

**TB diagnostic based on antigens from *M. tuberculosis***

This application is a continuation-in-part of:

- US Patent Application No. 09/050,739, filed 30 March 1998, which claims priority  
5 from US Provisional Application No. 60/044,624, filed 18 April 1997, US Provisional  
Application No. 60/070,488, filed 5 January 1998, and Danish Patent Applications  
Nos. DK 1997 00376, filed 2 April 1997, and DK 1997 01277, filed 10 November  
1997; and
- US Patent Application No. 10/138,473 filed 2 May 2002 which is a continuation-in  
10 part of
  - the above-mentioned US Patent Application No. 09/050,739;
  - US Patent Application No. 09/791,171, filed 20 February 2001, which is a  
divisional of the above mentioned US Patent Application No. 09/050,739, claiming  
the same priorities;
  - 15 and US Patent Application No. 09/415,884, filed 8 October 1999, which claims  
priority from US Provisional Application No. 60/116,673, filed 21 January 1999 and  
Danish Patent Application No. DK 1998 01281, filed 8 October 1998, US Provisional  
Application No. 60/044,624, filed 18 April 1997, US Provisional Application No.  
60/070,488, filed 5 January 1998, and Danish Patent Applications Nos. DK 1997  
20 00376, filed 2 April 1997, and DK 1997 01277, filed 10 November 1997.

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practice of this invention. Documents incorporated by reference into this text are not  
admitted to be prior art.

35 It is noted that in this disclosure and particularly in the claims, terms such as "comprises",  
"comprised", "comprising" and the like can have the meaning attributed to it in U.S. Patent  
law; e.g., they can mean "includes", "included", "including", and the like; and that terms  
such as "consisting essentially of" and "consists essentially of" have the meaning ascribed  
to them in U.S. Patent law, e.g., they allow for elements not explicitly recited, but exclude  
40 elements that are found in the prior art or that affect a basic or novel characteristic of the  
invention.

## FIELD OF THE INVENTION

The present invention relates to a number of immunologically active, novel polypeptide fragments derived from the *Mycobacterium tuberculosis*, vaccines and other immunologic compositions containing the fragments as immunogenic components, and methods of production and use of the polypeptides. The invention also relates to novel nucleic acid fragments derived from *M. tuberculosis* which are useful in the preparation of the polypeptide fragments of the invention or in the diagnosis of infection with *M. tuberculosis*. The invention further relates to certain fusion polypeptides, notably fusions between ESAT-6 and MPT59.

## BACKGROUND OF THE INVENTION

Human tuberculosis (hereinafter designated "TB") caused by *Mycobacterium tuberculosis* is a severe global health problem responsible for approximately 3 million deaths annually, according to the WHO. The worldwide incidence of new TB cases has been progressively falling for the last decade but the recent years has markedly changed this trend due to the advent of AIDS and the appearance of multidrug resistant strains of *M. tuberculosis*.

The only vaccine presently available for clinical use is BCG, a vaccine which efficacy remains a matter of controversy. BCG generally induces a high level of acquired resistance in animal models of TB, but several human trials in developing countries have failed to demonstrate significant protection. Notably, BCG is not approved by the FDA for use in the United States.

This makes the development of a new and improved vaccine against TB an urgent matter which has been given a very high priority by the WHO. Many attempts to define protective mycobacterial substances have been made, and from 1950 to 1970 several investigators reported an increased resistance

after experimental vaccination. However, the demonstration of a specific long-term protective immune response with the potency of BCG has not yet been achieved by administration of soluble proteins or cell wall fragments, although progress is currently being made by relying on polypeptides derived from short term-culture filtrate, cf. the discussion below.

Immunity to *M. tuberculosis* is characterized by three basic features; i) Living bacilli efficiently induces a protective immune response in contrast to killed preparations; ii) Specifically sensitized T lymphocytes mediate this protection; iii) The most important mediator molecule seems to be interferon gamma (INF- $\gamma$ ).

Short term-culture filtrate (ST-CF) is a complex mixture of proteins released from *M. tuberculosis* during the first few days of growth in a liquid medium (Andersen et al., 1991). Culture filtrates has been suggested to hold protective antigens recognized by the host in the first phase of TB infection (Andersen et al. 1991, Orme et al. 1993). Recent data from several laboratories have demonstrated that experimental subunit vaccines based on culture filtrate antigens can provide high levels of acquired resistance to TB (Pal and Horwitz, 1992; Roberts et al., 1995; Andersen, 1994; Lindblad et al., 1997). Culture filtrates are, however, complex protein mixtures and until now very limited information has been available on the molecules responsible for this protective immune response. In this regard, only two culture filtrate antigens have been described as involved in protective immunity, the low mass antigen ESAT-6 (Andersen et al., 1995 and EP-A-0 706 571) and the 31 kDa molecule Ag85B (EP-0 432 203). There is therefore a need for the identification of further antigens involved in the induction of protective immunity against TB in order to eventually produce an effective subunit vaccine.

## OBJECT OF THE INVENTION

It is an object of the invention to provide novel antigens which are effective as components in a subunit vaccine against TB or which are useful as components in diagnostic  
5 compositions for the detection of infection with mycobacteria, especially virulence-associated mycobacteria. The novel antigens may also be important drug targets.

## SUMMARY OF THE INVENTION

The present invention is *i.a.* based on the identification and  
10 characterization of a number of previously uncharacterized culture filtrate antigens from *M. tuberculosis*. In animal models of TB, T cells mediating immunity are focused predominantly to antigens in the regions 6-12 and 17-30 kDa of ST-CF. In the present invention 8 antigens in the low molecular  
15 weight region (CFP7, CFP7A, CFP7B, CFP8A, CFP8B, CFP9, CFP10A, and CFP11) and 18 antigens (CFP16, CFP17, CFP19, CFP19B, CFP20, CFP21, CFP22, CFP22A, CFP23, CFP23A, CFP23B, CFP25, CFP26, CFP27, CFP28, CFP29, CFP30A, and CFP30B) in the 17-30 kDa region have been identified. Of these, CFP19A and  
20 CFP23 have been selected because they exhibit relatively high homologies with CFP21 and CFP25, respectively, in so far that a nucleotide homology sequence search in the Sanger Database (cf. below) with the genes encoding CFP21 and CFP25, (*cfp25* and *cfp21* respectively), shows homology to two *M. tuberculo-*  
25 *sis* DNA sequences, *orf19A* and *orf23*. The two sequences, *orf19a* and *orf23*, encode to putative proteins CFP19A and CFP23 with the molecular weights of approx. 19 and 23 kDa respectively. The identity, at amino acid level, to CFP21 and CFP25 is 46% and 50%, respectively, for both proteins. CFP21  
30 and CFP25 have been shown to be dominant T-cell antigens, and it is therefore believed that CFP19A and CFP23 are possible new T-cell antigens.



Furthermore, a 50 kDa antigen (CFP50) has been isolated from culture filtrate and so has also an antigen (CWP32) isolated from the cell wall in the 30 kDa region.

The present invention is also based on the identification of a number of putative antigens from *M. tuberculosis* which are not present in *Mycobacterium bovis* BCG strains. The nucleotide sequences encoding these putative antigens are: rd1-orf2, rd1-orf3, rd1-orf4, rd1-orf5, rd1-orf8, rd1-orf9a, and rd1-orf9b.

Finally, the invention is based on the surprising discovery that fusions between ESAT-6 and MPT59 are superior immunogens compared to the unfused proteins, respectively.

The encoding genes for 33 of the antigens have been determined, the distribution of a number of the antigens in various mycobacterial strains investigated and the biological activity of the products characterized. The panel hold antigens with potential for vaccine purposes as well as for diagnostic purposes, since the antigens are all secreted by metabolizing mycobacteria.

The following table lists the antigens of the invention by the names used herein as well as by reference to relevant SEQ ID NOs of N-terminal sequences, full amino acid sequences and sequences of DNA encoding the antigens:

Antigen	N-terminal sequence SEQ ID NO:	Nucleotide sequence SEQ ID NO:	Amino acid sequence SEQ ID NO:
25 CFP7		1	2
CFP7A	81	47	48
CFP7B	168	146	147
CFP8A	73	148	149
CFP8B	74	150	151
30 CFP9		3	4
CFP10A	169	140	141
CFP11	170	142	143
CFP16	79	63	64
CFP17	17	5	6
35 CFP19	82	49	50
CFP19A		51	52

Antigen	N-terminal sequence SEQ ID NO:	Nucleotide sequence SEQ ID NO:	Amino acid sequence SEQ ID NO:
CFP19B	80		
CFP20	18	7	8
CFP21	19	9	10
CFP22	20	11	12
5 CFP22A	83	53	54
CFP23		55	56
CFP23A	76		
CFP23B	75		
CFP25	21	13	14
10 CFP25A	78	65	66
CFP27	84	57	58
CFP28	22		
CFP29	23	15	16
CFP30A	85	59	60
15 CFP30B	171	144	145
CFP50	86	61	62
MPT51		41	42
CWP32	77	152	153
RD1-ORF8		67	68
20 RD1-ORF2		71	72
RD1-ORF9B		69	70
RD1-ORF3		87	88
RD1-ORF9A		93	94
RD1-ORF4		89	90
25 RD1-ORF5		91	92
MPT59 -			172
ESAT6			
ESAT6 -			173
MPT59			

30 It is well-known in the art that T-cell epitopes are respon-  
sible for the elicitation of the acquired immunity against  
TB, whereas B-cell epitopes are without any significant  
influence on acquired immunity and recognition of mycobacte-  
ria *in vivo*. Since such T-cell epitopes are linear and are  
35 known to have a minimum length of 6 amino acid residues, the  
present invention is especially concerned with the identifi-  
cation and utilisation of such T-cell epitopes.

Hence, in its broadest aspect the invention relates to a  
substantially pure polypeptide fragment which

- 40 a) comprises an amino acid sequence selected from the  
sequences shown in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14,  
16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60,  
62, 64, 66, 68, 70, any one of 72-86, 88, 90, 92, 94;

141, 143, 145, 147, 149, 151, 153, and any one of 168-171,

- b) comprises a subsequence of the polypeptide fragment defined in a) which has a length of at least 6 amino acid residues, said subsequence being immunologically equivalent to the polypeptide defined in a) with respect to the ability of evoking a protective immune response against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex, or
- c) comprises an amino acid sequence having a sequence identity with the polypeptide defined in a) or the subsequence defined in b) of at least 70% and at the same time being immunologically equivalent to the polypeptide defined in a) with respect to the ability of evoking a protective immune response against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex,

with the proviso that

- i) the polypeptide fragment is in essentially pure form when consisting of the amino acid sequence 1-96 of SEQ ID NO: 2 or when consisting of the amino acid sequence 87-108 of SEQ ID NO: 4 fused to  $\beta$ -galactosidase,

ii) the degree of sequence identity in c) is at least 95% when the polypeptide comprises a homologue of a polypeptide

which has the amino acid sequence SEQ ID NO: 12 or a subsequence thereof as defined in b), and

iii) the polypeptide fragment contains a threonine residue corresponding to position 213 in SEQ ID NO: 42 when comprising an amino acid sequence of at least 6 amino acids in SEQ ID NO: 42.

Other parts of the invention pertains to the DNA fragments encoding a polypeptide with the above definition as well as to DNA fragments useful for determining the presence of DNA encoding such polypeptides.

#### DETAILED DISCLOSURE OF THE INVENTION

In the present specification and claims, the term "polypeptide fragment" denotes both short peptides with a length of at least two amino acid residues and at most 10 amino acid residues, oligopeptides (11-100 amino acid residues), and longer peptides (the usual interpretation of "polypeptide", i.e. more than 100 amino acid residues in length) as well as proteins (the functional entity comprising at least one peptide, oligopeptide, or polypeptide which may be chemically modified by being glycosylated, by being lipidated, or by comprising prosthetic groups). The definition of polypeptides also comprises native forms of peptides/proteins in mycobacteria as well as recombinant proteins or peptides in any type of expression vectors transforming any kind of host, and also chemically synthesized peptides.

In the present context the term "substantially pure polypeptide fragment" means a polypeptide preparation which contains at most 5% by weight of other polypeptide material with which it is natively associated (lower percentages of other polypeptide material are preferred, e.g. at most 4%, at most 3%, at most 2%, at most 1%, and at most ½%). It is preferred that the substantially pure polypeptide is at least 96% pure, i.e. that the polypeptide constitutes at least 96%

by weight of total polypeptide material present in the preparation, and higher percentages are preferred, such as at least 97%, at least 98%, at least 99%, at least 99,25%, at least 99,5%, and at least 99,75%. It is especially preferred  
5 that the polypeptide fragment is in "essentially pure form", i.e. that the polypeptide fragment is essentially free of any other antigen with which it is natively associated, i.e. free of any other antigen from bacteria belonging to the tuberculosis complex. This can be accomplished by preparing the  
10 polypeptide fragment by means of recombinant methods in a non-mycobacterial host cell as will be described in detail below, or by synthesizing the polypeptide fragment by the well-known methods of solid or liquid phase peptide synthesis, e.g. by the method described by Merrifield or variations thereof.  
15

The term "subsequence" when used in connection with a polypeptide of the invention having a SEQ ID NO selected from 2, 4, 6, 8, 10, 12, 14, 16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, any one of 72-86, 88, 90,  
20 92, 94, 141, 143, 145, 147, 149, 151, 153, and any one of 168-171 denotes any continuous stretch of at least 6 amino acid residues taken from the *M. tuberculosis* derived polypeptides in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68,  
25 70, any one of 72-86, 88, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, or any one of 168-171 and being immunological equivalent thereto with respect to the ability of conferring increased resistance to infections with bacteria belonging to the tuberculosis complex. Thus, included is also a  
30 polypeptide from different sources, such as other bacteria or even from eukaryotic cells.

When referring to an "immunologically equivalent" polypeptide is herein meant that the polypeptide, when formulated in a vaccine or a diagnostic agent (i.e. together with a pharmaceutically acceptable carrier or vehicle and optionally an  
35 adjuvant), will

I) confer, upon administration (either alone or as an immunologically active constituent together with other antigens), an acquired increased specific resistance in a mouse and/or in a guinea pig and/or in a primate such as a human being against infections with bacteria belonging to the tuberculosis complex which is at least 20% of the acquired increased resistance conferred by *Mycobacterium bovis* BCG and also at least 20% of the acquired increased resistance conferred by the parent polypeptide comprising SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, any one of 72-86, 88, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, or any one of 168-171 (said parent polypeptide having substantially the same relative location and pattern in a 2DE gel prepared as the 2DE gel shown in Fig. 6, cf. the examples), the acquired increased resistance being assessed by the observed reduction in mycobacterial counts from spleen, lung or other organ homogenates isolated from the mouse or guinea pig receiving a challenge infection with a virulent strain of *M. tuberculosis*, or, in a primate such as a human being, being assessed by determining the protection against development of clinical tuberculosis in a vaccinated group versus that observed in a control group receiving a placebo or BCG (preferably the increased resistance is higher and corresponds to at least 50% of the protective immune response elicited by *M. bovis* BCG, such as at least 60%, or even more preferred to at least 80% of the protective immune response elicited by *M. bovis* BCG, such as at least 90%; in some cases it is expected that the increased resistance will supersede that conferred by *M. bovis* BCG, and hence it is preferred that the resistance will be at least 100%, such as at least 110% of said increased resistance); and/or

II) elicit a diagnostically significant immune response in a mammal indicating previous or ongoing sensitization

with antigens derived from mycobacteria belonging to the tuberculosis complex; this diagnostically significant immune response can be in the form of a delayed type hypersensitivity reaction which can e.g. be determined by a skin test, or can be in the form of IFN- $\gamma$  release determined e.g. by an IFN- $\gamma$  assay as described in detail below. A diagnostically significant response in a skin test setup will be a reaction which gives rise to a skin reaction which is at least 5 mm in diameter and which is at least 65% (preferably at least 75% such as at the least 85%) of the skin reaction (assessed as the skin reaction diameter) elicited by the parent polypeptide comprising SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, any one of 72-86, 88, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, or any one of 168-171.

The ability of the polypeptide fragment to confer increased immunity may thus be assessed by measuring in an experimental animal, e.g. a mouse or a guinea pig, the reduction in mycobacterial counts from the spleen, lung or other organ homogenates isolated from the experimental animal which have received a challenge infection with a virulent strain of mycobacteria belonging to the tuberculosis complex after previously having been immunized with the polypeptide, as compared to the mycobacterial counts in a control group of experimental animals infected with the same virulent strain, which experimental animals have not previously been immunized against tuberculosis. The comparison of the mycobacterial counts may also be carried out with mycobacterial counts from a group of experimental animals receiving a challenge infection with the same virulent strain after having been immunized with *Mycobacterium bovis* BCG.

The mycobacterial counts in homogenates from the experimental animals immunized with a polypeptide fragment according to the present invention must at the most be 5 times the counts

in the mice or guinea pigs immunized with *Mycobacterium bovis* BCG, such as at the most 3 times the counts, and preferably at the most 2 times the counts.

A more relevant assessment of the ability of the polypeptide  
 5 fragment of the invention to confer increased resistance is to compare the incidence of clinical tuberculosis in two groups of individuals (e.g. humans or other primates) where one group receives a vaccine as described herein which contains an antigen of the invention and the other group  
 10 receives either a placebo or an other known TB vaccine (e.g. BCG). In such a setup, the antigen of the invention should give rise to a protective immunity which is significantly higher than the one provided by the administration of the placebo (as determined by statistical methods known to the  
 15 skilled artisan).

The "tuberculosis-complex" has its usual meaning, i.e. the complex of mycobacteria causing TB which are *Mycobacterium tuberculosis*, *Mycobacterium bovis*, *Mycobacterium bovis* BCG, and *Mycobacterium africanum*.

20 In the present context the term "metabolizing mycobacteria" means live mycobacteria that are multiplying logarithmically and releasing polypeptides into the culture medium wherein they are cultured.

The term "sequence identity" indicates a quantitative measure  
 25 of the degree of homology between two amino acid sequences or between two nucleotide sequences of equal length: The

sequence identity can be calculated as  $\frac{(N_{ref}-N_{dif})100}{N_{ref}}$ , wherein

$N_{dif}$  is the total number of non-identical residues in the two sequences when aligned and wherein  $N_{ref}$  is the number of  
 30 residues in one of the sequences. Hence, the DNA sequence AGTCAGTC will have a sequence identity of 75% with the sequence AATCAATC ( $N_{dif}=2$  and  $N_{ref}=8$ ).



The sequence identity is used here to illustrate the degree of identity between the amino acid sequence of a given polypeptide and the amino acid sequence shown in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, any one of 72-86, 88, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, or any one of 168-171. The amino acid sequence to be compared with the amino acid sequence shown in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, any one of 72-86, 88, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, or any one of 168-171 may be deduced from a DNA sequence, e.g. obtained by hybridization as defined below, or may be obtained by conventional amino acid sequencing methods. The sequence identity is preferably determined on the amino acid sequence of a mature polypeptide, i.e. without taking any leader sequence into consideration.

As appears from the above disclosure, polypeptides which are not identical to the polypeptides having SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, any one of 72-86, 88, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, or any one of 168-171 are embraced by the present invention. The invention allows for minor variations which do not have an adverse effect on immunogenicity compared to the parent sequences and which may give interesting and useful novel binding properties or biological functions and immunogenicities etc.

Each polypeptide fragment may thus be characterized by specific amino acid and nucleic acid sequences. It will be understood that such sequences include analogues and variants produced by recombinant methods wherein such nucleic acid and polypeptide sequences have been modified by substitution, insertion, addition and/or deletion of one or more nucleotides in said nucleic acid sequences to cause the substitution, insertion, addition or deletion of one or more amino acid residues in the recombinant polypeptide. When the term DNA is used in the following, it should be understood that for the

number of purposes where DNA can be substituted with RNA, the term DNA should be read to include RNA embodiments which will be apparent for the man skilled in the art. For the purposes of hybridization, PNA may be used instead of DNA, as PNA has  
5 been shown to exhibit a very dynamic hybridization profile (PNA is described in Nielsen P E et al., 1991, Science 254: 1497-1500).

In both immunodiagnostics and vaccine preparation, it is often possible and practical to prepare antigens from seg-  
10 ments of a known immunogenic protein or polypeptide. Certain epitopic regions may be used to produce responses similar to those produced by the entire antigenic polypeptide. Potential antigenic or immunogenic regions may be identified by any of a number of approaches, e.g., Jameson-Wolf or Kyte-Doolittle  
15 antigenicity analyses or Hopp and Woods (1981) hydrophobicity analysis (see, e.g., Jameson and Wolf, 1988; Kyte and Doolittle, 1982; or U.S. Patent No. 4,554,101). Hydrophobicity analysis assigns average hydrophilicity values to each amino acid residue from these values average hydrophilicities can  
20 be calculated and regions of greatest hydrophilicity determined. Using one or more of these methods, regions of predicted antigenicity may be derived from the amino acid sequence assigned to the polypeptides of the invention.

Alternatively, in order to identify relevant T-cell epitopes  
25 which are recognized during an immune response, it is also possible to use a "brute force" method: Since T-cell epitopes are linear, deletion mutants of polypeptides having SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, any one of 72-86, 88,  
30 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, or any one of 168-171 will, if constructed systematically, reveal what regions of the polypeptides are essential in immune recognition, e.g. by subjecting these deletion mutants to the IFN- $\gamma$  assay described herein. Another method utilises overlapping  
35 oligomers (preferably synthetic having a length of e.g. 20 amino acid residues) derived from polypeptides having SEQ ID

NO: 2, 4, 6, 8, 10, 12, 14, 16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, any one of 72-86, 88, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, or any one of 168-171. Some of these will give a positive response in the  
5 IFN- $\gamma$  assay whereas others will not.

In a preferred embodiment of the invention, the polypeptide fragment of the invention comprises an epitope for a T-helper cell.

Although the minimum length of a T-cell epitope has been  
10 shown to be at least 6 amino acids, it is normal that such epitopes are constituted of longer stretches of amino acids. Hence it is preferred that the polypeptide fragment of the invention has a length of at least 7 amino acid residues, such as at least 8, at least 9, at least 10, at least 12, at  
15 least 14, at least 16, at least 18, at least 20, at least 22, at least 24, and at least 30 amino acid residues.

As will appear from the examples, a number of the polypeptides of the invention are natively translation products which include a leader sequence (or other short peptide  
20 sequences), whereas the product which can be isolated from short-term culture filtrates from bacteria belonging to the tuberculosis complex are free of these sequences. Although it may in some applications be advantageous to produce these polypeptides recombinantly and in this connection facilitate  
25 export of the polypeptides from the host cell by including information encoding the leader sequence in the gene for the polypeptide, it is more often preferred to either substitute the leader sequence with one which has been shown to be superior in the host system for effecting export, or to  
30 totally omit the leader sequence (e.g. when producing the polypeptide by peptide synthesis. Hence, a preferred embodiment of the invention is a polypeptide which is free from amino acid residues -30 to -1 in SEQ ID NO: 6 and/or -32 to -1 in SEQ ID NO: 10 and/or -8 to -1 in SEQ ID NO: 12 and/or  
35 -32 to -1 in SEQ ID NO: 14 and/or -33 to -1 in SEQ ID NO: 42

and/or -38 to -1 in SEQ ID NO: 52 and/or -33 to -1 in SEQ ID NO: 56 and/or -56 to -1 in SEQ ID NO: 58 and/or -28 to -1 in SEQ ID NO: 151.

5 In another preferred embodiment, the polypeptide fragment of the invention is free from any signal sequence; this is especially interesting when the polypeptide fragment is produced synthetically but even when the polypeptide fragments are produced recombinantly it is normally acceptable that they are not exported by the host cell to the periplasm  
10 or the extracellular space; the polypeptide fragments can be recovered by traditional methods (cf. the discussion below) from the cytoplasm after disruption of the host cells, and if there is need for refolding of the polypeptide fragments, general refolding schemes can be employed, cf. e.g. the  
15 disclosure in WO 94/18227 where such a general applicable refolding method is described.

A suitable assay for the potential utility of a given polypeptide fragment derived from SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, any one of 17-23, 42, 48, 50, 52, 54, 56, 58, 60,  
20 62, 64, 66, 68, 70, any one of 72-86, 88, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, or any one of 168-171 is to assess the ability of the polypeptide fragment to effect IFN- $\gamma$  release from primed memory T-lymphocytes. Polypeptide fragments which have this capability are according to the  
25 invention especially interesting embodiments of the invention: It is contemplated that polypeptide fragments which stimulate T lymphocyte immune response shortly after the onset of the infection are important in the control of the mycobacteria causing the infection before the mycobacteria  
30 have succeeded in multiplying up to the number of bacteria that would have resulted in fulminant infection.

Thus, an important embodiment of the invention is a polypeptide fragment defined above which

- 1) induces a release of IFN- $\gamma$  from primed memory T-lymphocytes withdrawn from a mouse within 2 weeks of primary infection or within 4 days after the mouse has been re-challenge infected with mycobacteria belonging to the tuberculosis complex, the induction performed by the addition of the polypeptide to a suspension comprising about 200,000 spleen cells per ml, the addition of the polypeptide resulting in a concentration of 1-4  $\mu$ g polypeptide per ml suspension, the release of IFN- $\gamma$  being assessable by determination of IFN- $\gamma$  in supernatant harvested 2 days after the addition of the polypeptide to the suspension, and/or
- 2) induces a release of IFN- $\gamma$  of at least 1,500 pg/ml above background level from about 1,000,000 human PBMC (peripheral blood mononuclear cells) per ml isolated from TB patients in the first phase of infection, or from healthy BCG vaccinated donors, or from healthy contacts to TB patients, the induction being performed by the addition of the polypeptide to a suspension comprising the about 1,000,000 PBMC per ml, the addition of the polypeptide resulting in a concentration of 1-4  $\mu$ g polypeptide per ml suspension, the release of IFN- $\gamma$  being assessable by determination of IFN- $\gamma$  in supernatant harvested 2 days after the addition of the polypeptide to the suspension; and/or
- 3) induces an IFN- $\gamma$  release from bovine PBMC derived from animals previously sensitized with mycobacteria belonging to the tuberculosis complex, said release being at least two times the release observed from bovine PBMC derived from animals not previously sensitized with mycobacteria belonging to the tuberculosis complex.

Preferably, in alternatives 1 and 2, the release effected by the polypeptide fragment gives rise to at least 1,500 pg/ml IFN- $\gamma$  in the supernatant but higher concentrations are preferred, e.g. at least 2,000 pg/ml and even at least 3,000

pg/ml IFN- $\gamma$  in the supernatant. The IFN- $\gamma$  release from bovine PBMC can e.g. be measured as the optical density (OD) index over background in a standard cytokine ELISA and should thus be at least two, but higher numbers such as at least 3, 5, 8, 5 and 10 are preferred.

The polypeptide fragments of the invention preferably comprises an amino acid sequence of at least 6 amino acid residues in length which has a higher sequence identity than 70 percent with SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, any one 10 of 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, any one of 72-86, 88, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, or any one of 168-171. A preferred minimum percentage of sequence identity is at least 80%, such as at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, 15 at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, and at least 99.5%.

As mentioned above, it will normally be interesting to omit the leader sequences from the polypeptide fragments of the invention. However, by producing fusion polypeptides, 20 superior characteristics of the polypeptide fragments of the invention can be achieved. For instance, fusion partners which facilitate export of the polypeptide when produced recombinantly, fusion partners which facilitate purification of the polypeptide, and fusion partners which enhance the 25 immunogenicity of the polypeptide fragment of the invention are all interesting possibilities. Therefore, the invention also pertains to a fusion polypeptide comprising at least one polypeptide fragment defined above and at least one fusion partner. The fusion partner can, in order to enhance immuno- 30 genicity, e.g. be selected from the group consisting of another polypeptide fragment as defined above (so as to allow for multiple expression of relevant epitopes), and an other polypeptide derived from a bacterium belonging to the tuberculosis complex, such as ESAT-6, MPB64, MPT64, and MPB59 or 35 at least one T-cell epitope of any of these antigens. Other immunogenicity enhancing polypeptides which could serve as

fusion partners are T-cell epitopes (e.g. derived from the polypeptides ESAT-6, MPB64, MPT64, or MPB59) or other immunogenic epitopes enhancing the immunogenicity of the target gene product, e.g. lymphokines such as INF- $\gamma$ , IL-2 and  
5 IL-12. In order to facilitate expression and/or purification the fusion partner can e.g. be a bacterial fimbrial protein, e.g. the pilus components pilin and papA; protein A; the ZZ-peptide (ZZ-fusions are marketed by Pharmacia in Sweden); the maltose binding protein; glutathione S-transferase;  $\beta$ -galac-  
10 tosidase; or poly-histidine.

Other interesting fusion partners are polypeptides which are lipidated and thereby effect that the immunogenic polypeptide is presented in a suitable manner to the immune system. This effect is e.g. known from vaccines based on the *Borrelia*  
15 *burgdorferi* OspA polypeptide, wherein the lipidated membrane anchor in the polypeptide confers a self-adjuvating effect to the polypeptide (which is natively lipidated) when isolated from cells producing it. In contrast, the OspA polypeptide is relatively silent immunologically when prepared without the  
20 lipidation anchor.

As evidenced in Example 6A, the fusion polypeptide consisting of MPT59 fused directly N-terminally to ESAT-6 enhances the immunogenicity of ESAT-6 beyond what would be expected from the immunogenicities of MPT59 and ESAT-6 alone. The precise  
25 reason for this surprising finding is not yet known, but it is expected that either the presence of both antigens lead to a synergistic effect with respect to immunogenicity or the presence of a sequence N-terminally to the ESAT-6 sequence protects this immune dominant protein from loss of important  
30 epitopes known to be present in the N-terminus. A third, alternative, possibility is that the presence of a sequence C-terminally to the MPT59 sequence enhances the immunologic properties of this antigen.

Hence, one part of the invention pertains to a fusion  
35 polypeptide fragment which comprises a first amino acid

sequence including at least one stretch of amino acids constituting a T-cell epitope derived from the *M. tuberculosis* protein ESAT-6 or MPT59, and a second amino acid sequence including at least one T-cell epitope derived from a *M.*

5 *tuberculosis* protein different from ESAT-6 (if the first stretch of amino acids are derived from ESAT-6) or MPT59 (if the first stretch of amino acids are derived from MPT59) and/or including a stretch of amino acids which protects the first amino acid sequence from *in vivo* degradation or post-

10 translational processing. The first amino acid sequence may be situated N- or C-terminally to the second amino acid sequence, but in line with the above considerations regarding protection of the ESAT-6 N-terminus it is preferred that the first amino acid sequence is C-terminal to the second when

15 the first amino acid sequence is derived from ESAT-6.

Although only the effect of fusion between MPT59 and ESAT6 has been investigated at present, it is believed that ESAT6 and MPT59 or epitopes derived therefrom could be advantageously be fused to other fusion partners having substantially

20 the same effect on overall immunogenicity of the fusion construct. Hence, it is preferred that such a fusion polypeptide fragment according of the invention is one, wherein the at least one T-cell epitope included in the second amino acid sequence is derived from a *M. tuberculosis*

25 polypeptide (the "parent" polypeptide) selected from the group consisting of a polypeptide fragment according to the present invention and described in detail above and in the examples, or the amino acid sequence could be derived from any one of the *M. tuberculosis* proteins DnaK, GroEL, urease,

30 glutamine synthetase, the proline rich complex, L-alanine dehydrogenase, phosphate binding protein, Ag 85 complex, HBHA (heparin binding hemagglutinin), MPT51, MPT64, superoxide dismutase, 19 kDa lipoprotein,  $\alpha$ -crystallin, GroES, MPT59 (when the first amino acid sequence is derived from ESAT-6),

35 and ESAT-6 (when the first amino acid sequence is derived from MPT59). It is preferred that the first and second T-cell epitopes each have a sequence identity of at least 70% with



the natively occurring sequence in the proteins from which they are derived and it is even further preferred that the first and/or second amino acid sequence has a sequence identity of at least 70% with the protein from which they are  
 5 derived. A most preferred embodiment of this fusion polypeptide is one wherein the first amino acid sequence is the amino acid sequence of ESAT-6 or MPT59 and/or the second amino acid sequence is the full-length amino acid sequence of the possible "parent" polypeptides listed above.

10 In the most preferred embodiment, the fusion polypeptide fragment comprises ESAT-6 fused to MPT59 (advantageously, ESAT-6 is fused to the C-terminus of MPT59) and in one special embodiment, there are no linkers introduced between the two amino acid sequences constituting the two parent  
 15 polypeptide fragments.

Another part of the invention pertains to a nucleic acid fragment in isolated form which

- 1) comprises a nucleic acid sequence which encodes a polypeptide or fusion polypeptide as defined above, or  
 20 comprises a nucleic acid sequence complementary thereto, and/or
- 2) has a length of at least 10 nucleotides and hybridizes readily under stringent hybridization conditions (as defined in the art, i.e. 5-10°C under the melting point  
 25  $T_m$ , cf. Sambrook et al, 1989, pages 11.45-11.49) with a nucleic acid fragment which has a nucleotide sequence selected from  
 SEQ ID NO: 1 or a sequence complementary thereto,  
 SEQ ID NO: 3 or a sequence complementary thereto,  
 30 SEQ ID NO: 5 or a sequence complementary thereto,  
 SEQ ID NO: 7 or a sequence complementary thereto,  
 SEQ ID NO: 9 or a sequence complementary thereto,  
 SEQ ID NO: 11 or a sequence complementary thereto,  
 SEQ ID NO: 13 or a sequence complementary thereto,

SEQ ID NO: 15 or a sequence complementary thereto,  
 SEQ ID NO: 41 or a sequence complementary thereto,  
 SEQ ID NO: 47 or a sequence complementary thereto,  
 SEQ ID NO: 49 or a sequence complementary thereto,  
 5 SEQ ID NO: 51 or a sequence complementary thereto,  
 SEQ ID NO: 53 or a sequence complementary thereto,  
 SEQ ID NO: 55 or a sequence complementary thereto,  
 SEQ ID NO: 57 or a sequence complementary thereto,  
 SEQ ID NO: 59 or a sequence complementary thereto,  
 10 SEQ ID NO: 61 or a sequence complementary thereto,  
 SEQ ID NO: 63 or a sequence complementary thereto,  
 SEQ ID NO: 65 or a sequence complementary thereto,  
 SEQ ID NO: 67 or a sequence complementary thereto,  
 SEQ ID NO: 69 or a sequence complementary thereto,  
 15 SEQ ID NO: 71 or a sequence complementary thereto,  
 SEQ ID NO: 87 or a sequence complementary thereto,  
 SEQ ID NO: 89 or a sequence complementary thereto,  
 SEQ ID NO: 91 or a sequence complementary thereto,  
 SEQ ID NO: 93 or a sequence complementary thereto,  
 20 SEQ ID NO: 140 or a sequence complementary thereto,  
 SEQ ID NO: 142 or a sequence complementary thereto,  
 SEQ ID NO: 144 or a sequence complementary thereto,  
 SEQ ID NO: 146 or a sequence complementary thereto,  
 SEQ ID NO: 148 or a sequence complementary thereto,  
 25 SEQ ID NO: 150 or a sequence complementary thereto, and  
 SEQ ID NO: 152 or a sequence complementary thereto,

with the proviso that when the nucleic acid fragment comprises a subsequence of SEQ ID NO: 41, then the nucleic acid fragment contains an A corresponding to position 781 in SEQ  
 30 ID NO: 41 and when the nucleic acid fragment comprises a subsequence of a nucleotide sequence exactly complementary to SEQ ID NO: 41, then the nucleic acid fragment comprises a T corresponding to position 781 in SEQ ID NO: 41.

It is preferred that the nucleic acid fragment is a DNA  
 35 fragment.

To provide certainty of the advantages in accordance with the invention, the preferred nucleic acid sequence when employed for hybridization studies or assays includes sequences that are complementary to at least a 10 to 40, or so, nucleotide stretch of the selected sequence. A size of at least 10 nucleotides in length helps to ensure that the fragment will be of sufficient length to form a duplex molecule that is both stable and selective. Molecules having complementary sequences over stretches greater than 10 bases in length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained.

Hence, the term "subsequence" when used in connection with the nucleic acid fragments of the invention is intended to indicate a continuous stretch of at least 10 nucleotides exhibits the above hybridization pattern. Normally this will require a minimum sequence identity of at least 70% with a subsequence of the hybridization partner having SEQ ID NO: 1, 3, 5, 7, 9, 11, 12, 15, 21, 41, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 87, 89, 91, 93, 140, 142, 144, 146, 148, 150, or 152. It is preferred that the nucleic acid fragment is longer than 10 nucleotides, such as at least 15, at least 20, at least 25, at least 30, at least 35, at least 40, at least 45, at least 50, at least 55, at least 60, at least 65, at least 70, and at least 80 nucleotides long, and the sequence identity should preferable also be higher than 70%, such as at least 75%, at least 80%, at least 85%, at least 90%, at least 92%, at least 94%, at least 96%, and at least 98%. It is most preferred that the sequence identity is 100%. Such fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, by application of nucleic acid reproduction technology, such as the PCR technology of U.S. Patent 4,603,102, or by introducing selected sequences into recombinant vectors for recombinant production.

It is well known that the same amino acid may be encoded by various codons, the codon usage being related, *inter alia*, to the preference of the organisms in question expressing the nucleotide sequence. Thus, at least one nucleotide or codon  
5 of a nucleic acid fragment of the invention may be exchanged by others which, when expressed, result in a polypeptide identical or substantially identical to the polypeptide encoded by the nucleic acid fragment in question. The invention thus allows for variations in the sequence such as  
10 substitution, insertion (including introns), addition, deletion and rearrangement of one or more nucleotides, which variations do not have any substantial effect on the polypeptide encoded by the nucleic acid fragment or a subsequence thereof. The term "substitution" is intended to mean the  
15 replacement of one or more nucleotides in the full nucleotide sequence with one or more different nucleotides, "addition" is understood to mean the addition of one or more nucleotides at either end of the full nucleotide sequence, "insertion" is intended to mean the introduction of one or more nucleotides  
20 within the full nucleotide sequence, "deletion" is intended to indicate that one or more nucleotides have been deleted from the full nucleotide sequence whether at either end of the sequence or at any suitable point within it, and "rearrangement" is intended to mean that two or more nucleotide  
25 residues have been exchanged with each other.

The nucleotide sequence to be modified may be of cDNA or genomic origin as discussed above, but may also be of synthetic origin. Furthermore, the sequence may be of mixed cDNA and genomic, mixed cDNA and synthetic or genomic and synthetic origin as discussed above. The sequence may have been  
30 modified, e.g. by site-directed mutagenesis, to result in the desired nucleic acid fragment encoding the desired polypeptide. The following discussion focused on modifications of nucleic acid encoding the polypeptide should be  
35 understood to encompass also such possibilities, as well as the possibility of building up the nucleic acid by ligation of two or more DNA fragments to obtain the desired nucleic

acid fragment, and combinations of the above-mentioned principles.

The nucleotide sequence may be modified using any suitable technique which results in the production of a nucleic acid  
5 fragment encoding a polypeptide of the invention.

The modification of the nucleotide sequence encoding the amino acid sequence of the polypeptide of the invention should be one which does not impair the immunological function of the resulting polypeptide.

10 A preferred method of preparing variants of the antigens disclosed herein is site-directed mutagenesis. This technique is useful in the preparation of individual peptides, or biologically functional equivalent proteins or peptides, derived from the antigen sequences, through specific mutagenesis of the underlying nucleic acid. The technique further  
15 provides a ready ability to prepare and test sequence variants, for example, incorporating one or more of the foregoing considerations, by introducing one or more nucleotide sequence changes into the nucleic acid. Site-specific mutagenesis allows the production of mutants through the use of  
20 specific oligonucleotide sequences which encode the nucleotide sequence of the desired mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer sequence of sufficient size and sequence complexity to  
25 form a stable duplex on both sides of the deletion junction being traversed. Typically, a primer of about 17 to 25 nucleotides in length is preferred, with about 5 to 10 residues on both sides of the junction of the sequence being altered.

In general, the technique of site-specific mutagenesis is  
30 well known in the art as exemplified by publications (Adelman et al., 1983). As will be appreciated, the technique typically employs a phage vector which exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis include vectors such as the M13

phage (Messing et al., 1981). These phage are readily commercially available and their use is generally well known to those skilled in the art.

- In general, site-directed mutagenesis in accordance herewith
- 5 is performed by first obtaining a single-stranded vector which includes within its sequence a nucleic acid sequence which encodes the polypeptides of the invention. An oligonucleotide primer bearing the desired mutated sequence is prepared, generally synthetically, for example by the method
- 10 of Crea et al. (1978). This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as E. coli polymerase I Klenow fragment, in order to complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand
- 15 encodes the original non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform appropriate cells, such as E. coli cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement.
- 20 The preparation of sequence variants of the selected nucleic acid fragments of the invention using site-directed mutagenesis is provided as a means of producing potentially useful species of the genes and is not meant to be limiting as there are other ways in which sequence variants of the nucleic acid
- 25 fragments of the invention may be obtained. For example, recombinant vectors encoding the desired genes may be treated with mutagenic agents to obtain sequence variants (see, e.g., a method described by Eichenlaub, 1979) for the mutagenesis of plasmid DNA using hydroxylamine.
- 30 The invention also relates to a replicable expression vector which comprises a nucleic acid fragment defined above, especially a vector which comprises a nucleic acid fragment encoding a polypeptide fragment of the invention.

The vector may be any vector which may conveniently be subjected to recombinant DNA procedures, and the choice of vector will often depend on the host cell into which it is to be introduced. Thus, the vector may be an autonomously replicating vector, *i.e.* a vector which exists as an extrachromosomal entity, the replication of which is independent of chromosomal replication; examples of such a vector are a plasmid, phage, cosmid, mini-chromosome or virus. Alternatively, the vector may be one which, when introduced in a host cell, is integrated in the host cell genome and replicated together with the chromosome(s) into which it has been integrated.

Expression vectors may be constructed to include any of the DNA segments disclosed herein. Such DNA might encode an antigenic protein specific for virulent strains of mycobacteria or even hybridization probes for detecting mycobacteria nucleic acids in samples. Longer or shorter DNA segments could be used, depending on the antigenic protein desired. Epitopic regions of the proteins expressed or encoded by the disclosed DNA could be included as relatively short segments of DNA. A wide variety of expression vectors is possible including, for example, DNA segments encoding reporter gene products useful for identification of heterologous gene products and/or resistance genes such as antibiotic resistance genes which may be useful in identifying transformed cells.

The vector of the invention may be used to transform cells so as to allow propagation of the nucleic acid fragments of the invention or so as to allow expression of the polypeptide fragments of the invention. Hence, the invention also pertains to a transformed cell harbouring at least one such vector according to the invention, said cell being one which does not natively harbour the vector and/or the nucleic acid fragment of the invention contained therein. Such a transformed cell (which is also a part of the invention) may be any suitable bacterial host cell or any other type of cell

- such as a unicellular eukaryotic organism, a fungus or yeast, or a cell derived from a multicellular organism, e.g. an animal or a plant. It is especially in cases where glycosylation is desired that a mammalian cell is used,
- 5 although glycosylation of proteins is a rare event in prokaryotes. Normally, however, a prokaryotic cell is preferred such as a bacterium belonging to the genera *Mycobacterium*, *Salmonella*, *Pseudomonas*, *Bacillus* and *Eschericia*. It is preferred that the transformed cell is an *E. coli*, *B. subtilis*, or *M. bovis* BCG cell, and it is especially preferred
- 10 that the transformed cell expresses a polypeptide according of the invention. The latter opens for the possibility to produce the polypeptide of the invention by simply recovering it from the culture containing the transformed cell. In the
- 15 most preferred embodiment of this part of the invention the transformed cell is *Mycobacterium bovis* BCG strain: Danish 1331, which is the *Mycobacterium bovis* strain Copenhagen from the Copenhagen BCG Laboratory, Statens Seruminstitut, Denmark.
- 20 The nucleic acid fragments of the invention allow for the recombinant production of the polypeptides fragments of the invention. However, also isolation from the natural source is a way of providing the polypeptide fragments as is peptide synthesis.
- 25 Therefore, the invention also pertains to a method for the preparation of a polypeptide fragment of the invention, said method comprising inserting a nucleic acid fragment as defined above into a vector which is able to replicate in a host cell, introducing the resulting recombinant vector into
- 30 the host cell (transformed cells may be selected using various techniques, including screening by differential hybridization, identification of fused reporter gene products, resistance markers, anti-antigen antibodies and the like), culturing the host cell in a culture medium under conditions
- 35 sufficient to effect expression of the polypeptide (of course the cell may be cultivated under conditions appropriate to



the circumstances, and if DNA is desired, replication conditions are used), and recovering the polypeptide from the host cell or culture medium; or

isolating the polypeptide from a short-term culture filtrate  
5 as defined in claim 1; or

isolating the polypeptide from whole mycobacteria of the tuberculosis complex or from lysates or fractions thereof, e.g. cell wall containing fractions, or

synthesizing the polypeptide by solid or liquid phase peptide  
10 synthesis.

The medium used to grow the transformed cells may be any conventional medium suitable for the purpose. A suitable vector may be any of the vectors described above, and an appropriate host cell may be any of the cell types listed  
15 above. The methods employed to construct the vector and effect introduction thereof into the host cell may be any methods known for such purposes within the field of recombinant DNA. In the following a more detailed description of the possibilities will be given:

20 In general, of course, prokaryotes are preferred for the initial cloning of nucleic sequences of the invention and constructing the vectors useful in the invention. For example, in addition to the particular strains mentioned in the more specific disclosure below, one may mention by way of  
25 example, strains such as *E. coli* K12 strain 294 (ATCC No. 31446), *E. coli* B, and *E. coli* X 1776 (ATCC No. 31537). These examples are, of course, intended to be illustrative rather than limiting.

Prokaryotes are also preferred for expression. The  
30 aforementioned strains, as well as *E. coli* W3110 (F-, lambda-, prototrophic, ATCC No. 273325), bacilli such as *Bacillus subtilis*, or other enterobacteriaceae such as *Salmonella*

typhimurium or *Serratia marcesans*, and various *Pseudomonas* species may be used. Especially interesting are rapid-growing mycobacteria, e.g. *M. smegmatis*, as these bacteria have a high degree of resemblance with mycobacteria of the tuberculosis complex and therefore stand a good chance of reducing the need of performing post-translational modifications of the expression product.

In general, plasmid vectors containing replicon and control sequences which are derived from species compatible with the host cell are used in connection with these hosts. The vector ordinarily carries a replication site, as well as marking sequences which are capable of providing phenotypic selection in transformed cells. For example, *E. coli* is typically transformed using pBR322, a plasmid derived from an *E. coli* species (see, e.g., Bolivar et al., 1977, Gene 2: 95). The pBR322 plasmid contains genes for ampicillin and tetracycline resistance and thus provides easy means for identifying transformed cells. The pBR plasmid, or other microbial plasmid or phage must also contain, or be modified to contain, promoters which can be used by the microorganism for expression.

Those promoters most commonly used in recombinant DNA construction include the B-lactamase (penicillinase) and lactose promoter systems (Chang et al., 1978; Itakura et al., 1977; Goeddel et al., 1979) and a tryptophan (*trp*) promoter system (Goeddel et al., 1979; EPO Appl. Publ. No. 0036776). While these are the most commonly used, other microbial promoters have been discovered and utilized, and details concerning their nucleotide sequences have been published, enabling a skilled worker to ligate them functionally with plasmid vectors (Siebwenlist et al., 1980). Certain genes from prokaryotes may be expressed efficiently in *E. coli* from their own promoter sequences, precluding the need for addition of another promoter by artificial means.

After the recombinant preparation of the polypeptide according to the invention, the isolation of the polypeptide may for instance be carried out by affinity chromatography (or other conventional biochemical procedures based on chromatography), using a monoclonal antibody which substantially specifically binds the polypeptide according to the invention. Another possibility is to employ the simultaneous electroelution technique described by Andersen et al. in J. Immunol. Methods 161: 29-39.

- 10 According to the invention the post-translational modifications involves lipidation, glycosylation, cleavage, or elongation of the polypeptide.

In certain aspects, the DNA sequence information provided by this invention allows for the preparation of relatively short DNA (or RNA or PNA) sequences having the ability to specifically hybridize to mycobacterial gene sequences. In these aspects, nucleic acid probes of an appropriate length are prepared based on a consideration of the relevant sequence. The ability of such nucleic acid probes to specifically hybridize to the mycobacterial gene sequences lend them particular utility in a variety of embodiments. Most importantly, the probes can be used in a variety of diagnostic assays for detecting the presence of pathogenic organisms in a given sample. However, either uses are envisioned, including the use of the sequence information for the preparation of mutant species primers, or primers for use in preparing other genetic constructs.

Apart from their use as starting points for the synthesis of polypeptides of the invention and for hybridization probes (useful for direct hybridization assays or as primers in e.g. PCR or other molecular amplification methods) the nucleic acid fragments of the invention may be used for effecting *in vivo* expression of antigens, *i.e.* the nucleic acid fragments may be used in so-called DNA vaccines. Recent research have revealed that a DNA fragment cloned in a vector which is non-

replicative in eukaryotic cells may be introduced into an animal (including a human being) by e.g. intramuscular injection or percutaneous administration (the so-called "gene gun" approach). The DNA is taken up by e.g. muscle cells and the  
5 gene of interest is expressed by a promoter which is functioning in eukaryotes, e.g. a viral promoter, and the gene product thereafter stimulates the immune system. These newly discovered methods are reviewed in Ulmer et al., 1993, which hereby is included by reference.

10 Hence, the invention also relates to a vaccine comprising a nucleic acid fragment according to the invention, the vaccine effecting *in vivo* expression of antigen by an animal, including a human being, to whom the vaccine has been administered, the amount of expressed antigen being effective to  
15 confer substantially increased resistance to infections with mycobacteria of the tuberculosis complex in an animal, including a human being.

The efficacy of such a "DNA vaccine" can possibly be enhanced by administering the gene encoding the expression product  
20 together with a DNA fragment encoding a polypeptide which has the capability of modulating an immune response. For instance, a gene encoding lymphokine precursors or lymphokines (e.g. IFN- $\gamma$ , IL-2, or IL-12) could be administered together with the gene encoding the immunogenic protein,  
25 either by administering two separate DNA fragments or by administering both DNA fragments included in the same vector. It also is a possibility to administer DNA fragments comprising a multitude of nucleotide sequences which each encode relevant epitopes of the polypeptides disclosed herein so as  
30 to effect a continuous sensitization of the immune system with a broad spectrum of these epitopes.

As explained above, the polypeptide fragments of the invention are excellent candidates for vaccine constituents or for constituents in an immune diagnostic agent due to their  
35 extracellular presence in culture media containing metaboli-

zing virulent mycobacteria belonging to the tuberculosis complex, or because of their high homologies with such extra-cellular antigens, or because of their absence in *M. bovis* BCG.

- 5 Thus, another part of the invention pertains to an immunologic composition comprising a polypeptide or fusion polypeptide according to the invention. In order to ensure optimum performance of such an immunologic composition it is preferred that it comprises an immunologically and pharma-  
10 ceutically acceptable carrier, vehicle or adjuvant.

Suitable carriers are selected from the group consisting of a polymer to which the polypeptide(s) is/are bound by hydrophobic non-covalent interaction, such as a plastic, e.g. polystyrene, or a polymer to which the polypeptide(s) is/are  
15 covalently bound, such as a polysaccharide, or a polypeptide, e.g. bovine serum albumin, ovalbumin or keyhole limpet haemocyanin. Suitable vehicles are selected from the group consisting of a diluent and a suspending agent. The adjuvant is preferably selected from the group consisting of dimethyl-  
20 dioctadecylammonium bromide (DDA), Quil A, poly I:C, Freund's incomplete adjuvant, IFN- $\gamma$ , IL-2, IL-12, monophosphoryl lipid A (MPL), and muramyl dipeptide (MDP).

A preferred immunologic composition according to the present invention comprising at least two different polypeptide  
25 fragments, each different polypeptide fragment being a polypeptide or a fusion polypeptide defined above. It is preferred that the immunologic composition comprises between 3-20 different polypeptide fragments or fusion polypeptides.

Such an immunologic composition may preferably be in the form  
30 of a vaccine or in the form of a skin test reagent.

In line with the above, the invention therefore also pertain to a method for producing an immunologic composition according to the invention, the method comprising preparing,

synthesizing or isolating a polypeptide according to the invention, and solubilizing or dispersing the polypeptide in a medium for a vaccine, and optionally adding other *M. tuberculosis* antigens and/or a carrier, vehicle and/or adjuvant substance.

Preparation of vaccines which contain peptide sequences as active ingredients is generally well understood in the art, as exemplified by U.S. Patents 4,608,251; 4,601,903; 4,599,231; 4,599,230; 4,596,792; and 4,578,770, all incorporated herein by reference. Typically, such vaccines are prepared as injectables either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection may also be prepared. The preparation may also be emulsified. The active immunogenic ingredient is often mixed with excipients which are pharmaceutically acceptable and compatible with the active ingredient. Suitable excipients are, for example, water, saline, dextrose, glycerol, ethanol, or the like, and combinations thereof. In addition, if desired, the vaccine may contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, or adjuvants which enhance the effectiveness of the vaccines.

The vaccines are conventionally administered parenterally, by injection, for example, either subcutaneously or intramuscularly. Additional formulations which are suitable for other modes of administration include suppositories and, in some cases, oral formulations. For suppositories, traditional binders and carriers may include, for example, polyalkalene glycols or triglycerides; such suppositories may be formed from mixtures containing the active ingredient in the range of 0.5% to 10%, preferably 1-2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, and the like. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained release

formulations or powders and contain 10-95% of active ingredient, preferably 25-70%.

The proteins may be formulated into the vaccine as neutral or salt forms. Pharmaceutically acceptable salts include acid addition salts (formed with the free amino groups of the peptide) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups may also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, 2-ethylamino ethanol, histidine, procaine, and the like.

The vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be therapeutically effective and immunogenic. The quantity to be administered depends on the subject to be treated, including, e.g., the capacity of the individual's immune system to mount an immune response, and the degree of protection desired. Suitable dosage ranges are of the order of several hundred micrograms active ingredient per vaccination with a preferred range from about 0.1  $\mu\text{g}$  to 1000  $\mu\text{g}$ , such as in the range from about 1  $\mu\text{g}$  to 300  $\mu\text{g}$ , and especially in the range from about 10  $\mu\text{g}$  to 50  $\mu\text{g}$ . Suitable regimens for initial administration and booster shots are also variable but are typified by an initial administration followed by subsequent inoculations or other administrations.

The manner of application may be varied widely. Any of the conventional methods for administration of a vaccine are applicable. These are believed to include oral application on a solid physiologically acceptable base or in a physiologically acceptable dispersion, parenterally, by injection or the like. The dosage of the vaccine will depend on the route of administration and will vary according to the age of the

person to be vaccinated and, to a lesser degree, the size of the person to be vaccinated.

Some of the polypeptides of the vaccine are sufficiently immunogenic in a vaccine, but for some of the others the  
5 immune response will be enhanced if the vaccine further comprises an adjuvant substance.

Various methods of achieving adjuvant effect for the vaccine include use of agents such as aluminum hydroxide or phosphate (alum), commonly used as 0.05 to 0.1 percent solution in  
10 phosphate buffered saline, admixture with synthetic polymers of sugars (Carbopol) used as 0.25 percent solution, aggregation of the protein in the vaccine by heat treatment with temperatures ranging between 70° to 101°C for 30 second to 2 minute periods respectively. Aggregation by reactivating with  
15 pepsin treated (Fab) antibodies to albumin, mixture with bacterial cells such as *C. parvum* or endotoxins or lipopolysaccharide components of gram-negative bacteria, emulsion in physiologically acceptable oil vehicles such as mannide monooleate (Aracel A) or emulsion with 20 percent solution of a  
20 perfluorocarbon (Fluosol-DA) used as a block substitute may also be employed. According to the invention DDA (dimethyldioctadecylammonium bromide) is an interesting candidate for an adjuvant, but also Freund's complete and incomplete adjuvants as well as QuilA and RIBI are interesting possibilities.  
25 Further possibilities are monophosphoryl lipid A (MPL), and muramyl dipeptide (MDP).

Another highly interesting (and thus, preferred) possibility of achieving adjuvant effect is to employ the technique described in Gosselin et al., 1992 (which is hereby incorporated by reference herein). In brief, the presentation of a  
30 relevant antigen such as an antigen of the present invention can be enhanced by conjugating the antigen to antibodies (or antigen binding antibody fragments) against the Fcγ receptors on monocytes/macrophages. Especially conjugates between



antigen and anti-Fc $\gamma$ RI have been demonstrated to enhance immunogenicity for the purposes of vaccination.

Other possibilities involve the use of immune modulating substances such as lymphokines (e.g. IFN- $\gamma$ , IL-2 and IL-12) or synthetic IFN- $\gamma$  inducers such as poly I:C in combination with the above-mentioned adjuvants. As discussed in example 3, it is contemplated that such mixtures of antigen and adjuvant will lead to superior vaccine formulations.

In many instances, it will be necessary to have multiple administrations of the vaccine, usually not exceeding six vaccinations, more usually not exceeding four vaccinations and preferably one or more, usually at least about three vaccinations. The vaccinations will normally be at from two to twelve week intervals, more usually from three to five week intervals. Periodic boosters at intervals of 1-5 years, usually three years, will be desirable to maintain the desired levels of protective immunity. The course of the immunization may be followed by *in vitro* proliferation assays of PBL (peripheral blood lymphocytes) co-cultured with ESAT-6 or ST-CF, and especially by measuring the levels of IFN- $\gamma$  released from the primed lymphocytes. The assays may be performed using conventional labels, such as radionuclides, enzymes, fluorescers, and the like. These techniques are well known and may be found in a wide variety of patents, such as U.S. Patent Nos. 3,791,932; 4,174,384 and 3,949,064, as illustrative of these types of assays.

Due to genetic variation, different individuals may react with immune responses of varying strength to the same polypeptide. Therefore, the vaccine according to the invention may comprise several different polypeptides in order to increase the immune response. The vaccine may comprise two or more polypeptides, where all of the polypeptides are as defined above, or some but not all of the peptides may be derived from a bacterium belonging to the *M. tuberculosis* complex. In the latter example the polypeptides not necessa-

rily fulfilling the criteria set forth above for polypeptides may either act due to their own immunogenicity or merely act as adjuvants. Examples of such interesting polypeptides are MPB64, MPT64, and MPB59, but any other substance which can be  
5 isolated from mycobacteria are possible candidates.

The vaccine may comprise 3-20 different polypeptides, such as 3-10 different polypeptides.

One reason for admixing the polypeptides of the invention with an adjuvant is to effectively activate a cellular immune  
10 response. However, this effect can also be achieved in other ways, for instance by expressing the effective antigen in a vaccine in a non-pathogenic microorganism. A well-known example of such a microorganism is *Mycobacterium bovis* BCG.

Therefore, another important aspect of the present invention  
15 is an improvement of the living BCG vaccine presently available, which is a vaccine for immunizing an animal, including a human being, against TB caused by mycobacteria belonging to the tuberculosis-complex, comprising as the effective component a microorganism, wherein one or more copies of a DNA  
20 sequence encoding a polypeptide as defined above has been incorporated into the genome of the microorganism in a manner allowing the microorganism to express and secrete the polypeptide.

In the present context the term "genome" refers to the chromosome of the microorganisms as well as extrachromosomally  
25 DNA or RNA, such as plasmids. It is, however, preferred that the DNA sequence of the present invention has been introduced into the chromosome of the non-pathogenic microorganism, since this will prevent loss of the genetic material introduced.  
30

It is preferred that the non-pathogenic microorganism is a bacterium, e.g. selected from the group consisting of the genera *Mycobacterium*, *Salmonella*, *Pseudomonas* and *Eschericia*.

It is especially preferred that the non-pathogenic microorganism is *Mycobacterium bovis* BCG, such as *Mycobacterium bovis* BCG strain: Danish 1331.

- The incorporation of one or more copies of a nucleotide sequence encoding the polypeptide according to the invention in a mycobacterium from a *M. bovis* BCG strain will enhance the immunogenic effect of the BCG strain. The incorporation of more than one copy of a nucleotide sequence of the invention is contemplated to enhance the immune response even more, and consequently an aspect of the invention is a vaccine wherein at least 2 copies of a DNA sequence encoding a polypeptide is incorporated in the genome of the microorganism, such as at least 5 copies. The copies of DNA sequences may either be identical encoding identical polypeptides or be variants of the same DNA sequence encoding identical or homologues of a polypeptide, or in another embodiment be different DNA sequences encoding different polypeptides where at least one of the polypeptides is according to the present invention.
- 20 The living vaccine of the invention can be prepared by cultivating a transformed non-pathogenic cell according to the invention, and transferring these cells to a medium for a vaccine, and optionally adding a carrier, vehicle and/or adjuvant substance.
- 25 The invention also relates to a method of diagnosing TB caused by *Mycobacterium tuberculosis*, *Mycobacterium africanum* or *Mycobacterium bovis* in an animal, including a human being, comprising intradermally injecting, in the animal, a polypeptide according to the invention or a skin test reagent described above, a positive skin response at the location of injection being indicative of the animal having TB, and a negative skin response at the location of injection being indicative of the animal not having TB. A positive response is a skin reaction having a diameter of at least 5 mm, but larger reactions are preferred, such as at least 1 cm, 1.5

cm, and at least 2 cm in diameter. The composition used as the skin test reagent can be prepared in the same manner as described for the vaccines above.

- In line with the disclosure above pertaining to vaccine
- 5 preparation and use, the invention also pertains to a method for immunising an animal, including a human being, against TB caused by mycobacteria belonging to the tuberculosis complex, comprising administering to the animal the polypeptide of the invention, or a vaccine composition of the invention as
- 10 described above, or a living vaccine described above. Preferred routes of administration are the parenteral (such as intravenous and intraarterially), intraperitoneal, intramuscular, subcutaneous, intradermal, oral, buccal, sublingual, nasal, rectal or transdermal route.
- 15 The protein ESAT-6 which is present in short-term culture filtrates from mycobacteria as well as the *esat-6* gene in the mycobacterial genome has been demonstrated to have a very limited distribution in other mycobacterial strains that *M. tuberculosis*, e.g. *esat-6* is absent in both BCG and the
- 20 majority of mycobacterial species isolated from the environment, such as *M. avium* and *M. terrae*. It is believed that this is also the case for at least one of the antigens of the present invention and their genes and therefore, the diagnostic embodiments of the invention are especially well-suited
- 25 for performing the diagnosis of on-going or previous infection with virulent mycobacterial strains of the tuberculosis complex, and it is contemplated that it will be possible to distinguish between 1) subjects (animal or human) which have been previously vaccinated with e.g. BCG vaccines or sub-
- 30 jected to antigens from non-virulent mycobacteria and 2) subjects which have or have had active infection with virulent mycobacteria.

A number of possible diagnostic assays and methods can be envisaged:

When diagnosis of previous or ongoing infection with virulent mycobacteria is the aim, a blood sample comprising mononuclear cells (*i.a.* T-lymphocytes) from a patient could be contacted with a sample of one or more polypeptides of the invention. This contacting can be performed *in vitro* and a positive reaction could e.g. be proliferation of the T-cells or release cytokines such as  $\gamma$ -interferon into the extracellular phase (e.g. into a culture supernatant); a suitable *in vivo* test would be a skin test as described above. It is also conceivable to contact a serum sample from a subject to contact with a polypeptide of the invention, the demonstration of a binding between antibodies in the serum sample and the polypeptide being indicative of previous or ongoing infection.

The invention therefore also relates to an *in vitro* method for diagnosing ongoing or previous sensitization in an animal or a human being with bacteria belonging to the tuberculosis complex, the method comprising providing a blood sample from the animal or human being, and contacting the sample from the animal with the polypeptide of the invention, a significant release into the extracellular phase of at least one cytokine by mononuclear cells in the blood sample being indicative of the animal being sensitized. By the term "significant release" is herein meant that the release of the cytokine is significantly higher than the cytokine release from a blood sample derived from a non-tuberculous subject (e.g. a subject which does not react in a traditional skin test for TB). Normally, a significant release is at least two times the release observed from such a sample.

Alternatively, a sample of a possibly infected organ may be contacted with an antibody raised against a polypeptide of the invention. The demonstration of the reaction by means of methods well-known in the art between the sample and the antibody will be indicative of ongoing infection. It is of course also a possibility to demonstrate the presence of anti-mycobacterial antibodies in serum by contacting a serum

sample from a subject with at least one of the polypeptide fragments of the invention and using well-known methods for visualizing the reaction between the antibody and antigen.

Also a method of determining the presence of mycobacterial  
5 nucleic acids in an animal, including a human being, or in a sample, comprising administering a nucleic acid fragment of the invention to the animal or incubating the sample with the nucleic acid fragment of the invention or a nucleic acid fragment complementary thereto, and detecting the presence of  
10 hybridized nucleic acids resulting from the incubation (by using the hybridization assays which are well-known in the art), is also included in the invention. Such a method of diagnosing TB might involve the use of a composition comprising at least a part of a nucleotide sequence as defined  
15 above and detecting the presence of nucleotide sequences in a sample from the animal or human being to be tested which hybridize with the nucleic acid fragment (or a complementary fragment) by the use of PCR technique.

The fact that certain of the disclosed antigens are not  
20 present in *M. bovis* BCG but are present in virulent mycobacteria point them out as interesting drug targets; the antigens may constitute receptor molecules or toxins which facilitate the infection by the mycobacterium, and if such functionalities are blocked the infectivity of the mycobacte-  
25 rium will be diminished.

To determine particularly suitable drug targets among the antigens of the invention, the gene encoding at least one of the polypeptides of the invention and the necessary control sequences can be introduced into avirulent strains of myco-  
30 bacteria (e.g. BCG) so as to determine which of the polypeptides are critical for virulence. Once particular proteins are identified as critical for/contributory to virulence, anti-mycobacterial agents can be designed rationally to inhibit expression of the critical genes or to  
35 attack the critical gene products. For instance, antibodies

or fragments thereof (such as Fab and (Fab')<sub>2</sub> fragments can be prepared against such critical polypeptides by methods known in the art and thereafter used as prophylactic or therapeutic agents. Alternatively, small molecules can be  
5 screened for their ability to selectively inhibit expression of the critical gene products, e.g. using recombinant expression systems which include the gene's endogenous promoter, or for their ability to directly interfere with the action of the target. These small molecules are then used as thera-  
10 peutics or as prophylactic agents to inhibit mycobacterial virulence.

Alternatively, anti-mycobacterial agents which render a virulent mycobacterium avirulent can be operably linked to expression control sequences and used to transform a virulent  
15 mycobacterium. Such anti-mycobacterial agents inhibit the replication of a specified mycobacterium upon transcription or translation of the agent in the mycobacterium. Such a "newly avirulent" mycobacterium would constitute a superb alternative to the above described modified BCG for vaccine  
20 purposes since it would be immunologically very similar to a virulent mycobacterium compared to e.g. BCG.

Finally, a monoclonal or polyclonal antibody, which is specifically reacting with a polypeptide of the invention in an immuno assay, or a specific binding fragment of said anti-  
25 body, is also a part of the invention. The production of such polyclonal antibodies requires that a suitable animal be immunized with the polypeptide and that these antibodies are subsequently isolated, suitably by immune affinity chromatography. The production of monoclonals can be effected by  
30 methods well-known in the art, since the present invention provides for adequate amounts of antigen for both immunization and screening of positive hybridomas.

## LEGENDS TO THE FIGURES

Fig. 1: Long term memory immune mice are very efficiently protected towards an infection with *M. tuberculosis*. Mice were given a challenge of *M. tuberculosis* and spleens were isolated at different time points. Spleen lymphocytes were stimulated *in vitro* with ST-CF and the release of IFN- $\gamma$  investigated (panel A). The counts of CFU in the spleens of the two groups of mice are indicated in panel B. The memory immune mice control infection within the first week and produce large quantities of IFN- $\gamma$  in response to antigens in ST-CF.

Fig. 2: T cells involved in protective immunity are predominantly directed to molecules from 6-12 and 17-38 kDa. Splenic T cells were isolated four days after the challenge with *M. tuberculosis* and stimulated *in vitro* with narrow molecular mass fractions of ST-CF. The release of IFN- $\gamma$  was investigated

Fig. 3: Nucleotide sequence (SEQ ID NO: 1) of *cfp7*. The deduced amino acid sequence (SEQ ID NO: 2) of CFP7 is given in conventional one-letter code below the nucleotide sequence. The putative ribosome-binding site is written in underlined italics as are the putative -10 and -35 regions. Nucleotides written in bold are those encoding CFP7.

Fig. 4. Nucleotide sequence (SEQ ID NO: 3) of *cfp9*. The deduced amino acid sequence (SEQ ID NO: 4) of CFP9 is given in conventional one-letter code below the nucleotide sequence. The putative ribosome-binding site Shine Delgarno sequence is written in underlined italics as are the putative -10 and -35 regions. Nucleotides in bold writing are those encoding CFP9. The nucleotide sequence obtained from the lambda 226 phage is double underlined.

Fig. 5: Nucleotide sequence of *mpt51*. The deduced amino acid sequence of MPT51 is given in a one-letter code below the



nucleotide sequence. The signal is indicated in italics. the putative potential ribosome-binding site is underlined. The nucleotide difference and amino acid difference compared to the nucleotide sequence of MPB51 (Ohara et al., 1995) are  
 5 underlined at position 780. The nucleotides given in italics are not present in *M. tuberculosis* H37Rv.

Fig. 6: the position of the purified antigens in the 2DE system have been determined and mapped in a reference gel. The newly purified antigens are encircled and the position of  
 10 well-known proteins are also indicated.

#### EXAMPLE 1

##### *Identification of single culture filtrate antigens involved in protective immunity*

A group of efficiently protected mice was generated by infecting 8-12 weeks old female C57Bl/6j mice with  $5 \times 10^4$  *M. tuberculosis* i.v. After 30 days of infection the mice were subjected to 60 days of antibiotic treatment with isoniazid and were then left for 200-240 days to ensure the establishment of resting long-term memory immunity. Such memory immune  
 20 mice are very efficiently protected against a secondary infection (Fig. 1). Long lasting immunity in this model is mediated by a population of highly reactive CD4 cells recruited to the site of infection and triggered to produce large amounts of IFN- $\gamma$  in response to ST-CF (Fig. 1)  
 25 (Andersen et al. 1995).

We have used this model to identify single antigens recognized by protective T cells. Memory immune mice were reinfected with  $1 \times 10^6$  *M. tuberculosis* i.v. and splenic lymphocytes were harvested at day 4-6 of reinfection, a time  
 30 point where this population is highly reactive to ST-CF. The antigens recognized by these T cells were mapped by the multi-elution technique (Andersen and Heron, 1993). This technique divides complex protein mixtures separated in SDS-

PAGE into narrow fractions in a physiological buffer. These fractions were used to stimulate spleen lymphocytes *in vitro* and the release of IFN- $\gamma$  was monitored (Fig. 2). Long-term memory immune mice did not recognize these fractions before  
 5 TB infection, but splenic lymphocytes obtained during the recall of protective immunity recognized a range of culture filtrate antigens and peak production of IFN- $\gamma$  was found in response to proteins of apparent molecular weight 6-12 and 17-30 kDa (Fig. 2). It is therefore concluded that culture  
 10 filtrate antigens within these regions are the major targets recognized by memory effector T-cells triggered to release IFN- $\gamma$  during the first phase of a protective immune response.

## EXAMPLE 2

### 15 *Cloning of genes expressing low mass culture filtrate antigens*

In example 1 it was demonstrated that antigens in the low molecular mass fraction are recognized strongly by cells isolated from memory immune mice. Monoclonal antibodies (mAbs) to these antigens were therefore generated by immuni-  
 20 zing with the low mass fraction in RIBI adjuvant (first and second immunization) followed by two injections with the fractions in aluminium hydroxide. Fusion and cloning of the reactive cell lines were done according to standard procedures (Kohler and Milstein 1975). The procedure resulted in  
 25 the provision of two mAbs: ST-3 directed to a 9 kDa culture filtrate antigen (CFP9) and PV-2 directed to a 7 kDa antigen (CFP7), when the molecular weight is estimated from migration of the antigens in an SDS-PAGE.

In order to identify the antigens binding to the Mab's, the  
 30 following experiments were carried out:

The recombinant  $\lambda$ gt11 *M. tuberculosis* DNA library constructed by R. Young (Young, R.A. et al. 1985) and obtained through the World Health Organization IMMTUB programme

(WHO.0032.wibr) was screened for phages expressing gene products which would bind the monoclonal antibodies ST-3 and PV-2.

Approximately  $1 \times 10^5$  pfu of the gene library (containing  
5 approximately 25% recombinant phages) were plated on *Escher-*  
*icia coli* Y1090 (DlacU169, proA<sup>+</sup>, Dlon, araD139, supF,  
trpC22::tn10 [pMC9] ATCC#37197) in soft agar and incubated  
for 2,5 hours at 42°C.

The plates were overlaid with sheets of nitrocellulose satu-  
10 rated with isopropyl- $\beta$ -D-thiogalactopyranoside and incubation  
was continued for 2,5 hours at 37°C. The nitrocellulose was  
removed and incubated with samples of the monoclonal anti-  
bodies in PBS with Tween 20 added to a final concentration of  
0.05%. Bound monoclonal antibodies were visualized by horse-  
15 radish peroxidase-conjugated rabbit anti-mouse immunoglobu-  
lins (P260, Dako, Glostrup, DK) and a staining reaction  
involving 5,5',3,3'-tetramethylbenzidine and H<sub>2</sub>O<sub>2</sub>.

Positive plaques were recloned and the phages originating  
from a single plaque were used to lysogenize *E. coli* Y1089  
20 (DlacU169, proA<sup>+</sup>, Dlon, araD139, strA, hfl150 [chr::tn10]  
[pMC9] ATCC nr. 37196). The resultant lysogenic strains were  
used to propagate phage particles for DNA extraction. These  
lysogenic *E. coli* strains have been named:

AA226 (expressing ST-3 reactive polypeptide CFP9) which has  
25 been deposited 28 June 1993 with the collection of Deutsche  
Sammlung von Mikroorganismen und Zellkulturen GmbH (DSM)  
under the accession number DSM 8377 and in accordance with  
the provisions of the Budapest Treaty, and

AA242 (expressing PV-2 reactive polypeptide CFP7) which has  
30 been deposited 28 June 1993 with the collection of Deutsche  
Sammlung von Mikroorganismen und Zellkulturen GmbH (DSM)  
under the accession number DSM 8379 and in accordance with  
the provisions of the Budapest Treaty.

These two lysogenic *E. coli* strains are disclosed in WO 95/01441 as are the mycobacterial polypeptide products expressed thereby. However, no information concerning the amino acid sequences of these polypeptides or their genetic origin are given, and therefore only the direct expression products of AA226 and AA242 are made available to the public.

The st-3 binding protein is expressed as a protein fused to  $\beta$ -galactosidase, whereas the pv-2 binding protein appears to be expressed in an unfused version.

10 Sequencing of the nucleotide sequence encoding the PV-2 and ST-3 binding protein

In order to obtain the nucleotide sequence of the gene encoding the pv-2 binding protein, the approximately 3 kb *M. tuberculosis* derived *EcoRI* - *EcoRI* fragment from AA242 was subcloned in the *EcoRI* site in the pBluescriptSK + (Stratagene) and used to transform *E. coli* XL-1Blue (Stratagene).

Similarly, to obtain the nucleotide sequence of the gene encoding the st-3 binding protein, the approximately 5 kb *M. tuberculosis* derived *EcoRI* - *EcoRI* fragment from AA226 was subcloned in the *EcoRI* site in the pBluescriptSK + (Stratagene) and used to transform *E. coli* XL-1Blue (Stratagene).

The complete DNA sequence of both genes were obtained by the dideoxy chain termination method adapted for supercoiled DNA by use of the Sequenase DNA sequencing kit version 1.0 (United States Biochemical Corp., Cleveland, OH) and by cycle sequencing using the Dye Terminator system in combination with an automated gel reader (model 373A; Applied Biosystems) according to the instructions provided. The sequences DNA are shown in SEQ ID NO: 1 (CFP7) and in SEQ ID NO: 3 (CFP9) as well as in Figs. 3 and 4, respectively. Both strands of the DNA were sequenced.

CFP7

An open reading frame (ORF) encoding a sequence of 96 amino acid residues was identified from an ATG start codon at position 91-93 extending to a TAG stop codon at position 379-381. The deduced amino acid sequence is shown in SEQ ID NO: 2 (and in Fig. 3 where conventional one-letter amino acid codes are used).

CFP7 appear to be expressed in *E. coli* as an unfused version. The nucleotide sequence at position 78-84 is expected to be the Shine Delgarno sequence and the sequences from position 47-50 and 14-19 are expected to be the -10 and -35 regions, respectively:

CFP9

The protein recognised by ST-3 was produced as a  $\beta$ -galactosidase fusion protein, when expressed from the AA226 lambda phage. The fusion protein had an approx. size of 116 - 117kDa (Mw for  $\beta$ -galactosidase 116.25 kDa) which may suggest that only part of the CFP9 gene was included in the lambda clone (AA226).

Based on the 90 bp nucleotide sequence obtained on the insert from lambda phage AA226, a search of homology to the nucleotide sequence of the *M. tuberculosis* genome was performed in the Sanger database (Sanger *Mycobacterium tuberculosis* database):

<http://www.sanger.ac.uk/pathogens/TB-blast-server.html>;

Williams, 1996). 100% identity to the cloned sequence was found on the MTCY48 cosmid. An open reading frame (ORF) encoding a sequence of 109 amino acid residues was identified from a GTG start codon at position 141 - 143 extending to a TGA stop codon at position 465 - 467. The deduced amino acid

sequence is shown in Fig. 4 using conventional one letter code.

The nucleotide sequence at position 123 - 130 is expected to be the Shine Delgarno sequence and the sequences from position 73 - 78 and 4 - 9 are expected to be the -10 and -35 region respectively (Fig. 4). The ORF overlapping with the 5'-end of the sequence of AA229 is shown in Fig. 4 by double underlining.

#### Subcloning CFP7 and CFP9 in expression vectors

10

The two ORFs encoding CFP7 and CFP9 were PCR cloned into the pMST24 (Theisen et al., 1995) expression vector pRVN01 or the pQE-32 (QIAGEN) expression vector pRVN02, respectively.

The PCR amplification was carried out in a thermal reactor (Rapid cycler, Idaho Technology, Idaho) by mixing 10 ng plasmid DNA with the mastermix (0.5  $\mu$ M of each oligonucleotide primer, 0.25  $\mu$ M BSA (Stratagene), low salt buffer (20 mM Tris-HCl, pH 8.8, 10 mM KCl, 10 mM  $(\text{NH}_4)_2\text{SO}_4$ , 2 mM  $\text{MgSO}_4$  and 0.1% Triton X-100) (Stratagene), 0.25 mM of each deoxynucleoside triphosphate and 0.5 U Taq Plus Long DNA polymerase (Stratagene)). Final volume was 10  $\mu$ l (all concentrations given are concentrations in the final volume). Predenaturation was carried out at 94°C for 30 s. 30 cycles of the following was performed; Denaturation at 94°C for 30 s, annealing at 55°C for 30 s and elongation at 72°C for 1 min.

The oligonucleotide primers were synthesised automatically on a DNA synthesizer (Applied Biosystems, Foster City, Ca, ABI-391, PCR-mode), deblocked, and purified by ethanol precipitation.

The *cfp7* oligonucleotides (TABLE 1) were synthesised on the basis of the nucleotide sequence from the CFP7 sequence (Fig. 3). The oligonucleotides were engineered to include an *Sma*I

restriction enzyme site at the 5' end and a *Bam*HI restriction enzyme site at the 3' end for directed subcloning.

The *cfp9* oligonucleotides (TABLE 1) were synthesized partly on the basis of the nucleotide sequence from the sequence of the AA229 clone and partly from the identical sequence found in the Sanger database cosmid MTCY48 (Fig. 4). The oligonucleotides were engineered to include a *Sma*I restriction enzyme site at the 5' end and a *Hind*III restriction enzyme site at the 3' end for directed subcloning.

#### 10 CFP7

By the use of PCR a *Sma*I site was engineered immediately 5' of the first codon of the ORF of 291 bp, encoding the *cfp7* gene, so that only the coding region would be expressed, and a *Bam*HI site was incorporated right after the stop codon at the 3' end. The 291 bp PCR fragment was cleaved by *Sma*I and *Bam*HI, purified from an agarose gel and subcloned into the *Sma*I - *Bam*HI sites of the pMST24 expression vector. Vector DNA containing the gene fusion was used to transform the *E. coli* XL1-Blue (pRVN01).

#### 20 CFP9

By the use of PCR a *Sma*I site was engineered immediately 5' of the first codon of an ORF of 327 bp, encoding the *cfp9* gene, so that only the coding region would be expressed, and a *Hind*III site was incorporated after the stop codon at the 3' end. The 327 bp PCR fragment was cleaved by *Sma*I and *Hind*III, purified from an agarose gel, and subcloned into the *Sma*I - *Hind*III sites of the pQE-32 (QIAGEN) expression vector. Vector DNA containing the gene fusion was used to transform the *E. coli* XL1-Blue (pRVN02).

Purification of recombinant CFP7 and CFP9

The ORFs were fused N-terminally to the (His)<sub>6</sub>-tag (cf. EP-A-0 282 242). Recombinant antigen was prepared as follows: Briefly, a single colony of *E. coli* harbouring either the  
5 pRVN01 or the pRVN02 plasmid, was inoculated into Luria-Bertani broth containing 100 µg/ml ampicillin and 12.5 µg/ml tetracycline and grown at 37°C to OD<sub>600nm</sub> = 0.5. IPTG (isopropyl-β-D-thiogalactoside) was then added to a final concentration of 2 mM (expression was regulated either by the  
10 strong IPTG inducible P<sub>tac</sub> or the T5 promoter) and growth was continued for further 2 hours. The cells were harvested by centrifugation at 4,200 x g at 4°C for 8 min. The pelleted bacteria were stored overnight at -20°C. The pellet was resuspended in BC 40/100 buffer (20 mM Tris-HCl pH 7.9, 20%  
15 glycerol, 100 mM KCl, 40 mM Imidazole) and cells were broken by sonication (5 times for 30 s with intervals of 30 s) at 4°C. followed by centrifugation at 12,000 x g for 30 min at 4°C, the supernatant (crude extract) was used for purification of the recombinant antigens.

20 The two Histidine fusion proteins (His-rCFP7 and His-rCFP9) were purified from the crude extract by affinity chromatography on a Ni<sup>2+</sup>-NTA column from QIAGEN with a volume of 100 ml. His-rCFP7 and His-rCFP9 binds to Ni<sup>2+</sup>. After extensive washes of the column in BC 40/100 buffer, the fusion protein  
25 was eluted with a BC 1000/100 buffer containing 100 mM imidazole, 20 mM Tris pH 7.9, 20% glycerol and 1 M KCl. subsequently, the purified products were dialysed extensively against 10 mM Tris pH 8.0. His-rCFP7 and His-rCFP9 were then separated from contaminants by fast protein liquid chromatography (FPLC) over an anion-exchange column (Mono Q, Pharmacia, Sweden). in 10 mM Tris pH 8.0 with a linear gradient of  
30 NaCl from 0 to 1 M. Aliquots of the fractions were analyzed by 10%-20% gradient sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE). Fractions containing purified  
35 either purified His-rCFP7 or His-rCFP9 were pooled.



TABLE 1. Sequence of the *cfp7* and *cfp9* oligonucleotides<sup>a</sup>.

Orientation and oligonucleotide		Sequences (5' → 3')	Position <sup>b</sup> (nucleotide)
Sense			
5	pvr3	<u>GCAACACCCGGG</u> ATGTCGCAAATCATG (SEQ ID NO: 43)	91-105 (SEQ ID NO: 1)
	str2	<u>GTAACACCCGGG</u> GTGGCCGCCGACCCG (SEQ ID NO: 44)	141-155 (SEQ ID NO: 3)
Antisense			
	pVF4	CTACTAAGCTT <u>GGATCC</u> CTAGCCGCCCCATTTGGCGG (SEQ ID NO: 45)	381-362 (SEQ ID NO: 1)
	stF2	CTACTAAGCTT <u>CCATGG</u> TCAGGTCTTTTCGATGCTTAC (SEQ ID NO: 46)	467 - 447 (SEQ ID NO: 3)
10	<sup>a</sup> The <i>cfp7</i> oligonucleotides were based on the nucleotide sequence shown in Fig. 3 (SEQ ID NO: 1). The <i>cfp9</i> oligonucleotides were based on the nucleotide sequence shown in Fig. 4 (SEQ ID NO: 3). Nucleotides underlined are not contained in the nucleotide sequence of <i>cfp7</i> and <i>cfp9</i> .		
15	<sup>b</sup> The positions referred to are of the non-underlined part of the primers and correspond to the nucleotide sequence shown in Fig. 3 and Fig. 4, respectively.		

## EXAMPLE 2A

20 *Identification of antigens which are not expressed in BCG strains.*

In an effort to control the treat of TB, attenuated bacillus Calmette-Guérin (BCG) has been used as a live attenuated vaccine. BCG is an attenuated derivative of a virulent *Mycobacterium bovis*. The original BCG from the Pasteur Institute

25 in Paris, France was developed from 1908 to 1921 by 231 passages in liquid culture and has never been shown to revert to virulence in animals, indicating that the attenuating mutation(s) in BCG are stable deletions and/or multiple mutations which do not readily revert. While physiological

30 differences between BCG and *M. tuberculosis* and *M. bovis* has been noted, the attenuating mutations which arose during serial passage of the original BCG strain has been unknown until recently. The first mutations described are the loss of the gene encoding MPB64 in some BCG strains (Li et al., 1993,

35 Oettinger and Andersen, 1994) and the gene encoding ESAT-6 in all BCG strain tested (Harboe et al., 1996), later 3 large deletions in BCG have been identified (Mahairas et al., 1996). The region named RD1 includes the gene encoding ESAT-6

and an other (RD2) the gene encoding MPT64. Both antigens have been shown to have diagnostic potential and ESAT-6 has been shown to have properties as a vaccine candidate (cf. PCT/DK94/00273 and PCT/DK/00270). In order to find new *M. tuberculosis* specific diagnostic antigens as well as antigens for a new vaccine against TB, the RD1 region (17.499 bp) of *M. tuberculosis* H37Rv has been analyzed for Open Reading Frames (ORF). ORFs with a minimum length of 96 bp have been predicted using the algorithm described by Borodovsky and McIninch (1993), in total 27 ORFs have been predicted, 20 of these have possible diagnostic and/or vaccine potential, as they are deleted from all known BCG strains. The predicted ORFs include ESAT-6 (RD1-ORF7) and CFP10 (RD1-ORF6) described previously (Sørensen et al., 1995), as a positive control for the ability of the algorithm. In the present is described the potential of 7 of the predicted antigens for diagnosis of TB as well as potential as candidates for a new vaccine against TB.

Seven open reading frames (ORF) from the 17,499kb RD1 region (Accession no. U34848) with possible diagnostic and vaccine potential have been identified and cloned.

Identification of the ORF's *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a*, and *rd1-orf9b*.

The nucleotide sequence of *rd1-orf2* from *M. tuberculosis* H37Rv is set forth in SEQ ID NO: 71. The deduced amino acid sequence of RD1-ORF2 is set forth in SEQ ID NO: 72.

The nucleotide sequence of *rd1-orf3* from *M. tuberculosis* H37Rv is set forth in SEQ ID NO: 87. The deduced amino acid sequence of RD1-ORF2 is set forth in SEQ ID NO: 88.

The nucleotide sequence of *rd1-orf4* from *M. tuberculosis* H37Rv is set forth in SEQ ID NO: 89. The deduced amino acid sequence of RD1-ORF2 is set forth in SEQ ID NO: 90.

The nucleotide sequence of *rd1-orf5* from *M. tuberculosis* H37Rv is set forth in SEQ ID NO: 91. The deduced amino acid sequence of RD1-ORF2 is set forth in SEQ ID NO: 92.

The nucleotide sequence of *rd1-orf8* from *M. tuberculosis* H37Rv is set forth in SEQ ID NO: 67. The deduced amino acid sequence of RD1-ORF2 is set forth in SEQ ID NO: 68.

The nucleotide sequence of *rd1-orf9a* from *M. tuberculosis* H37Rv is set forth in SEQ ID NO: 93. The deduced amino acid sequence of RD1-ORF2 is set forth in SEQ ID NO: 94.

10 The nucleotide sequence of *rd1-orf9b* from *M. tuberculosis* H37Rv is set forth in SEQ ID NO: 69. The deduced amino acid sequence of RD1-ORF2 is set forth in SEQ ID NO: 70.

The DNA sequence *rd1-orf2* (SEQ ID NO: 71) contained an open reading frame starting with an ATG codon at position 889 -  
15 891 and ending with a termination codon (TAA) at position 2662 - 2664 (position numbers referring to the location in RD1). The deduced amino acid sequence (SEQ ID NO: 72) contains 591 residues corresponding to a molecular weight of 64,525.

20 The DNA sequence *rd1-orf3* (SEQ ID NO: 87) contained an open reading frame starting with an ATG codon at position 2807 - 2809 and ending with a termination codon (TAA) at position 3101 - 3103 (position numbers referring to the location in RD1). The deduced amino acid sequence (SEQ ID NO: 88) contains 98 residues corresponding to a molecular weight of  
25 9,799.

The DNA sequence *rd1-orf4* (SEQ ID NO: 89) contained an open reading frame starting with a GTG codon at position 4014 - 4012 and ending with a termination codon (TAG) at position  
30 3597 - 3595 (position numbers referring to the location in RD1). The deduced amino acid sequence (SEQ ID NO: 90) con-

tains 139 residues corresponding to a molecular weight of 14,210.

The DNA sequence rd1-orf5 (SEQ ID NO: 91) contained an open reading frame starting with a GTG codon at position 3128 -  
5 3130 and ending with a termination codon (TGA) at position 4241 - 4243 (position numbers referring to the location in RD1). The deduced amino acid sequence (SEQ ID NO: 92) contains 371 residues corresponding to a molecular weight of 37,647.

10 The DNA sequence rd1-orf8 (SEQ ID NO: 67) contained an open reading frame starting with a GTG codon at position 5502 - 5500 and ending with a termination codon (TAG) at position 5084 - 5082 (position numbers referring to the location in RD1), and the deduced amino acid sequence (SEQ ID NO: 68)  
15 contains 139 residues with a molecular weight of 11,737.

The DNA sequence rd1-orf9a (SEQ ID NO: 93) contained an open reading frame starting with a GTG codon at position 6146 - 6148 and ending with a termination codon (TAA) at position 7070 - 7072 (position numbers referring to the location in  
20 RD1). The deduced amino acid sequence (SEQ ID NO: 94) contains 308 residues corresponding to a molecular weight of 33,453.

The DNA sequence rd1-orf9b (SEQ ID NO: 69) contained an open reading frame starting with an ATG codon at position 5072 -  
25 5074 and ending with a termination codon (TAA) at position 7070 - 7072 (position numbers referring to the location in RD1). The deduced amino acid sequence (SEQ ID NO: 70) contains 666 residues corresponding to a molecular weight of 70,650.

Cloning of the ORF's *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a*, and *rd1-orf9b*.

The ORF's *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a* and *rd1-orf9b* were PCR cloned in the pMST24 (Thei-  
 5 sen et al., 1995) (*rd1-orf3*) or the pQE32 (QIAGEN) (*rd1-orf2*,  
*rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a* and *rd1-orf9b*) ex-  
 pression vector. Preparation of oligonucleotides and PCR  
 amplification of the *rd1-orf* encoding genes, was carried out  
 as described in example 2. Chromosomal DNA from *M. tuberculo-*  
 10 *sis* H37Rv was used as template in the PCR reactions. Oligonu-  
 cleotides were synthesized on the basis of the nucleotide  
 sequence from the RD1 region (Accession no. U34848). The  
 oligonucleotide primers were engineered to include an re-  
 striction enzyme site at the 5' end and at the 3' end by  
 15 which a later subcloning was possible. Primers are listed in  
 TABLE 2.

*rd1-orf2*. A *Bam*HI site was engineered immediately 5' of the  
 first codon of *rd1-orf2*, and a *Hind*III site was incorporated  
 right after the stop codon at the 3' end. The gene *rd1-orf2*  
 20 was subcloned in pQE32, giving pT096.

*rd1-orf3*. A *Sma*I site was engineered immediately 5' of the  
 first codon of *rd1-orf3*, and a *Nco*I site was incorporated  
 right after the stop codon at the 3' end. The gene *rd1-orf3*  
 was subcloned in pMST24, giving pT087.

25 *rd1-orf4*. A *Bam*HI site was engineered immediately 5' of the  
 first codon of *rd1-orf4*, and a *Hind*III site was incorporated  
 right after the stop codon at the 3' end. The gene *rd1-orf4*  
 was subcloned in pQE32, giving pT089.

30 *rd1-orf5*. A *Bam*HI site was engineered immediately 5' of the  
 first codon of *rd1-orf5*, and a *Hind*III site was incorporated  
 right after the stop codon at the 3' end. The gene *rd1-orf5*  
 was subcloned in pQE32, giving pT088.

*rd1-orf8*. A *Bam*HI site was engineered immediately 5' of the first codon of *rd1-orf8*, and a *Nco*I site was incorporated right after the stop codon at the 3' end. The gene *rd1-orf8* was subcloned in pMST24, giving pT098.

- 5 *rd1-orf9a*. A *Bam*HI site was engineered immediately 5' of the first codon of *rd1-orf9a*, and a *Hind*III site was incorporated right after the stop codon at the 3' end. The gene *rd1-orf9a* was subcloned in pQE32, giving pT091.

- 10 *rd1-orf9b*. A *Sca*I site was engineered immediately 5' of the first codon of *rd1-orf9b*, and a *Hind* III site was incorporated right after the stop codon at the 3' end. The gene *rd1-orf9b* was subcloned in pQE32, giving pT090.

- 15 The PCR fragments were digested with the suitable restriction enzymes, purified from an agarose gel and cloned into either pMST24 or pQE-32. The seven constructs were used to transform the *E. coli* XL1-Blue. Endpoints of the gene fusions were determined by the dideoxy chain termination method. Both strands of the DNA were sequenced.

- 20 Purification of recombinant RD1-ORF2, RD1-ORF3, RD1-ORF4, RD1-ORF5, RD1-ORF8, RD1-ORF9a and RD1-ORF9b.

- The rRD1-ORFs were fused N-terminally to the (His)<sub>6</sub> -tag. Recombinant antigen was prepared as described in example 2 (with the exception that pT091 was expressed at 30°C and not  
25 at 37°C), using a single colony of *E. coli* harbouring either the pT087, pT088, pT089, pT090, pT091, pT096 or pT098 for inoculation. Purification of recombinant antigen by Ni<sup>2+</sup> affinity chromatography was also carried out as described in example 2. Fractions containing purified His-rRD1-ORF2, His-  
30 rRD1-ORF3 His-rRD1-ORF4, His-rRD1-ORF5, His-rRD1-ORF8, His-rRD1-ORF9a or His-rRD1-ORF9b were pooled. The His-rRD1-ORF's were extensively dialysed against 10 mM Tris/HCl, pH 8.5, 3 M urea followed by an additional purification step performed on an anion exchange column (Mono Q) using fast protein liquid

chromatography (FPLC) (Pharmacia, Uppsala, Sweden). The purification was carried out in 10 mM Tris/HCl, pH 8.5, 3 M urea and protein was eluted by a linear gradient of NaCl from 0 to 1 M. Fractions containing the His-rRD1-ORF's were pooled and subsequently dialysed extensively against 25 mM Hepes, pH 8.0 before use.

Table 2. Sequence of the *rd1-orf's* oligonucleotides<sup>a</sup>.

	Orientation and oligonucleotide	Sequences (5'→ 3')	Position (nt)
10	Sense		
	RD1-ORF2f	<u>CTGGGGATCCGCATGACTGCTGAACCG</u>	886 - 903
	RD1-ORF3f	<u>CTTCCCGGGATGGAAAAATGTCAC</u>	2807 - 2822
	RD1-ORF4f	<u>GTAGGATCCTAGGAGACATCAGCGGC</u>	4028 - 4015
	RD1-ORF5f	<u>CTGGGGATCCGCGTGATCACCATGCTGTGG</u>	3028 - 3045
15	RD1-ORF8f	<u>CTCGGATCCTGTGGGTGCAGGTCCGGCGATGGGC</u>	5502 - 5479
	RD1-ORF9af	<u>GTGATGTGAGCTCAGGTGAAGAAGGTGAAG</u>	6144 - 6160
	RD1-ORF9bf	<u>GTGATGTGAGCTCCTATGGCGGCCGACTACGAC</u>	5072 - 5089
	Antisense		
	RD1-ORF2r	<u>TGCAAGCTTTTAACCGCGCTTGGGGGTGC</u>	2664 - 2644
20	RD1-ORF3r	<u>GATGCCATGGTTAGGCGAAGACGCCGGC</u>	3103 - 3086
	RD1-ORF4r	<u>CGATCTAAGCTTGGCAATGGAGGTCTA</u>	3582 - 3597
	RD1-ORF5r	<u>TGCAAGCTTTCACCAAGTCGTCCTCTTCGTC</u>	4243 - 4223
	RD1-ORF8r	<u>CTCCCATGGCTACGACAAGCTCTTCCGGCCGC</u>	5083 - 5105
	RD1-ORF9a/br	<u>CGATCTAAGCTTTCAACGACGTCCAGCC</u>	7073 - 7056

<sup>a</sup> The oligonucleotides were constructed from the Accession number U34484 nucleotide sequence (Mahairas et al., 1996). Nucleotides (nt) underlined are not contained in the nucleotide sequence of RD1-ORF's. The positions correspond to the nucleotide sequence of Accession number U34484.

The nucleotide sequences of *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a*, and *rd1-orf9b* from *M. tuberculosis* H37Rv are set forth in SEQ ID NO: 71, 87, 89, 91, 67, 93, and 69, respectively. The deduced amino acid sequences of *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a*, and *rd1-orf9b* are set forth in SEQ ID NO: 72, 88, 90, 92, 68, 94, and 70, respectively.

## EXAMPLE 3

*Cloning of the genes expressing 17-30 kDa antigens from ST-CF*Isolation of CFP17, CFP20, CFP21, CFP22, CFP25, and CFP28

ST-CF was precipitated with ammonium sulphate at 80% saturation. The precipitated proteins were removed by  
5 centrifugation and after resuspension washed with 8 M urea. CHAPS and glycerol were added to a final concentration of 0.5% (w/v) and 5% (v/v) respectively and the protein solution was applied to a Rotofor isoelectrical Cell (BioRad). The  
10 Rotofor Cell had been equilibrated with an 8 M urea buffer containing 0.5% (w/v) CHAPS, 5% (v/v) glycerol, 3% (v/v) Biolyt 3/5 and 1% (v/v) Biolyt 4/6 (BioRad). Isoelectric focusing was performed in a pH gradient from 3-6. The fractions were analyzed on silver-stained 10-20% SDS-PAGE. Frac-  
15 tions with similar band patterns were pooled and washed three times with PBS on a Centriprep concentrator (Amicon) with a 3 kDa cut off membrane to a final volume of 1-3 ml. An equal volume of SDS containing sample buffer was added and the protein solution boiled for 5 min before further separation  
20 on a Prep Cell (BioRad) in a matrix of 16% polyacrylamide under an electrical gradient. Fractions containing pure proteins with an molecular mass from 17-30 kDa were collected.

Isolation of CFP29

25 Anti-CFP29, reacting with CFP29 was generated by immunization of BALB/c mice with crushed gel pieces in RIBI adjuvant (first and second immunization) or aluminium hydroxide (third immunization and boosting) with two week intervals. SDS-PAGE gel pieces containing 2-5  $\mu$ g of CFP29 were used for each  
30 immunization. Mice were boosted with antigen 3 days before removal of the spleen. Generation of a monoclonal cell line producing antibodies against CFP29 was obtained essentially as described by Köhler and Milstein (1975). Screening of



supernatants from growing clones was carried out by immunoblotting of nitrocellulose strips containing ST-CF separated by SDS-PAGE. Each strip contained approximately 50  $\mu$ g of ST-CF. The antibody class of anti-CFP29 was identified as IgM by the mouse monoclonal antibody isotyping kit, RPN29 (Amersham) according to the manufacturer's instructions.

CFP29 was purified by the following method: ST-CF was concentrated 10 fold by ultrafiltration, and ammonium sulphate precipitation in the 45 to 55% saturation range was performed. The pellet was redissolved in 50 mM sodium phosphate, 1.5 M ammonium sulphate, pH 8.5, and subjected to thiophilic adsorption chromatography (Porath et al., 1985) on an Affi-T gel column (Kem-En-Tec). Protein was eluted by a linear 1.5 to 0 M gradient of ammonium sulphate and fractions collected in the range 0.44 to 0.31 M ammonium sulphate were identified as CFP29 containing fractions in Western blot experiments with mAb Anti-CFP29. These fractions were pooled and anion exchange chromatography was performed on a Mono Q HR 5/5 column connected to an FPLC system (Pharmacia). The column was equilibrated with 10 mM Tris-HCl, pH 8.5 and the elution was performed with a linear gradient from 0 to 500 mM NaCl. From 400 to 500 mM sodium chloride, rather pure CFP29 was eluted. As a final purification step the Mono Q fractions containing CFP29 were loaded on a 12.5% SDS-PAGE gel and pure CFP29 was obtained by the multi-elution technique (Andersen and Heron, 1993).

#### N-terminal sequencing and amino acid analysis

CFP17, CFP20, CFP21, CFP22, CFP25, and CFP28 were washed with water on a Centricon concentrator (Amicon) with cutoff at 10 kDa and then applied to a ProSpin concentrator (Applied Biosystems) where the proteins were collected on a PVDF membrane. The membrane was washed 5 times with 20% methanol before sequencing on a Procise sequencer (Applied Biosystems).

CFP29 containing fractions were blotted to PVDF membrane after tricine SDS-PAGE (Ploug et al., 1989). The relevant bands were excised and subjected to amino acid analysis (Barkholt and Jensen, 1989) and N-terminal sequence analysis  
 5 on a Procise sequencer (Applied Biosystems).

The following N-terminal sequences were obtained:

	For CFP17: A/S E L D A P A Q A G T E X A V	(SEQ ID NO: 17)
	For CFP20: A Q I T L R G N A I N T V G E	(SEQ ID NO: 18)
	For CFP21: D P X S D I A V V F A R G T H	(SEQ ID NO: 19)
10	For CFP22: T N S P L A T A T A T L H T N	(SEQ ID NO: 20)
	For CFP25: A X P D A E V V F A R G R F E	(SEQ ID NO: 21)
	For CFP28: X I/V Q K S L E L I V/T V/F T A D/Q E	(SEQ ID NO: 22)
	For CFP29: M N N L Y R D L A P V T E A A W A E I	(SEQ ID NO: 23)

"X" denotes an amino acid which could not be determined by  
 15 the sequencing method used, whereas a "/" between two amino acids denotes that the sequencing method could not determine which of the two amino acids is the one actually present.

#### Cloning the gene encoding CFP29

The N-terminal sequence of CFP29 was used for a homology  
 20 search in the EMBL database using the TFASTA program of the Genetics Computer Group sequence analysis software package. The search identified a protein, Linocin M18, from *Brevibacterium linens* that shares 74% identity with the 19 N-terminal amino acids of CFP29.

25 Based on this identity between the N-terminal sequence of CFP29 and the sequence of the Linocin M18 protein from *Brevibacterium linens*, a set of degenerated primers were constructed for PCR cloning of the *M. tuberculosis* gene encoding CFP29. PCR reactions were containing 10 ng of *M. tuberculosis*  
 30 chromosomal DNA in 1 x low salt Taq+ buffer from Stratagene supplemented with 250 µM of each of the four nucleotides (Boehringer Mannheim), 0,5 mg/ml BSA (IgG technology), 1% DMSO (Merck), 5 pmoles of each primer and 0.5 unit Tag+ DNA

polymerase (Stratagene) in 10  $\mu$ l reaction volume. Reactions were initially heated to 94°C for 25 sec. and run for 30 cycles of the program; 94°C for 15 sec., 55°C for 15 sec. and 72°C for 90 sec, using thermocycler equipment from Idaho Technology.

An approx. 300 bp fragment was obtained using primers with the sequences:

1: 5'-CCCGGCTCGAGAACCTSTACCGCGACCTSGCSCC (SEQ ID NO: 24)

2: 5'-GGGCCGGATCCGASGCSGCGTCCTTSACSGGYTGCCA (SEQ ID NO: 25)

10 -where S = G/C and Y = T/C

The fragment was excised from a 1% agarose gel, purified by Spin-X spin columns (Costar), cloned into pBluescript SK II+ - T vector (Stratagene) and finally sequenced with the Sequenase kit from United States Biochemical.

15 The first 150 bp of this sequence was used for a homology search using the Blast program of the Sanger *Mycobacterium tuberculosis* database:

([http://www.sanger.ac.uk/projects/M-tuberculosis/blast\\_server](http://www.sanger.ac.uk/projects/M-tuberculosis/blast_server)).

20 This program identified a *Mycobacterium tuberculosis* sequence on cosmid cy444 in the database that is nearly 100% identical to the 150 bp sequence of the CFP29 protein. The sequence is contained within a 795 bp open reading frame of which the 5' end translates into a sequence that is 100% identical to the N-terminally sequenced 19 amino acids of the purified CFP29  
25 protein.

Finally, the 795 bp open reading frame was PCR cloned under the same PCR conditions as described above using the primers:

3: 5'-GGAAGCCCCATATGAACAATCTCTACCG (SEQ ID NO: 26)

4: 5'-CGCGCTCAGCCCTTAGTGACTGAGCGCGACCG (SEQ ID NO: 27)

The resulting DNA fragments were purified from agarose gels as described above sequenced with primer 3 and 4 in addition to the following primers:

- 5: 5'-GGACGTTCAAGCGACACATCGCCG-3' (SEQ ID NO: 115)  
 5 6: 5'-CAGCACGAACGCGCCGTCGATGGC-3' (SEQ ID NO: 116)

Three independent clones were sequenced. All three clones were in 100% agreement with the sequence on cosmid cy444.

All other DNA manipulations were done according to Maniatis et al. (1989).

- 10 All enzymes other than Taq polymerase were from New England Biolabs.

#### Homology searches in the Sanger database

- For CFP17, CFP20, CFP21, CFP22, CFP25, and CFP28 the N-terminal amino acid sequence from each of the proteins were used  
 15 for a homology search using the blast program of the Sanger *Mycobacterium tuberculosis* database:

<http://www.sanger.ac.uk/pathogens/TB-blast-server.html>.

- For CFP29 the first 150 bp of the DNA sequence was used for the search. Furthermore, the EMBL database was searched for  
 20 proteins with homology to CFP29.

Thereby, the following information were obtained:

#### CFP17

- Of the 14 determined amino acids in CFP17 a 93% identical sequence was found with MTCY1A11.16c. The difference between  
 25 the two sequences is in the first amino acid: It is an A or an S in the N-terminal determined sequenced and a S in

MTCY1A11. From the N-terminal sequencing it was not possible to determine amino acid number 13.

Within the open reading frame the translated protein is 162 amino acids long. The N-terminal of the protein purified from culture filtrate starts at amino acid 31 in agreement with the presence of a signal sequence that has been cleaved off. This gives a length of the mature protein of 132 amino acids, which corresponds to a theoretical molecular mass of 13833 Da and a theoretical pI of 4.4. The observed mass in SDS-PAGE is 17 kDa.

#### CFP20

A sequence 100% identical to the 15 determined amino acids of CFP20 was found on the translated cosmid cscy09F9. A stop codon is found at amino acid 166 from the amino acid M at position 1. This gives a predicted length of 165 amino acids, which corresponds to a theoretical molecular mass of 16897 Da and a pI of 4.2. The observed molecular weight in a SDS-PAGE is 20 kDa.

Searching the GenEMBL database using the TFASTA algorithm (Pearson and Lipman, 1988) revealed a number of proteins with homology to the predicted 164 amino acids long translated protein.

The highest homology, 51.5% identity in a 163 amino acid overlap, was found to a Haemophilus influenza Rd toxR reg. (HIHI0751).

#### CFP21

A sequence 100% identical to the 14 determined amino acids of CFP21 was found at MTCY39. From the N-terminal sequencing it was not possible to determine amino acid number 3; this amino acid is a C in MTCY39. The amino acid C can not be detected

on a Sequencer which is probably the explanation of this difference.

Within the open reading frame the translated protein is 217 amino acids long. The N-terminally determined sequence from  
 5 the protein purified from culture filtrate starts at amino acid 33 in agreement with the presence of a signal sequence that has been cleaved off. This gives a length of the mature protein of 185 amino acids, which corresponds to a theoretical molecular weight at 18657 Da, and a theoretical pI at 4,6.  
 10 The observed weight in a SDS-PAGE is 21 kDa.

In a 193 amino acids overlap the protein has 32,6% identity to a cutinase precursor with a length of 209 amino acids (CUTI\_ALTBR P41744).

A comparison of the 14 N-terminal determined amino acids with  
 15 the translated region (RD2) deleted in *M. bovis* BCG revealed a 100% identical sequence (mb3484) (Mahairas et al. (1996)).

#### CFP22

A sequence 100% identical to the 15 determined amino acids of CFP22 was found at MTCY10H4. Within the open reading frame  
 20 the translated protein is 182 amino acids long. The N-terminal sequence of the protein purified from culture filtrate starts at amino acid 8 and therefore the length of the protein occurring in *M. tuberculosis* culture filtrate is 175 amino acids. This gives a theoretical molecular weight at  
 25 18517 Da and a pI at 6.8. The observed weight in a SDS-PAGE is 22 kDa.

In an 182 amino acids overlap the translated protein has 90,1% identity with E235739; a peptidyl-prolyl cis-trans isomerase.

CFP25

A sequence 93% identical to the 15 determined amino acids was found on the cosmid MTCY339.08c. The one amino acid that differs between the two sequences is a C in MTCY339.08c and a  
 5 X from the N-terminal sequence data. On a Sequencer a C can not be detected which is a probable explanation for this difference.

The N-terminally determined sequence from the protein purified from culture filtrate begins at amino acid 33 in  
 10 agreement with the presence of a signal sequence that has been cleaved off. This gives a length of the mature protein of 187 amino acids, which corresponds to a theoretical molecular weight at 19665 Da, and a theoretical pI at 4.9. The observed weight in a SDS-PAGE is 25 kDa.

15 In a 217 amino acids overlap the protein has 42.9% identity to CFP21 (MTCY39.35).

CFP28

No homology was found when using the 10 determined amino acid residues 2-8, 11, 12, and 14 of SEQ ID NO: 22 in the database  
 20 search.

CFP29

Sanger database searching: A sequence nearly 100% identical to the 150 bp sequence of the CFP29 protein was found on cosmid cy444. The sequence is contained within a 795 bp open  
 25 reading frame of which the 5' end translates into a sequence that is 100% identical to the N-terminally sequenced 19 amino acids of the purified CFP29 protein. The open reading frame encodes a 265 amino acid protein.

The amino acid analysis performed on the purified protein further confirmed the identity of CFP29 with the protein encoded in open reading frame on cosmid 444.

EMBL database searching: The open reading frame encodes a 265  
 5 amino acid protein that is 58% identical and 74% similar to  
 the Linocin M18 protein (61% identity on DNA level). This is  
 a 28.6 kDa protein with bacteriocin activity (Valdés-Stauber  
 and Scherer, 1994; Valdés-Stauber and Scherer, 1996). The two  
 10 proteins have the same length (except for 1 amino acid) and  
 share the same theoretical physicochemical properties. We  
 therefore suggest that CFP29 is a mycobacterial homolog to  
 the *Brevibacterium linens* Linocin M18 protein.

The amino acid sequences of the purified antigens as picked  
 from the Sanger database are shown in the following list. The  
 15 amino acids determined by N-terminal sequencing are marked  
 with bold.

CFP17 (SEQ ID NO: 6):

1 MTDMNPDIEK DQTSDEVTVE TTSVFRADFL **SELDAPAQAG** **TESAVSGVEG**  
 51 LPPGSALLVV KRGPNAGSRF LLDQAITSAG RHPDSDIFLD DVTVSRRHAE  
 20 101 FRLENNEFNV VDVGSLNGTY VNREPVDSAV LANGDEVQIG KFRLVFLTGP  
 151 KQGEDDGSTG GP

CFP20 (SEQ ID NO: 8):

1 **MAQITLRGNA** **INTVGELPAV** GSPAPAFTLT GGD LGVISSD QFRGKSVLLN  
 51 IFPSVDTPVC ATSVRTFDER AAASGATVLC VSKDLPFAQK RFCGAEGTEN  
 25 101 VMPASAFRDS FGEDYGVTTIA DGPMAGLLAR AIVVIGADGN VAYTELVPEI  
 151 AQEPNYEAL AALGA

CFP21 (SEQ ID NO: 10):

1 MTPRSLVRIV GVVVATTLLAL VSAPAGGRAA **HADPCSDIAV**  
 41 **VFARGTHQAS** GLGDVGEAFV DSLTSQVGGR SIGVYAVNYP ASDDYRASAS  
 30 91 NGSDDASAHI QRTVASCPNT RIVLGGYSQG ATVIDLSTSA MPPAVADHVA



141 AVALFGEPSS GFSSMLWGGG SLPTIGPLYS SKTINLCAPD DPICTGGGNI  
 191 MAHVSIVQSG MTSQAATFAA NRLDHAG

CFP22 (SEQ ID NO: 12):

1 MADCDSVTNS PLATATATLE TNRGDIKIAL FGNHAPKTVA NFVGLAQGTK  
 5 51 DYSTQNASGG PSGPFYDGAV FHRVIQGFMI QGGDPTGTGR GGPYKFADE  
 101 FHPELQFDKP YLLAMANAGP GTNGSQFFIT VGKTPHLNRR HTIFGEVIDA  
 151 ESQRVVEAIS KTATDGNDRP TDPVVIESIT IS

CFP25 (SEQ ID NO: 14):

1 MGAAAAMLAA VLLLTPTITVP AGYPGAVAPA TAACPDAEVV FARGRFEPPG  
 10 51 IGTVGNAFVS ALRSKVNKNV GYAVKYPAD NQIDVGANDM SAHIQSMANS  
 101 CPNTRLVPGG YSLGAAVTDV VLAVPTQMWG FTNPLPPGSD EHIAAVALFG  
 151 NGSQWVGPIIT NFSPAYNDRT IELCHGDDPV CHPADPNTWE ANWPQHLAGA  
 201 YVSSGMVNQA ADFVAGKLQ

CFP29 (SEQ ID NO: 16):

15 1 MNNLYRDLAP VTEAAWAEIE LEAARTFKRH IAGRRVVDVS DPGGPVTAAG  
 51 STGRLIDVKA PTNGVIAHLR ASKPLVRLRV PFTLSRNEID DVERGSKDSD  
 101 WEPVKEAAKK LAFVEDRTIF EGYSAASIEG IRSASSNPAL TLPEDPREIP  
 151 DVISQALSEL RLAGVDGPYS VLLSADVYTK VSETSDHGYP IREHLNRLVD  
 201 GDIIWAPAID GAFVLTTRGG DFDLQLGTDV AIGYASHDTD TVRLYLQETL  
 20 251 TFLCYTAEAS VALSH

For all six proteins the molecular weights predicted from the sequences are in agreement with the molecular weights observed on SDS-PAGE.

25 Cloning of the genes encoding CFP17, CFP20, CFP21, CFP22 and CFP25.

The genes encoding CFP17, CFP20, CFP21, CFP22 and CFP25 were all cloned into the expression vector pMCT6, by PCR amplification with gene specific primers, for recombinant expression in *E. coli* of the proteins.

PCR reactions contained 10 ng of *M. tuberculosis* chromosomal DNA in 1x low salt Tag+ buffer from Stratagene supplemented with 250 mM of each of the four nucleotides (Boehringer Mannheim), 0,5 mg/ml BSA (IgG technology), 1% DMSO (Merck), 5  
 5 pmoles of each primer and 0.5 unit Tag+ DNA polymerase (Stratagene) in 10 µl reaction volume. Reactions were initially heated to 94°C for 25 sec. and run for 30 cycles according to the following program; 94°C for 10 sec., 55°C for 10 sec. and 72°C for 90 sec, using thermocycler equipment from Idaho  
 10 Technology.

The DNA fragments were subsequently run on 1% agarose gels, the bands were excised and purified by Spin-X spin columns (Costar) and cloned into pBluescript SK II+ - T vector (Stratagene). Plasmid DNA was thereafter prepared from clones  
 15 harbouring the desired fragments, digested with suitable restriction enzymes and subcloned into the expression vector pMCT6 in frame with 8 histidine residues which are added to the N-terminal of the expressed proteins. The resulting clones were hereafter sequenced by use of the dideoxy chain  
 20 termination method adapted for supercoiled DNA using the Sequenase DNA sequencing kit version 1.0 (United States Biochemical Corp., USA) and by cycle sequencing using the Dye Terminator system in combination with an automated gel reader (model 373A; Applied Biosystems) according to the instruc-  
 25 tions provided. Both strands of the DNA were sequenced.

For cloning of the individual antigens, the following gene specific primers were used:

CFP17: Primers used for cloning of cfp17:

OPBR-51: ACAGATCTGTGACGGACATGAACCCG (SEQ ID NO: 117)  
 30 OPBR-52: TTTCCATGGTCACGGGCCCCCGGTACT (SEQ ID NO: 118)

OPBR-51 and OPBR-52 create BglIII and NcoI sites, respectively, used for the cloning in pMCT6.

CFP20: Primers used for cloning of cfp20:

OPBR-53: ACAGATCTGTGCCCATGGCACAGATA (SEQ ID NO: 119)

OPBR-54: TTTAAGCTTCTAGGCGCCAGCGCGGC (SEQ ID NO: 120)

OPBR-53 and OPBR-54 create BglIII and HindIII sites, respectively, used for the cloning in pMCT6.

CFP21: Primers used for cloning of cfp21:

OPBR-55: ACAGATCTGCGCATGCGGATCCGTGT (SEQ ID NO: 121)

OPBR-56: TTTTCCATGGTCATCCGGCGTGATCGAG (SEQ ID NO: 122)

OPBR-55 and OPBR-56 create BglIII and NcoI sites, respectively, used for the cloning in pMCT6.

CFP22: Primers used for cloning of cfp22:

OPBR-57: ACAGATCTGTAATGGCAGACTGTGAT (SEQ ID NO: 123)

OPBR-58: TTTTCCATGGTCAGGAGATGGTGATCGA (SEQ ID NO: 124)

OPBR-57 and OPBR-58 create BglIII and NcoI sites, respectively, used for the cloning in pMCT6.

CFP25: Primers used for cloning of cfp25:

OPBR-59: ACAGATCTGCCGGCTACCCCGGTGCC (SEQ ID NO: 125)

OPBR-60: TTTTCCATGGCTATTGCAGCTTTCCGGC (SEQ ID NO: 126)

OPBR-59 and OPBR-60 create BglIII and NcoI sites, respectively, used for the cloning in pMCT6.

Expression/purification of recombinant CFP17, CFP20, CFP21, CFP22 and CFP25 proteins.

Expression and metal affinity purification of recombinant proteins was undertaken essentially as described by the manufacturers. For each protein, 1 l LB-media containing 100

$\mu$ g/ml ampicillin, was inoculated with 10 ml of an overnight culture of XL1-Blue cells harbouring recombinant pMCT6 plasmids. Cultures were shaken at 37 °C until they reached a density of  $OD_{600} = 0.4 - 0.6$ . IPTG was hereafter added to a  
5 final concentration of 1 mM and the cultures were further incubated 4 - 16 hours. Cells were harvested, resuspended in 1X sonication buffer + 8 M urea and sonicated 5 X 30 sec. with 30 sec. pausing between the pulses.  
After centrifugation, the lysate was applied to a column  
10 containing 25 ml of resuspended Talon resin (Clontech, Palo Alto, USA). The column was washed and eluted as described by the manufacturers.

After elution, all fractions (1.5 ml each) were subjected to analysis by SDS-PAGE using the Mighty Small (Hoefer Scientific Instruments, USA) system and the protein concentrations  
15 were estimated at 280 nm. Fractions containing recombinant protein were pooled and dialysed against 3 M urea in 10 mM Tris-HCl, pH 8.5. The dialysed protein was further purified by FPLC (Pharmacia, Sweden) using a 6 ml Resource-Q column,  
20 eluted with a linear 0-1 M gradient of NaCl. Fractions were analyzed by SDS-PAGE and protein concentrations were estimated at  $OD_{280}$ . Fractions containing protein were pooled and dialysed against 25 mM Hepes buffer, pH 8.5.

Finally the protein concentration and the LPS content were  
25 determined by the BCA (Pierce, Holland) and LAL (Endosafe, Charleston, USA) tests, respectively.

## EXAMPLE 3A

*Identification of CFP7A, CFP8A, CFP8B, CFP16, CFP19, CFP19B, CFP22A, CFP23A, CFP23B, CFP25A, CFP27, CFP30A, CWP32 and CFP50.*

5 Identification of CFP16 and CFP19B.

ST-CF was precipitated with ammonium sulphate at 80% saturation. The precipitated proteins were removed by centrifugation and after resuspension washed with 8 M urea. CHAPS and glycerol were added to a final concentration of 0.5  
 10 % (w/v) and 5 % (v/v) respectively and the protein solution was applied to a Rotofor isoelectrical Cell (BioRad). The Rotofor Cell had been equilibrated with a 8M urea buffer containing 0.5 % (w/v) CHAPS, 5% (v/v) glycerol, 3% (v/v) Biolyt 3/5 and 1% (v/v) Biolyt 4/6 (BioRad). Isoelectric  
 15 focusing was performed in a pH gradient from 3-6. The fractions were analyzed on silver-stained 10-20% SDS-PAGE. Fractions with similar band patterns were pooled and washed three times with PBS on a Centriprep concentrator (Amicon) with a 3 kDa cut off membrane to a final volume of 1-3 ml. An equal  
 20 volume of SDS containing sample buffer was added and the protein solution boiled for 5 min before further separation on a Prep Cell (BioRad) in a matrix of 16% polyacrylamide under an electrical gradient. Fractions containing well separated bands in SDS-PAGE were selected for N-terminal  
 25 sequencing after transfer to PVDF membrane.

Isolation of CFP8A, CFP8B, CFP19, CFP23A, and CFP23B.

ST-CF was precipitated with ammonium sulphate at 80% saturation and redissolved in PBS, pH 7.4, and dialysed 3 times against 25mM Piperazin-HCl, pH 5.5, and subjected to chroma-  
 30 tofocusing on a matrix of PBE 94 (Pharmacia) in a column connected to an FPLC system (Pharmacia). The column was equilibrated with 25 mM Piperazin-HCl, pH 5.5, and the elution was performed with 10% PB74-HCl, pH 4.0 (Pharmacia).

Fractions with similar band patterns were pooled and washed three times with PBS on a Centriprep concentrator (Amicon) with a 3 kDa cut off membrane to a final volume of 1-3 ml and separated on a Prepcell as described above.

##### 5 Identification of CFP22A

ST-CF was concentrated approximately 10 fold by ultrafiltration and proteins were precipitated at 80 % saturation, redissolved in PBS, pH 7.4, and dialysed 3 times against PBS, pH 7.4. 5.1 ml of the dialysed ST-CF was treated  
 10 with RNase (0.2 mg/ml, QUIAGEN) and DNase (0.2 mg/ml, Boehringer Mannheim) for 6 h and placed on top of 6.4 ml of 48 % (w/v) sucrose in PBS, pH 7.4, in Sorvall tubes (Ultracrimp 03987, DuPont Medical Products) and ultracentrifuged for 20 h at  $257,300 \times g_{\max}$ , 10°C. The pellet was redissolved in 200  $\mu$ l  
 15 of 25 mM Tris-192 mM glycine, 0.1 % SDS, pH 8.3.

##### Identification of CFP7A, CFP25A, CFP27, CFP30A and CFP50

For CFP27, CFP30A and CFP50 ST-CF was concentrated approximately 10 fold by ultrafiltration and ammonium sulphate precipitation in the 45 to 55 % saturation range was per-  
 20 formed. Proteins were redissolved in 50 mM sodium phosphate, 1.5 M ammonium sulphate, pH 8.5, and subjected to thiophilic adsorption chromatography on an Affi-T gel column (Kem-En-Tec). Proteins were eluted by a 1.5 to 0 M decreasing gradient of ammonium sulphate. Fractions with similar band  
 25 patterns in SDS-PAGE were pooled and anion exchange chromatography was performed on a Mono Q HR 5/5 column connected to an FPLC system (Pharmacia). The column was equilibrated with 10 mM Tris-HCl, pH 8.5, and the elution was performed with a gradient of NaCl from 0 to 1 M. Fractions containing well  
 30 separated bands in SDS-PAGE were selected.

CFP7A and CFP25A were obtained as described above except for the following modification: ST-CF was concentrated approximately 10 fold by ultrafiltration and proteins were precipi-

tated at 80 % saturation, redissolved in PBS, pH 7.4, and dialysed 3 times against PBS, pH 7.4. Ammonium sulphate was added to a concentration of 1.5 M, and ST-CF proteins were loaded on an Affi T-gel column. Elution from the Affi T-gel  
5 column and anion exchange were performed as described above.

#### Isolation of CWP32

Heat treated H37Rv was subfractionated into subcellular fractions as described in Sørensen et al 1995. The Cell wall fraction was resuspended in 8 M urea, 0.2 % (w/v) N-octyl  $\beta$ -D  
10 glucopyranoside (Sigma) and 5 % (v/v) glycerol and the protein solution was applied to a Rotofor isoelectrical Cell (BioRad) which was equilibrated with the same buffer. Isoelectric focusing was performed in a pH gradient from 3-6. The fractions were analyzed by SDS-PAGE and fractions con-  
15 taining well separated bands were pooled and subjected to N-terminal sequencing after transfer to PVDF membrane.

#### N-terminal sequencing

Fractions containing CFP7A, CFP8A, CFP8B, CFP16, CFP19, CFP19B, CFP22A, CFP23A, CFP23B, CFP27, CFP30A, CWP32, and  
20 CFP50A were blotted to PVDF membrane after Tricine SDS-PAGE (Ploug et al, 1989). The relevant bands were excised and subjected to N-terminal amino acid sequence analysis on a Procise 494 sequencer (Applied Biosystems). The fraction containing CFP25A was blotted to PVDF membrane after 2-DE  
25 PAGE (isoelectric focusing in the first dimension and Tricin SDS-PAGE in the second dimension). The relevant spot was excised and sequenced as described above.

The following N-terminal sequences were obtained:

CFP7A:	AEDVRAEIVA SVLEVVVNEG DQIDKGDVVV LLESMYMEIP	
30	VLAEAAGTVS	(SEQ ID NO: 81)
CFP8A:	DPVDDAFIAKLNTAG	(SEQ ID NO: 73)
CFP8B:	DPVDAIINLDNYGX	(SEQ ID NO: 74)

	CFP16:	AKLSTDELLDAFKEM	(SEQ ID NO: 79)
	CFP19:	TTSPDPYAALPKLPS	(SEQ ID NO: 82)
	CFP19B:	DPAXAPDVPTAAQLT	(SEQ ID NO: 80)
	CFP22A:	TEYEGPKTKF HALMQ	(SEQ ID NO: 83)
5	CFP23A:	VIQ/AGMVT/GHIHXVAG	(SEQ ID NO: 76)
	CFP23B:	AEMKXFKNAIVQEID	(SEQ ID NO: 75)
	CFP25A:	AIEVSVLRVF TDSDG	(SEQ ID NO: 78)
	CWP32:	TNIVVLIKQVPDTWS	(SEQ ID NO: 77)
	CFP27:	TTIVALKYPG GVVMA	(SEQ ID NO: 84)
10	CFP30A:	SFPYFISPEX AMRE	(SEQ ID NO: 85)
	CFP50:	THYDVVVLGA GPGGY	(SEQ ID NO: 86)

N-terminal homology searching in the Sanger database and  
identification of the corresponding genes.

The N-terminal amino acid sequence from each of the proteins  
15 was used for a homology search using the blast program of the  
Sanger *Mycobacterium tuberculosis* database:

<http://www.sanger.ac.uk/projects/m-tuberculosis/TB-blast-server>.

For CFP23B, CFP23A, and CFP19B no similarities were found in  
the Sanger database. This could be due to the fact that only  
20 approximately 70% of the *M. tuberculosis* genome had been  
sequenced when the searches were performed. The genes en-  
coding these proteins could be contained in the remaining 30%  
of the genome for which no sequence data is yet available.

For CFP7A, CFP8A, CFP8B, CFP16, CFP19, CFP19B , CFP22A,  
25 CFP25A, CFP27, CFP30A, CWP32, and CFP50, the following infor-  
mation was obtained:

CFP7A: Of the 50 determined amino acids in CFP7A a 98% iden-  
tical sequence was found in cosmid csCY07D1 (contig 256):  
Score = 226 (100.4 bits), Expect = 1.4e-24, P = 1.4e-24  
30 Identities = 49/50 (98%), Positives = 49/50 (98%), Frame = -1



Query: 1 AEDVRAEIVASVLEVVVNEGDQIDKGDVVVLLESMYMEIPVLAAEAGTVS 50  
 AEDVRAEIVASVLEVVVNEGDQIDKGDVVVLLESM MEIPVLAAEAGTVS  
 Sbjct: 257679 AEDVRAEIVASVLEVVVNEGDQIDKGDVVVLLESMKMEIPVLAAEAGTVS 257530

(SEQ ID NOs: 127, 128, and 129)

- 5 The identity is found within an open reading frame of 71 amino acids length corresponding to a theoretical MW of CFP7A of 7305.9 Da and a pI of 3.762. The observed molecular weight in an SDS-PAGE gel is 7 kDa.

CFP8A: A sequence 80% identical to the 15 N-terminal amino acids was found on contig TB\_1884. The N-terminally determined sequence from the protein purified from culture filtrate starts at amino acid 32. This gives a length of the mature protein of 98 amino acids corresponding to a theoretical MW of 9700 Da and a pI of 3.72 This is in good agreement with the observed MW on SDS-PAGE at approximately 8 kDa. The full length protein has a theoretical MW of 12989 Da and a pI of 4.38.

CFP8B: A sequence 71% identical to the 14 N-terminal amino acids was found on contig TB\_653. However, careful re-evaluation of the original N-terminal sequence data confirmed the identification of the protein. The N-terminally determined sequence from the protein purified from culture filtrate starts at amino acid 29. This gives a length of the mature protein of 82 amino acids corresponding to a theoretical MW of 8337 Da and a pI of 4.23. This is in good agreement with the observed MW on SDS-PAGE at approximately 8 kDa. Analysis of the amino acid sequence predicts the presence of a signal peptide which has been cleaved of the mature protein found in culture filtrate.

- 30 CFP16: The 15 aa N-terminal sequence was found to be 100% identical to a sequence found on cosmid MTCY20H1.

The identity is found within an open reading frame of 130 amino acids length corresponding to a theoretical MW of CFP16 of 13440.4 Da and a pI of 4.59. The observed molecular weight in an SDS-PAGE gel is 16 kDa.

- 5 CFP19: The 15 aa N-terminal sequence was found to be 100% identical to a sequence found on cosmid MTCY270.

The identity is found within an open reading frame of 176 amino acids length corresponding to a theoretical MW of CFP19 of 18633.9 Da and a pI of 5.41. The observed molecular weight  
10 in an SDS-PAGE gel is 19 kDa.

CFP22A: The 15 aa N-terminal sequence was found to be 100% identical to a sequence found on cosmid MTCY1A6.

The identity is found within an open reading frame of 181 amino acids length corresponding to a theoretical MW of  
15 CFP22A of 20441.9 Da and a pI of 4.73. The observed molecular weight in an SDS-PAGE gel is 22 kDa.

CFP25A: The 15 aa N-terminal sequence was found to be 100% identical to a sequence found on contig 255.

The identity is found within an open reading frame of 228  
20 amino acids length corresponding to a theoretical MW of CFP25A of 24574.3 Da and a pI of 4.95. The observed molecular weight in an SDS-PAGE gel is 25 kDa.

CFP27: The 15 aa N-terminal sequence was found to be 100% identical to a sequence found on cosmid MTCY261.

- 25 The identity is found within an open reading frame of 291 amino acids length. The N-terminally determined sequence from the protein purified from culture filtrate starts at amino acid 58. This gives a length of the mature protein of 233 amino acids, which corresponds to a theoretical molecular

weigh at 24422.4 Da, and a theoretical pI at 4.64. The observed weight in an SDS-PAGE gel is 27 kDa.

CFP30A: Of the 13 determined amino acids in CFP30A, a 100% identical sequence was found on cosmid MTCY261.

- 5 The identity is found within an open reading frame of 248 amino acids length corresponding to a theoretical MW of CFP30A of 26881.0 Da and a pI of 5.41. The observed molecular weight in an SDS-PAGE gel is 30 kDa.

- 10 CWP32: The 15 amino acid N-terminal sequence was found to be 100% identical to a sequence found on contig 281. The identity was found within an open reading frame of 266 amino acids length, corresponding to a theoretical MW of CWP32 of 28083 Da and a pI of 4.563. The observed molecular weight in an SDS-PAGE gel is 32 kDa.

- 15 CFP50: The 15 aa N-terminal sequence was found to be 100% identical to a sequence found in MTVO38.06. The identity is found within an open reading frame of 464 amino acids length corresponding to a theoretical MW of CFP50 of 49244 Da and a pI of 5.66. The observed molecular weight in an SDS-PAGE gel is 50 kDa.

Use of homology searching in the EMBL database for identification of CFP19A and CFP23.

- Homology searching in the EMBL database (using the GCG package of the Biobase, Århus-DK) with the amino acid sequences of two earlier identified highly immunoreactive ST-CF proteins, using the TFASTA algorithm, revealed that these proteins (CFP21 and CFP25, EXAMPLE 3) belong to a family of fungal cutinase homologs. Among the most homologous sequences were also two *Mycobacterium tuberculosis* sequences found on cosmid MTCY13E12. The first, MTCY13E12.04 has 46% and 50% identity to CFP25 and CFP21 respectively. The second, MTCY13E12.05, has also 46% and 50% identity to CFP25 and

CFP21. The two proteins share 62.5% aa identity in a 184 residues overlap. On the basis of the high homology to the strong T-cell antigens CFP21 and CFP25, respectively, it is believed that CFP19A and CFP23 are possible new T-cell  
 5 antigens.

The first reading frame encodes a 254 amino acid protein of which the first 26 aa constitute a putative leader peptide that strongly indicates an extracellular location of the protein. The mature protein is thus 228 aa in length corresponding to a theoretical MW of 23149.0 Da and a Pi of 5.80.  
 10 The protein is named CFP23.

The second reading frame encodes an 231 aa protein of which the first 44 aa constitute a putative leader peptide that strongly indicates an extracellular location of the protein.  
 15 The mature protein is thus 187 aa in length corresponding to a theoretical MW of 19020.3 Da and a Pi of 7.03. The protein is named CFP19A.

The presence of putative leader peptides in both proteins (and thereby their presence in the ST-CF) is confirmed by  
 20 theoretical sequence analysis using the signalP program at the Expasy molecular Biology server

(<http://expasy.hcuge.ch/www/tools.html>).

Searching for homologies to CFP7A, CFP16, CFP19, CFP19A, CFP19B, CFP22A, CFP23, CFP25A, CFP27, CFP30A, CWP32 and CFP50  
 25 in the EMBL database.

The amino acid sequences derived from the translated genes of the individual antigens were used for homology searching in the EMBL and Genbank databases using the TFASTA algorithm, in order to find homologous proteins and to address eventual  
 30 functional roles of the antigens.

CFP7A: CFP7A has 44% identity and 70% similarity to hypothetical *Methanococcus jannaschii* protein (*M. jannaschii* from base 1162199-1175341), as well as 43% and 38% identity and 68 and 64% similarity to the C-terminal part of *B. stearothermophilus* pyruvate carboxylase and *Streptococcus mutans* biotin carboxyl carrier protein.

CFP7A contains a consensus sequence EAMKM for a biotin binding site motif which in this case was slightly modified (ESMKM in amino acid residues 34 to 38). By incubation with alkaline phosphatase conjugated streptavidin after SDS-PAGE and transfer to nitrocellulose it was demonstrated that native CFP7A was biotinylated.

CFP16: RplL gene, 130 aa. Identical to the *M. bovis* 50s ribosomal protein L7/L12 (acc. No P37381).

15 CFP19: CFP19 has 47% identity and 55% similarity to *E.coli* pectinesterase homolog (ybhC gene) in a 150 aa overlap.

CFP19A: CFP19A has between 38% and 45% identity to several cutinases from different fungal sp.

In addition CFP19A has 46% identity and 61% similarity to CFP25 as well as 50% identity and 64% similarity to CFP21 (both proteins are earlier isolated from the ST-CF).

CFP19B: No apparent homology

CFP22A: No apparent homology

25 CFP23: CFP23 has between 38% and 46% identity to several cutinases from different fungal sp.

In addition CFP23 has 46% identity and 61% similarity to CFP25 as well as 50% identity and 63% similarity to CFP21 (both proteins are earlier isolated from the ST-CF).

CFP25A: CFP25A has 95% identity in a 241 aa overlap to a putative *M. tuberculosis* thymidylate synthase (450 aa accession No p28176).

CFP27: CFP27 has 81% identity to a hypothetical *M. leprae* protein and 64% identity and 78% similarity to *Rhodococcus* sp. proteasome beta-type subunit 2 (prcB(2) gene).

CFP30A: CFP30A has 67% identity to *Rhodococcus* proteasome alfa-type 1 subunit.

CWP32: The CWP32 N-terminal sequence is 100% identical to the *Mycobacterium leprae* sequence MLCB637.03.

CFP50: The CFP50 N-terminal sequence is 100% identical to a putative lipoamide dehydrogenase from *M. leprae* (Accession 415183)

Cloning of the genes encoding CFP7A, CFP8A, CFP8B, CFP16, CFP19, CFP19A, CFP22A, CFP23, CFP25A, CFP27, CFP30A, CWP32, and CFP50.

The genes encoding CFP7A, CFP8A, CFP8B, CFP16, CFP19, CFP19A, CFP22A, CFP23, CFP25A, CFP27, CFP30A, CWP32 and CFP50 were all cloned into the expression vector pMCT6, by PCR amplification with gene specific primers, for recombinant expression in *E. coli* of the proteins.

PCR reactions contained 10 ng of *M. tuberculosis* chromosomal DNA in 1X low salt Taq+ buffer from Stratagene supplemented with 250 mM of each of the four nucleotides (Boehringer Mannheim), 0,5 mg/ml BSA (IgG technology), 1% DMSO (Merck), 5 pmoles of each primer and 0.5 unit Taq+ DNA polymerase (Stratagene) in 10 ml reaction volume. Reactions were initially heated to 94°C for 25 sec. and run for 30 cycles of the program; 94°C for 10 sec., 55°C for 10 sec. and 72°C for 90 sec, using thermocycler equipment from Idaho Technology.

The DNA fragments were subsequently run on 1% agarose gels, the bands were excised and purified by Spin-X spin columns (Costar) and cloned into pBluescript SK II+ - T vector (Stratagene). Plasmid DNA was hereafter prepared from clones

5 harbouring the desired fragments, digested with suitable restriction enzymes and subcloned into the expression vector pMCT6 in frame with 8 histidines which are added to the N-terminal of the expressed proteins. The resulting clones were hereafter sequenced by use of the dideoxy chain termination

10 method adapted for supercoiled DNA using the Sequenase DNA sequencing kit version 1.0 (United States Biochemical Corp., USA) and by cycle sequencing using the Dye Terminator system in combination with an automated gel reader (model 373A; Applied Biosystems) according to the instructions provided.

15 Both strands of the DNA were sequenced.

For cloning of the individual antigens, the following gene specific primers were used:

CFP7A: Primers used for cloning of *cfp7A*:

	OPBR-79:	AAGAGTAGATCTATGATGGCCGAGGATGTTTCGG	(SEQ ID NO: 95)
20	OPBR-80:	CGGCGACGACGGATCCTACCGCGTCGG	(SEQ ID NO: 96)

OPBR-79 and OPBR-80 create *Bgl*III and *Bam*HI sites, respectively, used for the cloning in pMCT6.

CFP8A: Primers used for cloning of *cfp8A*:

	CFP8A-F:	CTGAGATCTATGAACCTACGGCGCC	(SEQ ID NO: 154)
25	CFP8A-R:	CTCCCATGGTACCCTAGGACCCGGGCAGCCCCGGC	(SEQ ID NO: 155)

CFP8A-F and CFP8A-R create *Bgl*III and *Nco*I sites, respectively, used for the cloning in pMCT6.

CFP8B: Primers used for cloning of *cfp8B*:

	CFP8B-F:	CTGAGATCTATGAGGCTGTCGTTGACCGC	(SEQ ID NO: 156)
30	CFP8B-R:	CTCCCCGGGCTTAATAGTTGTTGCAGGAGC	(SEQ ID NO: 157)

CFP8B-F and CFP8B-R create *Bgl*III and *Sma*I sites, respectively, used for the cloning in pMCT6.

CFP16: Primers used for cloning of *cfp16*:

OPBR-104: CCGGGAGATCTATGGCAAAGCTCTCCACCGACG (SEQ ID NOs: 111 and 130)  
 5 OPBR-105: CGCTGGGCAGAGCTACTTGACGGTGACGGTGG (SEQ ID NOs: 112 and 131)

OPBR-104 and OPBR-105 create *Bgl*III and *Nco*I sites, respectively, used for the cloning in pMCT6.

CFP19: Primers used for cloning of *cfp19*:

OPBR-96: GAGGAAGATCTATGACAACTTCACCCGACCCG (SEQ ID NO: 107)  
 10 OPBR-97: CATGAAGCCATGGCCCGCAGGCTGCATG (SEQ ID NO: 108)

OPBR-96 and OPBR-97 create *Bgl*III and *Nco*I sites, respectively, used for the cloning in pMCT6.

CFP19A: Primers used for cloning of *cfp19A*:

OPBR-88: CCCCCAGATCTGCACCACCGGCATCGGCGGGC (SEQ ID NO: 99)  
 15 OPBR-89: GCGGCGGATCCGTTGCTTAGCCGG (SEQ ID NO: 100)

OPBR-88 and OPBR-89 create *Bgl*III and *Bam*HI sites, respectively, used for the cloning in pMCT6.

CFP22A: Primers used for cloning of *cfp22A*:

OPBR-90: CCGGCTGAGATCTATGACAGAATACGAAGGGC (SEQ ID NO: 101)  
 20 OPBR-91: CCCC GCCAGGGA ACTAGAGGCGGC (SEQ ID NO: 102)

OPBR-90 and OPBR-91 create *Bgl*III and *Nco*I sites, respectively, used for the cloning in pMCT6.

CFP23: Primers used for cloning of *cfp23*:

OPBR-86: CCTTGGGAGATCTTTGGACCCCGGTTGC (SEQ ID NO: 97)  
 25 OPBR-87: GACGAGATCTTATGGGCTTACTGAC (SEQ ID NO: 98)



OPBR-86 and OPBR-87 both create a *Bgl*III site used for the cloning in pMCT6.

CFP25A: Primers used for cloning of *cfp25A*:

OPBR-106: GGCCCAGATCTATGGCCATTGAGGTTTCGGTGTTC (SEQ ID NO: 113)  
 5 OPBR-107: CGCCGTGTTGCATGGCAGCGCTGAGC (SEQ ID NO: 114)

OPBR-106 and OPBR-107 create *Bgl*III and *Nco*I sites, respectively, used for the cloning in pMCT6.

CFP27: Primers used for cloning of *cfp27*:

OPBR-92: CTGCCGAGATCTACCACCATTGTGCGCGCTGAAATACCC (SEQ ID NO: 103)  
 10 OPBR-93: CGCCATGGCCTTACGCGCCAACCTCG (SEQ ID NO: 104)

OPBR-92 and OPBR-93 create *Bgl*III and *Nco*I sites, respectively, used for the cloning in pMCT6.

CFP30A: Primers used for cloning of *cfp30A*:

OPBR-94: GGCGGAGATCTGTGAGTTTTCGTAATTCATC (SEQ ID NO: 105)  
 15 OPBR-95: CGCGTCGAGCCATGGTTAGGCGCAG (SEQ ID NO: 106)

OPBR-94 and OPBR-95 create *Bgl*III and *Nco*I sites, respectively, used for the cloning in pMCT6.

CWP32: Primers used for cloning of *cwp32*:

CWP32-F: GCTTAGATCTATGATTTTCTGGGCAACCAGGTA (SEQ ID NO: 158)  
 20 CWP32-R: GCTTCCATGGGCGAGGCACAGGCGTGGGAA (SEQ ID NO: 159)

CWP32-F and CWP32-R create *Bgl*III and *Nco*I sites, respectively, used for the cloning in pMCT6.

CFP50: Primers used for cloning of *cfp50*:

OPBR-100: GGCCGAGATCTGTGACCCACTATGACGTCGTCG (SEQ ID NO: 109)  
 25 OPBR-101: GGCGCCCATGGTCAGAAATGATCATGTGGCCAA (SEQ ID NO: 110)

OPBR-100 and OPBR-101 create *Bgl*III and *Nco*I sites, respectively, used for the cloning in pMCT6.

Expression/purification of recombinant CFP7A, CFP8A, CFP8B, CFP16, CFP19, CFP19A, CFP22A, CFP23, CFP25A, CFP27, CFP30A, CWP32, and CFP50 proteins.

Expression and metal affinity purification of recombinant proteins was undertaken essentially as described by the manufacturers. For each protein, 1 l LB-media containing 100  $\mu$ g/ml ampicillin, was inoculated with 10 ml of an overnight  
10 culture of XL1-Blue cells harbouring recombinant pMCT6 plasmids. Cultures were shaken at 37°C until they reached a density of  $OD_{600} = 0.4 - 0.6$ . IPTG was hereafter added to a final concentration of 1 mM and the cultures were further incubated 4-16 hours. Cells were harvested, resuspended in 1X  
15 sonication buffer + 8 M urea and sonicated 5 X 30 sec. with 30 sec. pausing between the pulses.

After centrifugation, the lysate was applied to a column containing 25 ml of resuspended Talon resin (Clontech, Palo Alto, USA). The column was washed and eluted as described by  
20 the manufacturers.

After elution, all fractions (1.5 ml each) were subjected to analysis by SDS-PAGE using the Mighty Small (Hoefer Scientific Instruments, USA) system and the protein concentrations were estimated at 280 nm. Fractions containing recombinant  
25 protein were pooled and dialysed against 3 M urea in 10 mM Tris-HCl, pH 8.5. The dialysed protein was further purified by FPLC (Pharmacia, Sweden) using a 6 ml Resource-Q column, eluted with a linear 0-1 M gradient of NaCl. Fractions were analyzed by SDS-PAGE and protein concentrations were estimated at  $OD_{280}$ . Fractions containing protein were pooled and  
30 dialysed against 25 mM Hepes buffer, pH 8.5.

Finally the protein concentration and the LPS content were determined by the BCA (Pierce, Holland) and LAL (Endosafe, Charleston, USA) tests, respectively.

#### EXAMPLE 3B

##### 5 *Identification of CFP7B, CFP10A, CFP11 and CFP30B.*

##### Isolation of CFP7B

ST-CF was precipitated with ammonium sulphate at 80% saturation and redissolved in PBS, pH 7.4, and dialyzed 3 times against 25 mM Piperazin-HCl, pH 5.5, and subjected to chromatofocusing on a matrix of PBE 94 (Pharmacia) in a column connected to an FPLC system (Pharmacia). The column was equilibrated with 25 mM Piperazin-HCl, pH 5.5, and the elution was performed with 10% PB74-HCl, pH 4.0 (Pharmacia). Fractions with similar band patterns were pooled and washed three times with PBS on a Centriprep concentrator (Amicon) with a 3 kDa cut off membrane to a final volume of 1-3 ml. An equal volume of SDS containing sample buffer was added and the protein solution boiled for 5 min before further separation on a MultiEluter (BioRad) in a matrix of 10-20 % polyacrylamid (Andersen, P. & Heron, I., 1993). The fraction containing a well separated band below 10 kDa was selected for N-terminal sequencing after transfer to a PVDF membrane.

##### Isolation of CFP11

ST-CF was precipitated with ammonium sulphate at 80% saturation. The precipitated proteins were removed by centrifugation and after resuspension washed with 8 M urea. CHAPS and glycerol were added to a final concentration of 0.5 % (w/v) and 5% (v/v) respectively and the protein solution was applied to a Rotofor isoelectrical Cell (BioRad). The Rotofor Cell had been equilibrated with an 8M urea buffer containing 0.5 % (w/v) CHAPS, 5% (v/v) glycerol, 3% (v/v) Biolyt 3/5 and

1% (v/v) Biolyt 4/6 (BioRad). Isoelectric focusing was performed in a pH gradient from 3-6. The fractions were analyzed on silver-stained 10-20% SDS-PAGE. The fractions in the pH gradient 5.5 to 6 were pooled and washed three times with PBS  
 5 on a Centriprep concentrator (Amicon) with a 3 kDa cut off membrane to a final volume of 1 ml. 300 mg of the protein preparation was separated on a 10-20% Tricine SDS-PAGE (Ploug et al 1989) and transferred to a PVDF membrane and Coomassie stained. The lowest band occurring on the membrane  
 10 was excised and submitted for N-terminal sequencing.

#### Isolation of CFP10A and CFP30B

ST-CF was concentrated approximately 10-fold by ultrafiltration and ammonium sulphate precipitation at 80 % saturation. Proteins were redissolved in 50 mM sodium phosphate, 1.5 M  
 15 ammonium sulphate, pH 8.5, and subjected to thiophilic adsorption chromatography on an Affi-T gel column (Kem-En-Tec). Proteins were eluted by a 1.5 to 0 M decreasing gradient of ammonium sulphate. Fractions with similar band patterns in SDS-PAGE were pooled and anion exchange chromatography was  
 20 performed on a Mono Q HR 5/5 column connected to an FPLC system (Pharmacia). The column was equilibrated with 10 mM Tris-HCl, pH 8.5, and the elution was performed with a gradient of NaCl from 0 to 1 M. Fractions containing well separated bands in SDS-PAGE were selected.

25 Fractions containing CFP10A and CFP30B were blotted to PVDF membrane after 2-DE-PAGE (Ploug et al, 1989). The relevant spots were excised and subjected to N-terminal amino acid sequence analysis.

#### N-terminal sequencing

30 N-terminal amino acid sequence analysis was performed on a Procise 494 sequencer (Applied Biosystems).

The following N-terminal sequences were obtained:

CFP7B:	PQGTVKWFNAEKGFG	(SEQ ID NO: 168)
CFP10A:	NVTVSIPTILRPXXX	(SEQ ID NO: 169)
CFP11:	TRFMTDPHAMRDMAG	(SEQ ID NO: 170)
CFP30B:	PKRSEYRQGTPNWVD	(SEQ ID NO: 171)

- 5 "X" denotes an amino acid which could not be determined by the sequencing method used.

N-terminal homology searching in the Sanger database and identification of the corresponding genes.

- 10 The N-terminal amino acid sequence from each of the proteins was used for a homology search using the blast program of the Sanger *Mycobacterium tuberculosis* genome database:

<http://www.sanger.ac.uk/projects/m-tuberculosis/TB-blast-server>.

- 15 For CFP11 a sequence 100% identical to 15 N-terminal amino acids was found on contig TB\_1314. The identity was found within an open reading frame of 98 amino acids length corresponding to a theoretical MW of 10977 Da and a pI of 5.14.

- Amino acid number one can also be an Ala (insted of a Thr) as this sequence was also obtained (results not shown), and a 100% identical sequence to this N-terminal is found on contig  
20 TB\_671 and on locus MTCI364.09.

- For CFP7B a sequence 100% identical to 15 N-terminal amino acids was found on contig TB\_2044 and on locus MTY15C10.04 with EMBL accession number: z95436. The identity was found within an open reading frame of 67 amino acids length corresponding to a theoretical MW of 7240 Da and a pI of 5.18.  
25

For CFP10A a sequence 100% identical to 12 N-terminal amino acids was found on contig TB\_752 and on locus CY130.20 with EMBL accession number: Q10646 and Z73902. The identity was found within an open reading frame of 93 amino acids length

corresponding to a theoretical MW of 9557 Da and a pI of 4.78.

For CFP30B a sequence 100% identical to 15 N-terminal amino acids was found on contig TB\_335. The identity was found  
5 within an open reading frame of 261 amino acids length corresponding to a theoretical MW of 27345 Da and a pI of 4.24.

The amino acid sequences of the purified antigens as picked from the Sanger database are shown in the following list.

10 CFP7B (SEQ ID NO: 147)

1 MPQGTVKWFN AEKGFGFIAP EDGSADV FVH YTEIQGTGFR TLEENQKVEF  
51 EIGHSPKGPQ ATGVRSL

CFP10A (SEQ ID NO: 141)

1 MNVTVSIPTI LRPHTGGQKS VSASGDTLGA VISDLEANYS GISERLMDPS  
15 51 SPGKLHRFVN IYVNDEDVRF SGGLATAIAD GDSVTILPAV AGG

CFP11 protein sequence (SEQ ID NO: 143)

1 MATRFMTDPH AMRDMAGRFE VHAQTVEDEA RRMWASAQNI SGAGWSGMAE  
51 ATSLDTMAQM NQAFRNIVNM LHGVRDGLVR DANNYEQQEQ ASQQILSS

CFP30B (SEQ ID NO: 145)

20 1 MPKRSEYRQG TPNWVDLQTT DQSAAKKFYT SLFGWGYDDN PVPGGGGVYS  
51 MATLNGEAVA AIAPMPPGAP EGMPPIWNTY IAVDDVDAVV DKVVPGGGQV  
101 MMPAFDIGDA GRMSFITDPT GAAVGLWQAN RHIGATLVNE TGTLIWNELL  
151 TDKPDLALAF YEAVVGLTHS SMEIAAGQNY RVLKAGDAEV GGCMEPPMPG  
201 VPNHWHVYFA VDDADATAAK AAAAGGQVIA EPADIPSVGR FAVLSDPQGA  
25 251 IFSVLKPAPQ Q

Cloning of the genes encoding CFP7B, CFP10A, CFP11, and CFP30B.

PCR reactions contained 10 ng of *M. tuberculosis* chromosomal DNA in 1X low salt Taq+ buffer from Stratagene supplemented  
 5 with 250 mM of each of the four nucleotides (Boehringer Mannheim), 0,5 mg/ml BSA (IgG technology), 1% DMSO (Merck), 5 pmoles of each primer and 0.5 unit Tag+ DNA polymerase (Stratagene) in 10 ml reaction volume. Reactions were initially heated to 94°C for 25 sec. and run for 30 cycles of the  
 10 program; 94°C for 10 sec., 55°C for 10 sec. and 72°C for 90 sec., using thermocycler equipment from Idaho Technology.

The DNA fragments were subsequently run on 1% agarose gels, the bands were excised and purified by Spin-X spin columns (Costar) and cloned into pBluscript SK II+ - T vector (Stratagene). Plasmid DNA was hereafter prepared from clones  
 15 harbouring the desired fragments, digested with suitable restriction enzymes and subcloned into the expression vector pMCT6 in frame with 8 histidines which are added to the N-terminal of the expressed proteins. The resulting clones were  
 20 hereafter sequenced by use of the dideoxy chain termination method adapted for supercoiled DNA using the Sequenase DNA sequencing kit version 1.0 (United States Biochemical Corp., USA) and by cycle sequencing using the Dye Terminator system in combination with an automated gel reader (model 373A;  
 25 Applied Biosystems) according to the instructions provided. Both strands of the DNA were sequenced.

For cloning of the individual antigens, the following gene specific primers were used:

CFP7B: Primers used for cloning of *cfp7B*:

30	CFP7B-F:	CTGAGATCTAGAATGCCACAGGGAAGTGTG	(SEQ ID NO: 160)
	CFP7B-R:	TCTCCCGGGGGTAACTCAGAGCGAGCGGAC	(SEQ ID NO: 161)

CFP7B-F and CFP7B-R create *Bgl*II and *Sma*I sites, respectively, used for the cloning in pMCT6.

CFP10A: Primers used for cloning of *cfp10A*:

CFP10A-F: CTGAGATCTATGAACGTCACCGTATCC (SEQ ID NO: 162)  
 CFP10A-R: TCTCCCGGGGCTCACCCACCGGCCACG (SEQ ID NO: 163)

CFP10A -F and CFP10A -R create *Bgl*III and *Sma*I sites, respectively, used for the cloning in pMCT6.

CFP11: Primers used for cloning of *cfp11*:

CFP11-F: CTGAGATCTATGGCAACACGTTTTATGACG (SEQ ID NO: 164)  
 CFP11-R: CTCCCCGGGTTAGCTGCTGAGGATCTGCTH (SEQ ID NO: 165)

CFP11-F and CFP11-R create *Bgl*III and *Sma*I sites, respectively, used for the cloning in pMCT6.

CFP30B: Primers used for cloning of *cfp30B*:

CFP30B-F: CTGAAGATCTATGCCCAAGAGAAGCGAATAC (SEQ ID NO: 166)  
 CFP30B -R: CGGCAGCTGCTAGCATTCTCCGAATCTGCCG (SEQ ID NO: 167)

CFP30B-F and CFP30B-R create *Bgl*III and *Pvu*II sites, respectively, used for the cloning in pMCT6.

Expression/purification of recombinant CFP7B, CFP10A, CFP11 and CFP30B protein.

Expression and metal affinity purification of recombinant protein was undertaken essentially as described by the manufacturers. 1 l LB-media containing 100 µg/ml ampicillin, was inoculated with 10 ml of an overnight culture of XL1-Blue cells harbouring recombinant pMCT6 plasmid. The culture was shaken at 37 °C until it reached a density of OD<sub>600</sub> = 0.5. IPTG was hereafter added to a final concentration of 1 mM and the culture was further incubated 4 hours. Cells were harvested, resuspended in 1X sonication buffer + 8 M urea and sonicated 5 X 30 sec. with 30 sec. pausing between the pulses.



After centrifugation, the lysate was applied to a column containing 25 ml of resuspended Talon resin (Clontech, Palo Alto, USA). The column was washed and eluted as described by the manufacturers.

- 5 After elution, all fractions (1.5 ml each) were subjected to analysis by SDS-PAGE using the Mighty Small (Hoefer Scientific Instruments, USA) system and the protein concentrations were estimated at 280 nm. Fractions containing recombinant protein were pooled and dialysed against 3 M urea in 10 mM
- 10 Tris-HCl, pH 8.5. The dialysed protein was further purified by FPLC (Pharmacia, Sweden) using a 6 ml Resource-Q column, eluted with a linear 0-1 M gradient of NaCl. Fractions were analysed by SDS-PAGE and protein concentrations were estimated at OD<sub>280</sub>. Fractions containing protein were pooled and
- 15 dialysed against 25 mM Hepes buffer, pH 8.5.

Finally the protein concentration and the LPS content was determined by the BCA (Pierce, Holland) and LAL (Endosafe, Charleston, USA) tests, respectively.

#### EXAMPLE 4

- 20 *Cloning of the gene expressing CFP26 (MPT51)*

##### Synthesis and design of probes

- 25 Oligonucleotide primers were synthesized automatically on a DNA synthesizer (Applied Biosystems, Forster City, Ca, ABI-391, PCR-mode) deblocked and purified by ethanol precipitation.

- 30 Three oligonucleotides were synthesized (TABLE 3) on the basis of the nucleotide sequence from *mpb51* described by Ohara et al. (1995). The oligonucleotides were engineered to include an *EcoRI* restriction enzyme site at the 5' end and at the 3' end by which a later subcloning was possible.

Additional four oligonucleotides were synthesized on the basis of the nucleotide sequence from MPT51 (Fig. 5 and SEQ ID NO: 41). The four combinations of the primers were used for the PCR studies.

## 5 DNA cloning and PCR technology

Standard procedures were used for the preparation and handling of DNA (Sambrook et al., 1989). The gene *mpt51* was cloned from *M. tuberculosis* H37Rv chromosomal DNA by the use of the polymerase chain reactions (PCR) technology as described previously (Oettinger and Andersen, 1994). The PCR product was cloned in the pBluescriptSK + (Stratagene).

### Cloning of *mpt51*

The gene, the signal sequence and the Shine Delgarno region of MPT51 was cloned by use of the PCR technology as two fragments of 952 bp and 815 bp in pBluescript SK +, designated pT052 and pT053.

### DNA Sequencing

The nucleotide sequence of the cloned 952 bp *M. tuberculosis* H37Rv PCR fragment, pT052, containing the Shine Dalgarno sequence, the signal peptide sequence and the structural gene of MPT51, and the nucleotide sequence of the cloned 815 bp PCR fragment containing the structural gene of MPT51, pT053, were determined by the dideoxy chain termination method adapted for supercoiled DNA by use of the Sequenase DNA sequencing kit version 1.0 (United States Biochemical Corp., Cleveland, OH) and by cycle sequencing using the Dye Terminator system in combination with an automated gel reader (model 373A; Applied Biosystems) according to the instructions provided. Both strands of the DNA were sequenced.

The nucleotide sequences of pT052 and pT053 and the deduced amino acid sequence are shown in Figure 5. The DNA sequence

contained an open reading frame starting with a ATG codon at position 45 - 47 and ending with a termination codon (TAA) at position 942 - 944. The nucleotide sequence of the first 33 codons was expected to encode the signal sequence. On the basis of the known N-terminal amino acid sequence (Ala - Pro - Tyr - Glu - Asn) of the purified MPT51 (Nagai et al., 1991) and the features of the signal peptide, it is presumed that the signal peptidase recognition sequence (Ala-X-Ala) (von Heijne, 1984) is located in front of the N-terminal region of the mature protein at position 144. Therefore, a structural gene encoding MPT51, *mpt51*, derived from *M. tuberculosis* H37Rv was found to be located at position 144 - 945 of the sequence shown in Fig. 5. The nucleotide sequence of *mpt51* differed with one nucleotide compared to the nucleotide sequence of MPB51 described by Ohara et al. (1995) (Fig. 5). In *mpt51* at position 780 was found a substitution of a guanine to an adenine. From the deduced amino acid sequence this change occurs at a first position of the codon giving a amino acid change from alanine to threonine. Thus it is concluded, that *mpt51* consists of 801 bp and that the deduced amino acid sequence contains 266 residues with a molecular weight of 27,842, and MPT51 show 99,8% identity to MPB51.

#### Subcloning of *mpt51*

An *EcoRI* site was engineered immediately 5' of the first codon of *mpt51* so that only the coding region of the gene encoding MPT51 would be expressed, and an *EcoRI* site was incorporated right after the stop codon at the 3' end.

DNA of the recombinant plasmid pT053 was cleaved at the *EcoRI* sites. The 815 bp fragment was purified from an agarose gel and subcloned into the *EcoRI* site of the pMAL-cR1 expression vector (New England Biolabs), pT054. Vector DNA containing the gene fusion was used to transform the *E. coli* XL1-Blue by the standard procedures for DNA manipulation.

The endpoints of the gene fusion were determined by the dideoxy chain termination method as described under section DNA sequencing. Both strands of the DNA were sequenced.

Preparation and purification of rMPT51

- 5 Recombinant antigen was prepared in accordance with instructions provided by New England Biolabs. Briefly, single colonies of *E. coli* harbouring the pT054 plasmid were inoculated into Luria-Bertani broth containing 50 µg/ml ampicillin and 12.5 µg/ml tetracycline and grown at 37°C to  $2 \times 10^8$
- 10 cells/ml. Isopropyl-β-D-thiogalactoside (IPTG) was then added to a final concentration of 0.3 mM and growth was continued for further 2 hours. The pelleted bacteria were stored overnight at -20°C in new column buffer (20 mM Tris/HCl, pH 7.4, 200 mM NaCl, 1 mM EDTA, 1 mM dithiothreitol (DTT)) and thawed
- 15 at 4°C followed by incubation with 1 mg/ml lysozyme on ice for 30 min and sonication (20 times for 10 sec with intervals of 20 sec). After centrifugation at 9,000 x g for 30 min at 4°C, the maltose binding protein -MPT51 fusion protein (MBP-rMPT51) was purified from the crude extract by affinity
- 20 chromatography on amylose resin column. MBP-rMPT51 binds to amylose. After extensive washes of the column, the fusion protein was eluted with 10 mM maltose. Aliquots of the fractions were analyzed on 10% SDS-PAGE. Fractions containing the fusion protein of interest were pooled and was dialysed
- 25 extensively against physiological saline.

Protein concentration was determined by the BCA method supplied by Pierce (Pierce Chemical Company, Rockford, IL).

TABLE 3.

Orientation and oligonucleotide <sup>a</sup>		Sequence of the <i>mpt51</i> oligonucleotides <sup>a</sup> . Sequences (5' → 3')	Position <sup>b</sup> (nucleotide)
5	Sense		
	MPT51-1	<u>CTCGAATT</u> CGCCGGGTGCACACAG (SEQ ID NO: 28)	6 - 21 (SEQ ID NO: 41)
	MPT51-3	<u>CTCGAATT</u> CGCCCCATACGAGAAC (SEQ ID NO: 29)	143 - 158 (SEQ ID NO: 41)
	MPT51-5	GTGTATCTGCTGGAC (SEQ ID NO: 30)	228 - 242 (SEQ ID NO: 41)
	MPT51-7	CCGACTGGCTGGCCG (SEQ ID NO: 31)	418 - 432 (SEQ ID NO: 41)
10	Antisense		
	MPT51-2	<u>GAGGAATT</u> CGCTTAGCGGATCGCA (SEQ ID NO: 32)	946 - 932 (SEQ ID NO: 41)
	MPT51-4	CCCACATTCGGTTGG (SEQ ID NO: 33)	642 - 628 (SEQ ID NO: 41)
	MPT51-6	GTCCAGCAGATACAC (SEQ ID NO: 34)	242 - 228 (SEQ ID NO: 41)

<sup>a</sup> The oligonucleotides MPT51-1 and MPT51-2 were constructed from the MPB51 nucleotide sequence (Ohara et al., 1995). The other oligonucleotides constructions were based on the nucleotide sequence obtained from *mpt51* reported in this work. Nucleotides (nt) underlined are not contained in the nucleotide sequence of MPB/T51.

<sup>b</sup> The positions referred to are of the non-underlined parts of the primers and correspond to the nucleotide sequence shown in SEQ ID NO: 41.

#### Cloning of *mpt51* in the expression vector pMST24.

A PCR fragment was produced from pT052 using the primer combination MPT51-F and MPT51-R (TABLE 4). A *Bam*HI site was engineered immediately 5' of the first codon of *mpt51* so that only the coding region of the gene encoding MPT51 would be expressed, and an *Nco*I site was incorporated right after the stop codon at the 3' end.

The PCR product was cleaved at the *Bam*HI and the *Nco*I site. The 811 bp fragment was purified from an agarose gel and subcloned into the *Bam*HI and the *Nco*I site of the pMST24 expression vector, pT086. Vector DNA containing the gene fusion was used to transform the *E. coli* XL1-Blue by the standard procedures for DNA manipulation.

The nucleotide sequence of complete gene fusion was determined by the dideoxy chain termination method as described under section DNA sequencing. Both strands of the DNA were sequenced.

Preparation and purification of rMPT51.

Recombinant antigen was prepared from single colonies of *E. coli* harbouring the pT086 plasmid inoculated into Luria-Bertani broth containing 50 µg/ml ampicillin and 12.5 µg/ml tetracycline and grown at 37°C to  $2 \times 10^8$  cells/ml. Isopropyl-β-D-thiogalactoside (IPTG) was then added to a final concentration of 1 mM and growth was continued for further 2 hours. The pelleted bacteria were resuspended in BC 100/20 buffer (100 mM KCl, 20 mM Imidazole, 20 mM Tris/HCl, pH 7.9, 20 % glycerol). Cells were broken by sonication (20 times for 10 sec with intervals of 20 sec). After centrifugation at 9,000 x g for 30 min. at 4°C the insoluble matter was resuspended in BC 100/20 buffer with 8 M urea followed by sonication and centrifugation as above. The His tag-MPT51 fusion protein (His-rMPT51) was purified by affinity chromatography on Ni-NTA resin column (Qiagen, Hilden, Germany). His-rMPT51 binds to Ni-NTA. After extensive washes of the column, the fusion protein was eluted with BC 100/40 buffer (100 mM KCl, 40 mM Imidazole, 20 mM Tris/HCl, pH 7.9, 20 % glycerol) with 8 M urea and BC 1000/40 buffer (1000 mM KCl, 40 mM Imidazole, 20 mM Tris/HCl, pH 7.9, 20 % glycerol) with 8 M urea. His-rMPT51 was extensively dialysed against 10 mM Tris/HCl, pH 8.5, 3 M urea followed by purification using fast protein liquid chromatography (FPLC) (Pharmacia, Uppsala, Sweden), over an anion exchange column (Mono Q) using 10 mM Tris/HCl, pH 8.5, 3 M urea with a 0 - 1 M NaCl linear gradient. Fractions containing rMPT51 were pooled and subsequently dialysed extensively against 25 mM Hepes, pH 8.0 before use.

Protein concentration was determined by the BCA method supplied by Pierce (Pierce Chemical Company, Rockford, IL). The lipopolysaccharide (LPS) content was determined by the limulus amoebocyte lysate test (LAL) to be less than 0.004 ng/µg rMPT51, and this concentration had no influence on cellular activity.

TABLE 4. Sequence of the *mpt51* oligonucleotides.

Orientation and oligonucleotide	Sequences (5' → 3')	Position (nt)
Sense		
5 MPT51-F	<u>CTCGGATCCTG</u> CCCCATACGAGAACCTG	139 - 156
Antisense		
MPT51-R	CTCCCATGGTTAGCGGATCGCACCG	939 - 924

## EXAMPLE 4A

*Cloning of the ESAT6-MPT59 and the MPT59-ESAT6 hybrids.*

10 Background for ESAT-MPT59 and MPT59-ESAT6 fusion

Several studies have demonstrated that ESAT-6 is a an immunogen which is relatively difficult to adjuvate in order to obtain consistent results when immunizing therewith. To detect an *in vitro* recognition of ESAT-6 after immunization with the antigen is very difficult compared to the strong recognition of the antigen that has been found during the recall of memory immunity to *M. tuberculosis*. ESAT-6 has been found in ST-CF in a truncated version were amino acids 1-15 have been deleted. The deletion includes the main T-cell epitopes recognized by C57BL/6j mice (Brandt et al., 1996). This result indicates that ESAT-6 either is N-terminally processed or proteolytically degraded in STCF. In order to optimize ESAT-6 as an immunogen, a gene fusion between ESAT-6 and another major T cell antigen MPT59 has been constructed. Two different construct have been made: MPT59-ESAT-6 (SEQ ID NO: 172) and ESAT-6-MPT59 (SEQ ID NO: 173). In the first hybrid ESAT-6 is N-terminally protected by MPT59 and in the latter it is expected that the fusion of two dominant T-cell antigens can have a synergistic effect.

The genes encoding the ESAT6-MPT59 and the MPT59-ESAT6 hybrids were cloned into the expression vector pMCT6, by PCR amplification with gene specific primers, for recombinant expression in *E. coli* of the hybrid proteins.

#### 5 Construction of the hybrid MPT59-ESAT6.

The cloning was carried out in three steps. First the genes encoding the two components of the hybrid, ESAT6 and MPT59, were PCR amplified using the following primer constructions:

##### ESAT6:

- 10 OPBR-4: GGCGCCGGCAAGCTTGCCATGACAGAGCAGCAGTGG (SEQ ID NO: 132)  
 OPBR-28: CGAACTCGCCGGATCCCGTGTTCGC (SEQ ID NO: 133)

OPBR-4 and OPBR-28 create HindIII and BamHI sites, respectively.

##### MPT59:

- 15 OPBR-48: GGCAACCGCGAGATCTTCTCCCGCCGGGGC (SEQ ID NO: 134)  
 OPBR-3: GGCAAGCTTGCCGGCGCCTAACGAAC (SEQ ID NO: 135)

OPBR-48 and OPBR-3 create BglII and HindIII, respectively. Additionally OPBR-3 deletes the stop codon of MPT59.

- PCR reactions contained 10 ng of *M. tuberculosis* chromosomal DNA in 1x low salt Taq+ buffer from Stratagene supplemented with 250 mM of each of the four nucleotides (Boehringer Mannheim), 0,5 mg/ml BSA (IgG technology), 1% DMSO (Merck), 5 pmoles of each primer and 0.5 unit Tag+ DNA polymerase (Stratagene) in 10 µl reaction volume. Reactions were initially heated to 94°C for 25 sec. and run for 30 cycles of the program; 94°C for 10 sec., 55°C for 10 sec. and 72°C for 90 sec, using thermocycler equipment from Idaho Technology.

The DNA fragments were subsequently run on 1% agarose gels, the bands were excised and purified by Spin-X spin columns



(Costar). The two PCR fragments were digested with HindIII and ligated. A PCR amplification of the ligated PCR fragments encoding MPT59-ESAT6 was carried out using the primers OPBR-48 and OPBR-28. PCR reaction was initially heated to 94°C for 25 sec. and run for 30 cycles of the program; 94°C for 30 sec., 55°C for 30 sec. and 72°C for 90 sec. The resulting PCR fragment was digested with BglII and BamHI and cloned into the expression vector pMCT6 in frame with 8 histidines which are added to the N-terminal of the expressed protein hybrid. The resulting clones were hereafter sequenced by use of the dideoxy chain termination method adapted for supercoiled DNA using the Sequenase DNA sequencing kit version 1.0 (United States Biochemical Corp., USA) and by cycle sequencing using the Dye Terminator system in combination with an automated gel reader (model 373A; Applied Biosystems) according to the instructions provided. Both strands of the DNA were sequenced.

#### Construction of the hybrid ESAT6-MPT59.

Construction of the hybrid ESAT6-MPT59 was carried out as described for the hybrid MPT59-ESAT6. The primers used for the construction and cloning were:

#### ESAT6:

OPBR-75: GGACCCAGATCTATGACAGAGCAGCAGTGG (SEQ ID NO: 136)

OPBR-76: CCGGCAGCCCCGGCCGGGAGAAAAGCTTTGCGAACATCCCAGTGACG (SEQ ID NO: 137)

OPBR-75 and OPBR-76 create BglII and HindIII sites, respectively. Additionally OPBR-76 deletes the stop codon of ESAT6.

#### MPT59:

OPBR-77: GTTCGCAAAGCTTTTCTCCCGCCGGGGCTGCCGGTCGAGTACC (SEQ ID NO: 138)

OPBR-18: CCTTCGGTGGATCCCGTCAG (SEQ ID NO: 139)

OPBR-77 and OPBR-18 create HindIII and BamHI sites, respectively.

Expression/purification of MPT59-ESAT6 and ESAT6-MPT59 hybrid proteins.

Expression and metal affinity purification of recombinant proteins was undertaken essentially as described by the  
5 manufacturers. For each protein, 1 l LB-media containing 100  $\mu$ g/ml ampicillin, was inoculated with 10 ml of an overnight culture of XL1-Blue cells harbouring recombinant pMCT6 plas-  
mids. Cultures were shaken at 37 °C until they reached a  
density of  $OD_{600} = 0.4 - 0.6$ . IPTG was hereafter added to a  
10 final concentration of 1 mM and the cultures were further  
incubated 4 - 16 hours. Cells were harvested, resuspended in  
1X sonication buffer + 8 M urea and sonicated 5 X 30 sec.  
with 30 sec. pausing between the pulses.

After centrifugation, the lysate was applied to a column  
15 containing 25 ml of resuspended Talon resin (Clontech, Palo  
Alto, USA). The column was washed and eluted as described by  
the manufacturers.

After elution, all fractions (1.5 ml each) were subjected to  
analysis by SDS-PAGE using the Mighty Small (Hoefer Scien-  
20 tific Instruments, USA) system and the protein concentrations  
were estimated at 280 nm. Fractions containing recombinant  
protein were pooled and dialysed against 3 M urea in 10 mM  
Tris-HCl, pH 8.5. The dialysed protein was further purified  
by FPLC (Pharmacia, Sweden) using a 6 ml Resource-Q column,  
25 eluted with a linear 0-1 M gradient of NaCl. Fractions were  
analyzed by SDS-PAGE and protein concentrations were esti-  
mated at  $OD_{280}$ . Fractions containing protein were pooled and  
dialysed against 25 mM Hepes buffer, pH 8.5.

Finally the protein concentration and the LPS content were  
30 determined by the BCA (Pierce, Holland) and LAL (Endosafe,  
Charleston, USA) tests, respectively.

The biological activity of the MPT59-ESAT6 fusion protein is  
described in Example 6A.

## EXAMPLE 5

*Mapping of the purified antigens in a 2DE system.*

In order to characterize the purified antigens they were mapped in a 2-dimensional electrophoresis (2DE) reference system. This consists of a silver stained gel containing ST-CF proteins separated by isoelectrical focusing followed by a separation according to size in a polyacrylamide gel electrophoresis. The 2DE was performed according to Hochstrasser et al. (1988). 85  $\mu$ g of ST-CF was applied to the isoelectrical focusing tubes where BioRad ampholytes BioLyt 4-6 (2 parts) and BioLyt 5-7 (3 parts) were included. The first dimension was performed in acrylamide/piperazin diacrylamide tube gels in the presence of urea, the detergent CHAPS and the reducing agent DTT at 400 V for 18 hours and 800 V for 2 hours. The second dimension 10-20% SDS-PAGE was performed at 100 V for 18 hours and silver stained. The identification of CFP7, CFP7A, CFP7B, CFP8A, CFP8B, CFP9, CFP11, CFP16, CFP17, CFP19, CFP20, CFP21, CFP22, CFP25, CFP27, CFP28, CFP29, CFP30A, CFP50, and MPT51 in the 2DE reference gel were done by comparing the spot pattern of the purified antigen with ST-CF with and without the purified antigen. By the assistance of an analytical 2DE software system (Phoretix International, UK) the spots have been identified in Fig. 6. The position of MPT51 and CFP29 were confirmed by a Western blot of the 2DE gel using the Mab's anti-CFP29 and HBT 4.

## EXAMPLE 6

*Biological activity of the purified antigens.*IFN- $\gamma$  induction in the mouse model of TB infection

The recognition of the purified antigens in the mouse model of memory immunity to TB (described in example 1) was investigated. The results shown in TABLE 5 are representative for three experiments.

A very high IFN- $\gamma$  response was induced by two of the antigens CFP17 and CFP21 at almost the same high level as ST-CF.

TABLE 5

IFN- $\gamma$  release from splenic memory effector cells from C57BL/6J mice isolated after reinfection with *M. tuberculosis* after stimulation with native antigens.

	Antigen <sup>a</sup>	IFN- $\gamma$ (pg/ml) <sup>b</sup>
	ST-CF	12564
	CFP7	ND <sup>d</sup>
10	CFP9	ND
	CFP17	9251
	CFP20	2388
	CFP21	10732
	CFP22 + CFP25 <sup>c</sup>	5342
15	CFP26 (MPT51)	ND
	CFP28	2818
	CFP29	3700

The data is derived from a representative experiment out of three.

<sup>a</sup> ST-CF was tested in a concentration of 5  $\mu$ g/ml and the individual antigens in a concentration of 2  $\mu$ g/ml.

<sup>b</sup> Four days after rechallenge a pool of cells from three mice were tested. The results are expressed as mean of duplicate values and the difference between duplicate cultures are < 15% of mean. The IFN- $\gamma$  release of cultures incubated without antigen was 390 pg/ml.

<sup>c</sup> A pool of CFP22 and CFP25 was tested.

<sup>d</sup> ND, not determined.

#### Skin test reaction in TB infected guinea pigs

The skin test activity of the purified proteins was tested in *M. tuberculosis* infected guinea pigs.

30 1 group of guinea pigs was infected via an ear vein with  $1 \times 10^4$  CFU of *M. tuberculosis* H37Rv in 0,2 ml PBS. After 4

weeks skin tests were performed and 24 hours after injection erythema diameter was measured.

As seen in TABLES 6 and 6a all of the antigens induced a significant Delayed Type Hypersensitivity (DTH) reaction.

5

TABLE 6

DTH erythema diameter in guinea pigs infected with  $1 \times 10^4$  CFU of *M. tuberculosis*, after stimulation with native antigens.

	Antigen <sup>a</sup>	Skin reaction (mm) <sup>b</sup>
	Control	2.00
10	PPD <sup>c</sup>	15.40 (0.53)
	CFP7	ND <sup>e</sup>
	CFP9	ND
	CFP17	11.25 (0.84)
	CFP20	8.88 (0.13)
15	CFP21	12.44 (0.79)
	CFP22 + CFP25 <sup>d</sup>	9.19 (3.10)
	CFP26 (MPT51)	ND
	CFP28	2.90 (1.28)
	CFP29	6.63 (0.88)

20 The values presented are the mean of erythema diameter of four animals and the SEM's are indicated in the brackets. For PPD and CFP29 the values are mean of erythema diameter of ten animals.

<sup>a</sup> The antigens were tested in a concentration of 0,1  $\mu$ g except for CFP29 which was tested in a concentration of 0,8  $\mu$ g.

25 <sup>b</sup> The skin reactions are measured in mm erythema 24 h after intradermal injection.

<sup>c</sup> 10 TU of PPD was used.

<sup>d</sup> A pool of CFP22 and CFP25 was tested.

<sup>e</sup> ND, not determined.

30 Together these analyses indicate that most of the antigens identified were highly biologically active and recognized during TB infection in different animal models.

TABLE 6a

DTH erythema diameter of recombinant antigens in outbred guinea pigs infected with  $1 \times 10^4$  CFU of *M. Tuberculosis*.

	Antigen <sup>a</sup>	Skin reaction (mm) <sup>b</sup>	
5	Control	2.9	(0.3)
	PPD <sup>c</sup>	14.5	(1.0)
	CFP 7a	13.6	(1.4)
	CFP 17	6.8	(1.9)
	CFP 20	6.4	(1.4)
10	CFP 21	5.3	(0.7)
	CFP 25	10.8	(0.8)
	CFP 29	7.4	(2.2)
	MPT 51	4.9	(1.1)

The values presented are the mean of erythema diameter of four animals and the SEM's are indicated in the brackets. For Control, PPD, and CFP 20 the values are mean of erythema diameter of eight animals.

<sup>a</sup> The antigens were tested in a concentration of 1,0 µg.

<sup>b</sup> The skin test reactions are measured in mm erythema 24 h after intradermal infection.

20 <sup>c</sup> 10 TU of PPD was used.

#### *Biological activity of the purified recombinant antigens.*

#### Interferon-γ induction in the mouse model of TB infection.

**Primary infections.** 8 to 12 weeks old female C57BL/6j(H-2<sup>b</sup>), CBA/J(H-2<sup>k</sup>), DBA.2(H-2<sup>d</sup>) and A.SW(H-2<sup>s</sup>) mice (Bomholtegaard, 25 Ry) were given intravenous infections via the lateral tail vein with an inoculum of  $5 \times 10^4$  *M. tuberculosis* suspended in PBS in a vol. of 0.1 ml. 14 days postinfection the animals were sacrificed and spleen cells were isolated and tested for the recognition of recombinant antigen.

30 As seen in TABLE 7 the recombinant antigens rCFP7A, rCFP17, rCFP21, rCFP25, and rCFP29 were all recognized in at least two strains of mice at a level comparable to ST-CF. rMPT51 and rCFP7 were only recognized in one or two strains respectively, at a level corresponding to no more than 1/3 of the

response detected after ST-CF stimulation. Neither of the antigens rCFP20 and rCFP22 were recognized by any of the four mouse strains.

**Memory responses.** 8-12 weeks old female C57BL/6j(H-2<sup>b</sup>) mice (Bomholtegaard, Ry) were given intravenous infections via the lateral tail vein with an inoculum of  $5 \times 10^4$  *M. tuberculosis* suspended in PBS in a vol. of 0.1 ml. After 1 month of infection the mice were treated with isoniazid (Merck and Co., Rahway, NJ) and rifabutin (Farmatalia Carlo Erba, Milano, Italy) in the drinking water, for two months. The mice were rested for 4-6 months before being used in experiments. For the study of the recall of memory immunity, animals were infected with an inoculum of  $1 \times 10^6$  bacteria i.v. and sacrificed at day 4 postinfection. Spleen cells were isolated and tested for the recognition of recombinant antigen. As seen from TABLE 8, IFN- $\gamma$  release after stimulation with rCFP17, rCFP21 and rCFP25 was at the same level as seen from spleen cells stimulated with ST-CF. Stimulation with rCFP7, rCFP7A and rCFP29 all resulted in an IFN- $\gamma$  no higher than 1/3 of the response seen with ST-CF. rCFP22 was not recognized by IFN- $\gamma$  producing cells. None of the antigens stimulated IFN- $\gamma$  release in naive mice. Additionally none of the antigens were toxic to the cell cultures.

TABLE 7. T cell responses in primary TB infection.

Name	c57BL/6J(H2 <sup>b</sup> )	DBA.2(H2 <sup>d</sup> )	CBA/J(H2 <sup>k</sup> )	A.SW(H2 <sup>s</sup> )
rCFP7	+	+	-	-
rCFP7A	+++	+++	+++	+
rCFP17	+++	+	+++	+
rCFP20	-	-	-	-
rCFP21	+++	+++	+++	+
rCFP22	-	-	-	-
rCFP25	+++	++	+++	+

rCFP29	+++	+++	+++	++
rMPT51	+	-	-	-

Mouse IFN- $\gamma$  release during recall of memory immunity to *M. tuberculosis*.

- 5 -:no response; +: 1/3 of ST-CF; ++: 2/3 of ST-CF; +++: level of ST-CF.

**TABLE 8.** T cell responses in memory immune animals.

Name	Memory response
rCFP7	+
10 rCFP7A	++
rCFP17	+++
rCFP21	+++
rCFP22	-
rCFP29	+
15 rCFP25	+++
rMPT51	+

Mouse IFN- $\gamma$  release 14 days after primary infection with *M. tuberculosis*.

- 20 -:no response; +: 1/3 of ST-CF; ++: 2/3 of ST-CF; +++: level of ST-CF.

Interferon- $\gamma$  induction in human TB patients and BCG vaccinated people.

- 25 **Human donors:** PBMC were obtained from healthy BCG vaccinated donors with no known exposure to patients with TB and from patients with culture or microscopy proven infection with *Mycobacterium tuberculosis*. Blood samples were drawn from the TB patients 1-4 months after diagnosis.

- 30 **Lymphocyte preparations and cell culture:** PBMC were freshly isolated by gradient centrifugation of heparinized blood on Lymphoprep (Nycomed, Oslo, Norway). The cells were resuspended in complete medium: RPMI 1640 (Gibco, Grand Island, N.Y.) supplemented with 40  $\mu$ g/ml streptomycin, 40 U/ml penicillin,



and 0.04 mM/ml glutamine, (all from Gibco Laboratories, Paisley, Scotland) and 10% normal human ABO serum (NHS) from the local blood bank. The number and the viability of the cells were determined by trypan blue staining. Cultures were  
5 established with  $2.5 \times 10^5$  PBMC in 200  $\mu$ l in microtitre plates (Nunc, Roskilde, Denmark) and stimulated with no antigen, ST-CF, PPD ( $2.5 \mu$ g/ml); rCFP7, rCFP7A, rCFP17, rCFP20, rCFP21, rCFP22, rCFP25, rCFP26, rCFP29, in a final concentration of 5  $\mu$ g/ml. Phytohaemagglutinin, 1  $\mu$ g/ml (PHA,  
10 Difco laboratories, Detroit, MI. was used as a positive control. Supernatants for the detection of cytokines were harvested after 5 days of culture, pooled and stored at  $-80^\circ\text{C}$  until use.

**Cytokine analysis:** Interferon- $\gamma$  (IFN- $\gamma$ ) was measured with a  
15 standard ELISA technique using a commercially available pair of mAb's from Endogen and used according to the instructions for use. Recombinant IFN- $\gamma$  (Gibco laboratories) was used as a standard. The detection level for the assay was 50 pg/ml. The variation between the duplicate wells did not exceed 10 %  
20 of the mean. Responses of 9 individual donors are shown in TABLE 9.

A seen in TABLE 9 high levels of IFN- $\gamma$  release are obtained after stimulation with several of the recombinant antigens. rCFP7a and rCFP17 gives rise to responses comparable to STCF  
25 in almost all donors. rCFP7 seems to be most strongly recognized by BCG vaccinated healthy donors. rCFP21, rCFP25, rCFP26, and rCFP29 gives rise to a mixed picture with intermediate responses in each group, whereas low responses are obtained by rCFP20 and rCFP22.

**TABLE 9.** Mean values of results from the stimulation of human blood cells from 7 BCG vaccinated and 7 TB patients with recombinant antigens. SE values are given for each antigen. ST-CF and *M. avium* culture filtrate are shown for the comparison.

Controls, Healthy, BCG vaccinated, no known TB exposure

donor:	no ag	PHA	PPD	STCF	CFP7	CFP17	CFP7A	CFP20	CFP21	CFP22	CFP25	CFP26	CFP29
1	6	9564	6774	3966	7034	69	1799	58	152	73	182	946	86
2	48	12486	6603	8067	3146	10044	5267	29	6149	51	1937	526	2065
3	190	11929	10000	8299	8015	11563	8641	437	3194	669	2531	8076	6098
4	10	21029	4106	3537	1323	1939	5211	1	284	1	1344	20	125
5	1	18750	14209	13027	17725	8038	19002	1	3008	1	2103	974	8181

TB patients, 1-4 month after diagnosis

no	ag	PHA	PPD	STCF	CFP7	CFP17	CFP7A	CFP20	CFP21	CFP22	CFP25	CFP26	CFP29
6	9	8973	5096	6145	852	4250	4019	284	1131	48	2400	1078	4584
7	1	12413	6281	3393	168	6375	4505	11	4335	16	3082	1370	5115
8	4	11915	7671	7375	104	2753	3356	119	407	437	2069	712	5284
9	32	22130	16417	17213	8450	9783	16319	91	5957	67	10043	13313	9953

Example 6A

Four groups of 6-8 weeks old, female C57Bl/6J mice (Bomholte-gård, Denmark) were immunized subcutaneously at the base of the tail with vaccines of the following compositions:

- 5 Group 1: 10  $\mu$ g ESAT-6/DDA (250  $\mu$ g)  
 Group 2: 10  $\mu$ g MPT59/DDA (250 $\mu$ g)  
 Group 3: 10  $\mu$ g MPT59-ESAT-6 /DDA (250  $\mu$ g)  
 Group 4: Adjuvant control group: DDA (250  $\mu$ g) in NaCl

The animals were injected with a volume of 0.2 ml. Two weeks  
 10 after the first injection and 3 weeks after the second injection the mice were boosted a little further up the back.  
 One week after the last immunization the mice were bled and the blood cells were isolated. The immune response induced was monitored by release of IFN- $\gamma$  into the culture supernat-  
 15 ants when stimulated in vitro with relevant antigens (see the following table).

Immunogen 10 $\mu$ g/dose	For restimulation <sup>a)</sup> : Ag in vitro			
	no antigen	ST-CF	ESAT-6	MPT59
20 ESAT-6	219 $\pm$ 219	569 $\pm$ 569	835 $\pm$ 633	-
MPT59	0	802 $\pm$ 182	-	5647 $\pm$ 159
Hybrid: MPT59-ESAT-6	127 $\pm$ 127	7453 $\pm$ 581	15133 $\pm$ 861	16363 $\pm$ 1002

- a) Blood cells were isolated 1 week after the last immunization and the release of IFN- $\gamma$  (pg/ml) after 72h of antigen stimulation (5  
 25  $\mu$ g/ml) was measured.  
 The values shown are mean of triplicates performed on cells pooled from three mice  $\pm$  SEM
- b) - not determined

The experiment demonstrates that immunization with the hybrid  
 30 stimulates T cells which recognize ESAT-6 and MPT59 stronger than after single antigen immunization. Especially the recognition of ESAT-6 was enhanced by immunization with the MPT59-ESAT-6 hybrid. IFN- $\gamma$  release in control mice immunized with DDA never exceeded 1000 pg/ml.

## EXAMPLE 6B

The recombinant antigens were tested individually as subunit vaccines in mice. Eleven groups of 6-8 weeks old, female C57Bl/6j mice (Bomholtegård, Denmark) were immunized subcutaneously at the base of the tail with vaccines of the following composition:

- Group 1: 10  $\mu$ g CFP7
- Group 2: 10  $\mu$ g CFP17
- Group 3: 10  $\mu$ g CFP21
- 10 Group 4: 10  $\mu$ g CFP22
- Group 5: 10  $\mu$ g CFP25
- Group 6: 10  $\mu$ g CFP29
- Group 7: 10  $\mu$ g MPT51
- Group 8: 50  $\mu$ g ST-CF
- 15 Group 9: Adjuvant control group
- Group 10: BCG 2,5 x 10<sup>5</sup>/ml, 0,2 ml
- Group 11: Control group: Untreated

All the subunit vaccines were given with DDA as adjuvant. The animals were vaccinated with a volume of 0.2 ml. Two weeks after the first injection and three weeks after the second injection group 1-9 were boosted a little further up the back. One week after the last injection the mice were bled and the blood cells were isolated. The immune response induced was monitored by release of IFN- $\gamma$  into the culture supernatant when stimulated in vitro with the homologous protein.

6 weeks after the last immunization the mice were aerosol challenged with 5 x 10<sup>6</sup> viable *Mycobacterium tuberculosis*/ml. After 6 weeks of infection the mice were killed and the number of viable bacteria in lung and spleen of infected mice was determined by plating serial 3-fold dilutions of organ homogenates on 7H11 plates. Colonies were counted after 2-3 weeks of incubation. The protective efficacy is expressed as the difference between log<sub>10</sub> values of the geometric mean of

counts obtained from five mice of the relevant group and the geometric mean of counts obtained from five mouse of the relevant control group.

The results from the experiments are presented in the following table.

Immunogenicity and protective efficacy in mice, of ST-CF and 7 subunit vaccines

	Subunit Vaccine	Immunogenicity	Protective efficacy
	ST-CF	+++	+++
10	CFP7	++	-
	CFP17	+++	+++
	CFP21	+++	++
	CFP22	-	-
	CFP25	+++	+++
15	CFP29	+++	+++
	MPT51	+++	++

+++ Strong immunogen / high protection (level of BCG)  
 ++ Medium immunogen / medium protection  
 - No recognition / no protection

In conclusion, we have identified a number of proteins inducing high levels of protection. Three of these CFP17, CFP25 and CFP29 giving rise to similar levels of protection as ST-CF and BCG while two proteins CFP21 and MPT51 induces protections around 2/3 the level of BCG and ST-CF. Two of the proteins CFP7 and CFP22 did not induce protection in the mouse model.

#### EXAMPLE 7

*Species distribution of cfp7, cfp9, mpt51, rd1-orf2, rd1-orf3, rd1-orf4, rd1-orf5, rd1-orf8, rd1-orf9a and rd1-orf9b as well as of cfp7a, cfp7b, cfp10a, cfp17, cfp20, cfp21, cfp22, cfp22a, cfp23, cfp25 and cfp25a.*

Presence of *cfp7*, *cfp9*, *mpt51*, *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a* and *rd1-orf9b* in different mycobacterial species.

In order to determine the distribution of the *cfp7*, *cfp9*,  
 5 *mpt51*, *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a* and *rd1-orf9b* genes in species belonging to the *M. tuberculosis*-complex and in other mycobacteria PCR and/or Southern blotting was used. The bacterial strains used are listed in TABLE 10. Genomic DNA was prepared from mycobacte-  
 10 rial cells as described previously (Andersen et al. 1992).

PCR analyses were used in order to determine the distribution of the *cfp7*, *cfp9* and *mpt51* gene in species belonging to the tuberculosis-complex and in other mycobacteria. The bacterial strains used are listed in TABLE 10. PCR was performed on  
 15 genomic DNA prepared from mycobacterial cells as described previously (Andersen et al., 1992).

The oligonucleotide primers used were synthesised automatically on a DNA synthesizer (Applied Biosystems, Forster City, Ca, ABI-391, PCR-mode), deblocked, and purified by ethanol  
 20 precipitation. The primers used for the analyses are shown in TABLE 11.

The PCR amplification was carried out in a thermal reactor (Rapid cyclor, Idaho Technology, Idaho) by mixing 20 ng chromosomal with the mastermix (contained 0.5  $\mu$ M of each  
 25 oligonucleotide primer, 0.25  $\mu$ M BSA (Stratagene), low salt buffer (20 mM Tris-HCl, pH8.8, 10 mM KCl, 10 mM  $(\text{NH}_4)_2\text{SO}_4$ , 2 mM  $\text{MgSO}_4$  and 0,1% Triton X-100) (Stratagene), 0.25 mM of each deoxynucleoside triphosphate and 0.5 U Taq Plus Long DNA polymerase (Stratagene)). Final volume was 10  $\mu$ l (all concen-  
 30 trations given are concentrations in the final volume).

Predenaturation was carried out at 94°C for 30 s. 30 cycles of the following was performed: Denaturation at 94°C for 30 s, annealing at 55°C for 30 s and elongation at 72°C for 1 min.

The following primer combinations were used (the length of the amplified products are given in parentheses):

- mpt51*: MPT51-3 and MPT51-2 (820 bp), MPT51-3 and MPT51-6 (108 bp), MPT51-5 and MPT51-4 (415 bp), MPT51-7 and MPT51-4 (325 bp).
- cfp7*: pVF1 and PVR1 (274 bp), pVF1 and PVR2 (197 bp), pVF3 and PVR1 (302 bp), pVF3 and PVR2 (125 bp).
- cfp9*: stR3 and stF1 (351 bp).

TABLE 10.

10 Mycobacterial strains used in this Example.

	Species and strain(s)	Source
	1. <i>M. tuberculosis</i>	H 3 7 R vATCC <sup>a</sup> ( A T C C 27294)
15	2.	H 3 7 R aATCC ( A T C C 25177)
	3.	Erdman    Obtained from A. Lazlo, Ottawa, Canada
	4. <i>M. bovis</i> BCG substrain: Danish 1331	SSI <sup>b</sup>
20	5.	Chinese    SSI <sup>c</sup>
	6.	Canadian    SSI <sup>c</sup>
	7.	Glaxo    SSI <sup>c</sup>
	8.	Russia    SSI <sup>c</sup>
	9.	Pasteur    SSI <sup>c</sup>
25	10.	Japan    WHO <sup>e</sup>
	11. <i>M. bovis</i> MNC 27	SSI <sup>c</sup>
	12. <i>M. africanum</i>	Isolated from a Danish patient
	13. <i>M. leprae</i> (armadillo-derived)	Obtained from J. M. Colston, London, UK
	14. <i>M. avium</i> (ATCC 15769)	ATCC
30	15. <i>M. kansasii</i> (ATCC 12478)	ATCC
	16. <i>M. marinum</i> (ATCC 927)	ATCC
	17. <i>M. scrofulaceum</i> (ATCC 19275)	ATCC
	18. <i>M. intercellulare</i> (ATCC 15985)	ATCC
	19. <i>M. fortuitum</i> (ATCC 6841)	ATCC
35	20. <i>M. xenopi</i>	Isolated from a Danish patient
	21. <i>M. flavescens</i>	Isolated from a Danish patient
	22. <i>M. szulgai</i>	Isolated from a Danish patient
	23. <i>M. terrae</i>	SSI <sup>c</sup>
	24. <i>E. coli</i>	SSI <sup>d</sup>
40	25. <i>S. aureus</i>	SSI <sup>d</sup>

<sup>a</sup> American Type Culture Collection, USA.

<sup>b</sup> Statens Serum Institut, Copenhagen, Denmark.

<sup>c</sup> Our collection Department of Mycobacteriology, Statens Serum Institut, Copenhagen, Denmark.

<sup>d</sup> Department of Clinical Microbiology, Statens Serum Institut, Denmark.

<sup>e</sup> WHO International Laboratory for Biological Standards, Statens Serum Institut, Copenhagen, Denmark.

TABLE 11.

Sequence of the <i>mpt51</i> , <i>cfp7</i> and <i>cfp9</i> oligonucleotides.		
Orientation and oligonucleotide	Sequences (5'-3') <sup>a</sup>	Position <sup>b</sup> (nucleotides)
10 Sense		
MPT51- 1	<u>CTCGAATTCGCCGGTGCACACAG</u> (SEQ ID NO: 28)	6 - 21 (SEQ ID NO: 41)
MPT51- 3	<u>CTCGAATTCGCCCCATACGAGAAC</u> (SEQ ID NO: 29)	143 - 158 (SEQ ID NO: 41)
15 MPT51- 5	GTGTATCTGCTGGAC (SEQ ID NO: 30)	228 - 242 (SEQ ID NO: 41)
MPT51- 7	CCGACTGGCTGGCCG (SEQ ID NO: 31)	418 - 432 (SEQ ID NO: 41)
pVR1	<u>GTACGAGAATTCATGTCGCAAATCATG</u> (SEQ ID NO: 35)	91 - 105 (SEQ ID NO: 1)
20 pVR2	<u>GTACGAGAATTCGAGCTTGGGGTGCCG</u> (SEQ ID NO: 36)	168 - 181 (SEQ ID NO: 1)
stR3	<u>CGATTCCAAGCTTGTGGCCGCCGACCCG</u> (SEQ ID NO: 37)	141 - 155 (SEQ ID NO: 3)
Antisense		
MPT51- 2	<u>GAGGAATTCGCTTAGCGGATCGCA</u> (SEQ ID NO: 32)	946 - 932 (SEQ ID NO: 41)
25 MPT51- 4	CCCACATTCGGTTGG (SEQ ID NO: 33)	642 - 628 (SEQ ID NO: 41)
MPT51- 6	GTCCAGCAGATACAC (SEQ ID NO: 34)	242 - 228 (SEQ ID NO: 41)
pVF1	<u>CGTTAGGGATCCTCATCGCCATGGTGTG</u> (SEQ ID NO: 38)	340 - 323 (SEQ ID NO: 1)
30 pVF3	<u>CGTTAGGGATCCGGTTCCACTGTGCC</u> (SEQ ID NO: 39)	268 - 255 (SEQ ID NO: 1)
stF1	<u>CGTTAGGGATCCTCAGGTCTTTTCGATG</u> (SEQ ID NO: 40)	467 - 452 (SEQ ID NO: 3)

<sup>a</sup> Nucleotides underlined are not contained in the nucleotide sequences of *mpt51*, *cfp7*, and *cfp9*.

<sup>b</sup> The positions referred to are of the non-underlined parts of the primers and correspond to the nucleotide sequence shown in SEQ ID NOs: 41, 1, and 3 for *mpt51*, *cfp7*, and *cfp9*, respectively.

35 The Southern blotting was carried out as described previously (Oettinger and Andersen, 1994) with the following modifications: 2 µg of genomic DNA was digested with *PvuII*, electrophoresed in an 0.8% agarose gel, and transferred onto a nylon membrane (Hybond N-plus; Amersham International plc, Little Chalfont, United Kingdom) with a vacuum transfer device

40 (Milliblot, TM-v; Millipore Corp., Bedford, MA). The *cfp7*,



*cfp9*, *mpt51*, *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a* and *rd1-orf9b* gene fragments were amplified by PCR from the plasmids pRVN01, pRVN02, pT052, pT087, pT088, pT089, pT090, pT091, pT096 or pT098 by using the primers shown in TABLE 11 and TABLE 2 (in Example 2a). The probes were labelled non-radioactively with an enhanced chemiluminescence kit (ECL; Amersham International plc, Little Chalfont, United Kingdom). Hybridization and detection was performed according to the instructions provided by the manufacturer. The results are summarized in TABLES 12 and 13.

TABLE 12. Interspecies analysis of the *cfp7*, *cfp9* and *mpt51* genes by PCR and/or Southern blotting and of MPT51 protein by Western blotting.

		PCR			Southern blot			Western blot
Species and strain		<i>cfp7</i>	<i>cfp9</i>	<i>mpt51</i>	<i>cfp7</i>	<i>cfp9</i>	<i>mpt51</i>	MPT51
15	1. <i>M. tub.</i> H37Rv	+	+	+	+	+	+	+
	2. <i>M. tub.</i> H37Ra	+	+	+	N.D.	N.D.	+	+
	3. <i>M. tub.</i> Erdmann	+	+	+	+	+	+	+
	4. <i>M. bovis</i>	+	+	+			+	+
	5. <i>M. bovis</i> BCG Danish 1331	+	+	+	+	+	+	+
20	6. <i>M. bovis</i> BCG Japan	+	+	N.D.	+	+	+	N.D.
	7. <i>M. bovis</i> BCG Chinese	+	+	N.D.	+	+	N.D.	N.D.
	8. <i>M. bovis</i> BCG Canadian	+	+	N.D.	+	+	N.D.	N.D.
25	9. <i>M. bovis</i> BCG Glaxo	+	+	N.D.	+	+	N.D.	N.D.
	10. <i>M. bovis</i> BCG Russia	+	+	N.D.	+	+	N.D.	N.D.
	11. <i>M. bovis</i> BCG Pasteur	+	+	N.D.	+	+	N.D.	N.D.
30	12. <i>M. africanum</i>	+	+	+	+	+	+	+
	13. <i>M. leprae</i>	-	-	-	-	-	-	-
	14. <i>M. avium</i>	+	+	-	+	+	+	-
35	15. <i>M. kansasii</i>	+	-	-	+	+	+	-
	16. <i>M. marinum</i>	-	(+)	-	+	+	+	-
	17. <i>M. scrofulaceum</i>	-	-	-	-	-	-	-
	18. <i>M. intercellulare</i>	+	(+)	-	+	+	+	-

Species and strain	PCR			Southern blot			Western blot
	<i>cfp7</i>	<i>cfp9</i>	<i>mpt51</i>	<i>cfp7</i>	<i>cfp9</i>	<i>mpt51</i>	MPT51
19. <i>M. fortuitum</i>	-	-	-	-	-	-	-
20. <i>M. flavescens</i>	+	(+)	-	+	+	+	N.D.
21. <i>M. xenopi</i>	-	-	-	N.D.	N.D.	+	-
22. <i>M. szulgai</i>	(+)	(+)	-	-	+	-	-
5 23. <i>M. terrae</i>	-	-	N.D.	N.D.	N.D.	N.D.	N.D.

+, positive reaction; -, no reaction, N.D. not determined.

- cfp7*, *cfp9* and *mpt51* were found in the *M. tuberculosis* complex including BCG and the environmental mycobacteria; *M. avium*, *M. kansasii*, *M. marinum*, *M. intracellulare* and *M. flavescens*. *cfp9* was additionally found in *M. szulgai* and *mpt51* in *M. xenopi*.

Furthermore the presence of native MPT51 in culture filtrates from different mycobacterial strains was investigated with western blots developed with Mab HBT4.

- 15 There is a strong band at around 26 kDa in *M. tuberculosis* H37Rv, Ra, Erdman, *M. bovis* AN5, *M. bovis* BCG substrain Danish 1331 and *M. africanum*. No band was seen in the region in any other tested mycobacterial strains.

20 **TABLE 13a.** Interspecies analysis of the *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a* and *rd1-orf9b* genes by Southern blotting.

Species and strain	<i>rd1-orf2</i>	<i>rd1-orf3</i>	<i>rd1-orf4</i>	<i>rd1-orf5</i>	<i>rd1-orf8</i>	<i>rd1-orf9a</i>	<i>rd1-orf9b</i>
1. <i>M. tub.</i> H37Rv	+	+	+	+	+	+	+
2. <i>M. bovis</i>	+	+	+	+	N.D.	+	+
3. <i>M. bovis</i> BCG	+	-	-	-	N.D.	-	-
25 Danish 1331							
4. <i>M. bovis</i>	+	-	-	-	N.D.	-	-
BCG Japan							
5. <i>M. avium</i>	-	-	-	-	N.D.	-	-
6. <i>M. kansasii</i>	-	-	-	-	N.D.	-	-
30 7. <i>M. marinum</i>	+	-	+	-	N.D.	-	-
8. <i>M. scrofulaceum</i>	+	-	-	-	N.D.	-	-

Species and strain	<i>rd1-orf2</i>	<i>rd1-orf3</i>	<i>rd1-orf4</i>	<i>rd1-orf5</i>	<i>rd1-orf8</i>	<i>rd1-orf9a</i>	<i>rd1-orf9b</i>
9. <i>M. intercellulare</i>	-	-	-	-	N.D.	-	-
10. <i>M. fortuitum</i>	-	-	-	-	N.D.	-	-
11. <i>M. xenopi</i>	-	-	-	-	N.D.	-	-
12. <i>M. szulgai</i>	+	-	-	-	N.D.	-	-

5 +, positive reaction; -, no reaction, N.D. not determined.

Positive results for *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a* and *rd1-orf9b* were only obtained when using genomic DNA from *M. tuberculosis* and *M. bovis*, and not from *M. bovis* BCG or other mycobacteria analyzed except *rd1-orf4* which also was found in *M. marinum*.

Presence of *cfp7a*, *cfp7b*, *cfp10a*, *cfp17*, *cfp20*, *cfp21*, *cfp22*, *cfp22a*, *cfp23*, *cfp25* and *cfp25a* in different mycobacterial species.

Southern blotting was carried out as described for *rd1-orf2*, *rd1-orf3*, *rd1-orf4*, *rd1-orf5*, *rd1-orf8*, *rd1-orf9a* and *rd1-orf9b*. The *cfp7a*, *cfp7b*, *cfp10a*, *cfp17*, *cfp20*, *cfp21*, *cfp22*, *cfp22a*, *cfp23*, *cfp25* and *cfp25a* gene fragments were amplified by PCR from the recombinant pMCT6 plasmids encoding the individual genes. The primers used (same as the primers used for cloning) are described in example 3, 3A and 3B. The results are summarized in Table 13b.

TABLE 13b. Interspecies analysis of the *cfp7a*, *cfp7b*, *cfp10a*, *cfp17*, *cfp20*, *cfp21*, *cfp22*, *cfp22a*, *cfp23*, *cfp25*, and *cfp25a* genes by Southern blotting.

Species and strain	<i>cfp7a</i>	<i>cfp7b</i>	<i>cfp10a</i>	<i>cfp17</i>	<i>cfp20</i>	<i>cfp21</i>	<i>cfp22</i>	<i>cfp22a</i>	<i>cfp23</i>	<i>cfp25</i>	<i>cfp25a</i>
25 1. <i>M. tub.</i> H37Rv	+	+	+	+	+	+	+	+	+	+	+
2. <i>M. bovis</i>	+	+	+	+	+	+	+	+	+	+	+
3. <i>M. bovis</i> BCG Danish 1331	+	+	+	+	+	N.D.	+	+	+	+	+
4. <i>M. bovis</i> BCG Japan	+	+	+	+	+	+	+	+	+	+	+
5. <i>M. avium</i>	+	N.D.	-	+	-	+	+	+	+	+	-
6. <i>M. kansasii</i>	-	N.D.	+	-	-	-	+	-	+	-	-
7. <i>M. marinum</i>	+	+	-	+	+	+	+	+	+	+	+
8. <i>M. scrofulaceum</i>	-	-	+	-	+	+	-	+	+	+	-
35 9. <i>M. intercellulare</i>	+	+	-	+	-	+	+	-	+	+	-

Species and Strain	cfp7a	cfp7b	cfp10a	cfp17	cfp20	cfp31	cfp22	cfp22a	cfp23	cfp25	cfp25a
10. <i>M. fortuitum</i>	-	N.D.	-	-	-	-	-	-	+	-	-
11. <i>M. xenopi</i>	+	+	+	+	+	+	+	+	+	+	+
12. <i>M. szulgai</i>	+	+	-	+	+	+	+	+	+	+	+

+, positive reaction; -, no reaction, N.D. not determined

Table X: Sensitivity of the ORF3 antigens in African populations divided based on HIV infections status and smear result

	Sens (%) (n=72)	Sens (%) (n=79)	Sens (%) (n=13)
Smear	+	+	-
HIV	-	+	-
RD1-ORF3	59%	49%	70%
38kDa antigen	29%	15%	36%

In high endemic countries a serodiagnostic test would be ideal to supplement or replace current TB diagnostic approaches and in particular is smear negative TB patients and patients co-infected with HIV very difficult to diagnose with the traditional methods. We therefore evaluated the performance of the RD1-ORF3 antigen in sera collected from African patients groups including patients with a HIV co-infection and smear negative patients and the results were compared to the well known serodiagnostic antigen 38kDa. As seen in table x RD1-ORF3 was recognized by 59% of smear positive TB patients but most importantly almost 50 % of the HIV positive patients (compared to 15% for 38kDa antigen), and as many as 70% of the smear negative patients (compared to 36% for 38kDa antigen).

This demonstrates the superior qualities of RD1-ORF3 antigen as a serodiagnostic reagent also to be used in TB patients groups that traditionally is difficult to diagnose.

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## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

## (i) APPLICANT:

- (A) NAME: Statens Seruminstitut
- (B) STREET: Artillerivej 5
- (C) CITY: Copenhagen
- (E) COUNTRY: Denmark
- (F) POSTAL CODE (ZIP): 2300 S

(ii) TITLE OF INVENTION: Nucleic acid fragments and polypeptide fragments derived from M. tuberculosis

(iii) NUMBER OF SEQUENCES: 173

## (iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

## (2) INFORMATION FOR SEQ ID NO: 1:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 381 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: circular

(ii) MOLECULE TYPE: DNA (genomic)

## (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 91..381

## (ix) FEATURE:

- (A) NAME/KEY: -35\_signal
- (B) LOCATION: 14..19

## (ix) FEATURE:

- (A) NAME/KEY: -10\_signal
- (B) LOCATION: 47..50

## (ix) FEATURE:

- (A) NAME/KEY: RBS
- (B) LOCATION: 78..84

## (ix) FEATURE:

- (A) NAME/KEY: mat\_peptide
- (B) LOCATION: 91..381

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

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GGCCGCCGGT ACCTATGTGG CCGCCGATGC TGC GGACGCG TCGACCTATA CCGGGTTCTG      60
ATCGAACCCCT GCTGACCGAG AGGACTTGTG ATG TCG CAA ATC ATG TAC AAC TAC      114
      Met Ser Gln Ile Met Tyr Asn Tyr
      1              5
CCC GCG ATG TTG GGT CAC GCC GGG GAT ATG GCC GGA TAT GCC GGC ACG      162
Pro Ala Met Leu Gly His Ala Gly Asp Met Ala Gly Tyr Ala Gly Thr
      10              15              20
CTG CAG AGC TTG GGT GCC GAG ATC GCC GTG GAG CAG GCC GCG TTG CAG      210
Leu Gln Ser Leu Gly Ala Glu Ile Ala Val Glu Gln Ala Ala Leu Gln
      25              30              35              40
AGT GCG TGG CAG GGC GAT ACC GGG ATC ACG TAT CAG GCG TGG CAG GCA      258
Ser Ala Trp Gln Gly Asp Thr Gly Ile Thr Tyr Gln Ala Trp Gln Ala
      45              50              55
CAG TGG AAC CAG GCC ATG GAA GAT TTG GTG CGG GCC TAT CAT GCG ATG      306
Gln Trp Asn Gln Ala Met Glu Asp Leu Val Arg Ala Tyr His Ala Met
      60              65              70
TCC AGC ACC CAT GAA GCC AAC ACC ATG GCG ATG ATG GCC CGC GAC ACC      354
Ser Ser Thr His Glu Ala Asn Thr Met Ala Met Met Ala Arg Asp Thr
      75              80              85
GCC GAA GCC GCC AAA TGG GGC GGC TAG      381
Ala Glu Ala Ala Lys Trp Gly Gly
      90              95

```

## (2) INFORMATION FOR SEQ ID NO: 2:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 96 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

```

Met Ser Gln Ile Met Tyr Asn Tyr Pro Ala Met Leu Gly His Ala Gly
  1              5              10              15
Asp Met Ala Gly Tyr Ala Gly Thr Leu Gln Ser Leu Gly Ala Glu Ile
      20              25              30
Ala Val Glu Gln Ala Ala Leu Gln Ser Ala Trp Gln Gly Asp Thr Gly
      35              40              45
Ile Thr Tyr Gln Ala Trp Gln Ala Gln Trp Asn Gln Ala Met Glu Asp
      50              55              60
Leu Val Arg Ala Tyr His Ala Met Ser Ser Thr His Glu Ala Asn Thr
      65              70              75              80

```

Met Ala Met Met Ala Arg Asp Thr Ala Glu Ala Ala Lys Trp Gly Gly  
                     85                    90                    95

## (2) INFORMATION FOR SEQ ID NO: 3:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 467 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: circular

## (ii) MOLECULE TYPE: DNA (genomic)

## (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 141..467

## (ix) FEATURE:

- (A) NAME/KEY: -10\_signal
- (B) LOCATION: 73..78

## (ix) FEATURE:

- (A) NAME/KEY: -35\_signal
- (B) LOCATION: 4..9

## (ix) FEATURE:

- (A) NAME/KEY: RBS
- (B) LOCATION: 123..130

## (ix) FEATURE:

- (A) NAME/KEY: mat\_peptide
- (B) LOCATION: 141..467

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

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GGGTAGCCGG ACCACGGCTG GGCAAAGATG TGCAGGCCGC CATCAAGGCG GTCAAGGCCG      60
GCGACGGCGT CATAAACCCG GACGGCACCT TGTTGGCGGG CCCC GCGGTG CTGACGCCCC      120
ACGAGTACAA CTCCCGGCTG GTG GCC GCC GAC CCG GAG TCC ACC GCG GCG      170
                Met Ala Ala Asp Pro Glu Ser Thr Ala Ala
                  1             5             10
TTG CCC GAC GGC GCC GGG CTG GTC GTT CTG GAT GGC ACC GTC ACT GCC      218
Leu Pro Asp Gly Ala Gly Leu Val Val Leu Asp Gly Thr Val Thr Ala
                  15             20             25
GAA CTC GAA GCC GAG GGC TGG GCC AAA GAT CGC ATC CGC GAA CTG CAA      266
Glu Leu Glu Ala Glu Gly Trp Ala Lys Asp Arg Ile Arg Glu Leu Gln
                  30             35             40

```



GAG CTG CGT AAG TCG ACC GGG CTG GAC GTT TCC GAC CGC ATC CGG GTG	314
Glu Leu Arg Lys Ser Thr Gly Leu Asp Val Ser Asp Arg Ile Arg Val	
45 50 55	
GTG ATG TCG GTG CCT GCG GAA CGC GAA GAC TGG GCG CGC ACC CAT CGC	362
Val Met Ser Val Pro Ala Glu Arg Glu Asp Trp Ala Arg Thr His Arg	
60 65 70	
GAC CTC ATT GCC GGA GAA ATC TTG GCT ACC GAC TTC GAA TTC GCC GAC	410
Asp Leu Ile Ala Gly Glu Ile Leu Ala Thr Asp Phe Glu Phe Ala Asp	
75 80 85 90	
CTC GCC GAT GGT GTG GCC ATC GGC GAC GGC GTG CGG GTA AGC ATC GAA	458
Leu Ala Asp Gly Val Ala Ile Gly Asp Gly Val Arg Val Ser Ile Glu	
95 100 105	
AAG ACC TGA	467
Lys Thr	

## (2) INFORMATION FOR SEQ ID NO: 4:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 108 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met Ala Ala Asp Pro Glu Ser Thr Ala Ala Leu Pro Asp Gly Ala Gly	
1 5 10 15	
Leu Val Val Leu Asp Gly Thr Val Thr Ala Glu Leu Glu Ala Glu Gly	
20 25 30	
Trp Ala Lys Asp Arg Ile Arg Glu Leu Gln Glu Leu Arg Lys Ser Thr	
35 40 45	
Gly Leu Asp Val Ser Asp Arg Ile Arg Val Val Met Ser Val Pro Ala	
50 55 60	
Glu Arg Glu Asp Trp Ala Arg Thr His Arg Asp Leu Ile Ala Gly Glu	
65 70 75 80	
Ile Leu Ala Thr Asp Phe Glu Phe Ala Asp Leu Ala Asp Gly Val Ala	
85 90 95	
Ile Gly Asp Gly Val Arg Val Ser Ile Glu Lys Thr	
100 105	

## (2) INFORMATION FOR SEQ ID NO: 5:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 889 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double

(D) TOPOLOGY: circular

(ii) MOLECULE TYPE: DNA (genomic)

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Mycobacterium tuberculosis

(B) STRAIN: H37Rv

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 201..689

(ix) FEATURE:

(A) NAME/KEY: sig\_peptide

(B) LOCATION: 201..290

(ix) FEATURE:

(A) NAME/KEY: mat\_peptide

(B) LOCATION: 291..689

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

CGGGTCTGCA CGGATCCGGG CCGGGCAGGG CAATCGAGCC TGGGATCCGC TGGGGTGCGC	60
ACATCGCGGA CCCGTGCGCG GTACGGTCGA GACAGCGGCA CGAGAAAGTA GTAAGGGCGA	120
TAATAGGCGG TAAAGAGTAG CGGGAAGCCG GCCGAACGAC TCGGTCAGAC AACGCCACAG	180
CGGCCAGTGA GGAGCAGCGG GTG ACG GAC ATG AAC CCG GAT ATT GAG AAG	230
Met Thr Asp Met Asn Pro Asp Ile Glu Lys	
-30 -25	
GAC CAG ACC TCC GAT GAA GTC ACG GTA GAG ACG ACC TCC GTC TTC CGC	278
Asp Gln Thr Ser Asp Glu Val Thr Val Glu Thr Thr Ser Val Phe Arg	
-20 -15 -10 -5	
GCA GAC TTC CTC AGC GAG CTG GAC GCT CCT GCG CAA GCG GGT ACG GAG	326
Ala Asp Phe Leu Ser Glu Leu Asp Ala Pro Ala Gln Ala Gly Thr Glu	
1 5 10	
AGC GCG GTC TCC GGG GTG GAA GGG CTC CCG CCG GGC TCG GCG TTG CTG	374
Ser Ala Val Ser Gly Val Glu Gly Leu Pro Pro Gly Ser Ala Leu Leu	
15 20 25	
GTA GTC AAA CGA GGC CCC AAC GCC GGG TCC CGG TTC CTA CTC GAC CAA	422
Val Val Lys Arg Gly Pro Asn Ala Gly Ser Arg Phe Leu Leu Asp Gln	
30 35 40	
GCC ATC ACG TCG GCT GGT CGG CAT CCC GAC AGC GAC ATA TTT CTC GAC	470
Ala Ile Thr Ser Ala Gly Arg His Pro Asp Ser Asp Ile Phe Leu Asp	
45 50 55 60	
GAC GTG ACC GTG AGC CGT CGC CAT GCT GAA TTC CGG TTG GAA AAC AAC	518
Asp Val Thr Val Ser Arg Arg His Ala Glu Phe Arg Leu Glu Asn Asn	
65 70 75	

GAA TTC AAT GTC GTC GAT GTC GGG AGT CTC AAC GGC ACC TAC GTC AAC	566
Glu Phe Asn Val Val Asp Val Gly Ser Leu Asn Gly Thr Tyr Val Asn	
80 85 90	
CGC GAG CCC GTG GAT TCG GCG GTG CTG GCG AAC GGC GAC GAG GTC CAG	614
Arg Glu Pro Val Asp Ser Ala Val Leu Ala Asn Gly Asp Glu Val Gln	
95 100 105	
ATC GGC AAG TTC CGG TTG GTG TTC TTG ACC GGA CCC AAG CAA GGC GAG	662
Ile Gly Lys Phe Arg Leu Val Phe Leu Thr Gly Pro Lys Gln Gly Glu	
110 115 120	
GAT GAC GGG AGT ACC GGG GGC CCG TGA GCGCACCCGA TAGCCCCGCG	709
Asp Asp Gly Ser Thr Gly Gly Pro	
125 130	
CTGGCCGGGA TGTCGATCGG GCGGTCCTC GACCTGCTAC GACCGGATTT TCCTGATGTC	769
ACCATCTCCA AGATTCGATT CTTGGAGGCT GAGGGTCTGG TGACGCCCCG GCGGGCCTCA	829
TCGGGGTATC GCGGGTTCAC CGCATACGAC TGCGCACGGC TCGGATTCAT TCTCACTGCC	889

## (2) INFORMATION FOR SEQ ID NO: 6:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 162 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

Met Thr Asp Met Asn Pro Asp Ile Glu Lys Asp Gln Thr Ser Asp Glu	
-30 -25 -20 -15	
Val Thr Val Glu Thr Thr Ser Val Phe Arg Ala Asp Phe Leu Ser Glu	
-10 -5 1	
Leu Asp Ala Pro Ala Gln Ala Gly Thr Glu Ser Ala Val Ser Gly Val	
5 10 15	
Glu Gly Leu Pro Pro Gly Ser Ala Leu Leu Val Val Lys Arg Gly Pro	
20 25 30	
Asn Ala Gly Ser Arg Phe Leu Leu Asp Gln Ala Ile Thr Ser Ala Gly	
35 40 45 50	
Arg His Pro Asp Ser Asp Ile Phe Leu Asp Asp Val Thr Val Ser Arg	
55 60 65	
Arg His Ala Glu Phe Arg Leu Glu Asn Asn Glu Phe Asn Val Val Asp	
70 75 80	
Val Gly Ser Leu Asn Gly Thr Tyr Val Asn Arg Glu Pro Val Asp Ser	
85 90 95	

Ala Val Leu Ala Asn Gly Asp Glu Val Gln Ile Gly Lys Phe Arg Leu  
 100 105 110

Val Phe Leu Thr Gly Pro Lys Gln Gly Glu Asp Asp Gly Ser Thr Gly  
 115 120 125 130

Gly Pro

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 898 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: circular

(ii) MOLECULE TYPE: DNA (genomic)

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 201..698

(ix) FEATURE:

- (A) NAME/KEY: mat\_peptide
- (B) LOCATION: 201..698

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

TCGACTCCGG CGCCACCGGG CAGGATCACG GTGTCGACGG GGTCCGCCGGG GAATCCCACG	60
ATAACCACTC TTCGCGCCAT GAATGCCAGT GTTGGCCAGG CGCTGGCCTG GCGTCCACGC	120
CACACACCGC ACAGATTAGG ACACGCCGGC GGCAGAGCCC TGCCCGAAAG ACCGTGCACC	180
GGTCTTGGCA GACTGTGCCC ATG GCA CAG ATA ACC CTG CGA GGA AAC GCG	230
Met Ala Gln Ile Thr Leu Arg Gly Asn Ala	
1 5 10	
ATC AAT ACC GTC GGT GAG CTA CCT GCT GTC GGA TCC CCG GCC CCG GCC	278
Ile Asn Thr Val Gly Glu Leu Pro Ala Val Gly Ser Pro Ala Pro Ala	
15 20 25	
TTC ACC CTG ACC GGG GGC GAT CTG GGG GTG ATC AGC AGC GAC CAG TTC	326
Phe Thr Leu Thr Gly Gly Asp Leu Gly Val Ile Ser Ser Asp Gln Phe	
30 35 40	
CGG GGT AAG TCC GTG TTG CTG AAC ATC TTT CCA TCC GTG GAC ACA CCG	374
Arg Gly Lys Ser Val Leu Leu Asn Ile Phe Pro Ser Val Asp Thr Pro	
45 50 55	

GTG TGC GCG ACG AGT GTG CGA ACC TTC GAC GAG CGT GCG GCG GCA AGT Val Cys Ala Thr Ser Val Arg Thr Phe Asp Glu Arg Ala Ala Ala Ser 60 65 70	422
GGC GCT ACC GTG CTG TGT GTC TCG AAG GAT CTG CCG TTC GCC CAG AAG Gly Ala Thr Val Leu Cys Val Ser Lys Asp Leu Pro Phe Ala Gln Lys 75 80 85 90	470
CGC TTC TGC GGC GCC GAG GGC ACC GAA AAC GTC ATG CCC GCG TCG GCA Arg Phe Cys Gly Ala Glu Gly Thr Glu Asn Val Met Pro Ala Ser Ala 95 100 105	518
TTC CGG GAC AGC TTC GGC GAG GAT TAC GGC GTG ACC ATC GCC GAC GGG Phe Arg Asp Ser Phe Gly Glu Asp Tyr Gly Val Thr Ile Ala Asp Gly 110 115 120	566
CCG ATG GCC GGG CTG CTC GCC CGC GCA ATC GTG GTG ATC GGC GCG GAC Pro Met Ala Gly Leu Leu Ala Arg Ala Ile Val Val Ile Gly Ala Asp 125 130 135	614
GGC AAC GTC GCC TAC ACG GAA TTG GTG CCG GAA ATC GCG CAA GAA CCC Gly Asn Val Ala Tyr Thr Glu Leu Val Pro Glu Ile Ala Gln Glu Pro 140 145 150	662
AAC TAC GAA GCG GCG CTG GCC GCG CTG GGC GCC TAG GCTTTCACAA Asn Tyr Glu Ala Ala Leu Ala Ala Leu Gly Ala 155 160 165	708
GCCCCGCGCG TTCGGCGAGC AGCGCACGAT TTCGAGCGCT GCTCCCGAAA AGCGCCTCGG	768
TGGTCTTGGC CCGGCGGTAA TACAGGTGCA GGTCTGTGCTC CCACGTGAAG GCGATGGCAC	828
CGTGGATCTG AAGAGCGGAG CCGGCGCATA ACACAAAGGT TTCCGCGGTC TGC GCCTTCG	888
CCAGCGGCGC	898

## (2) INFORMATION FOR SEQ ID NO: 8:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 165 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

Met Ala Gln Ile Thr Leu Arg Gly Asn Ala Ile Asn Thr Val Gly Glu 1 5 10 15	
Leu Pro Ala Val Gly Ser Pro Ala Pro Ala Phe Thr Leu Thr Gly Gly 20 25 30	
Asp Leu Gly Val Ile Ser Ser Asp Gln Phe Arg Gly Lys Ser Val Leu 35 40 45	
Leu Asn Ile Phe Pro Ser Val Asp Thr Pro Val Cys Ala Thr Ser Val 50 55 60	

Arg Thr Phe Asp Glu Arg Ala Ala Ala Ser Gly Ala Thr Val Leu Cys  
 65 70 75 80  
 Val Ser Lys Asp Leu Pro Phe Ala Gln Lys Arg Phe Cys Gly Ala Glu  
 85 90 95  
 Gly Thr Glu Asn Val Met Pro Ala Ser Ala Phe Arg Asp Ser Phe Gly  
 100 105 110  
 Glu Asp Tyr Gly Val Thr Ile Ala Asp Gly Pro Met Ala Gly Leu Leu  
 115 120 125  
 Ala Arg Ala Ile Val Val Ile Gly Ala Asp Gly Asn Val Ala Tyr Thr  
 130 135 140  
 Glu Leu Val Pro Glu Ile Ala Gln Glu Pro Asn Tyr Glu Ala Ala Leu  
 145 150 155 160  
 Ala Ala Leu Gly Ala  
 165

## (2) INFORMATION FOR SEQ ID NO: 9:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1054 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: circular

## (ii) MOLECULE TYPE: DNA (genomic)

## (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 201..854

## (ix) FEATURE:

- (A) NAME/KEY: sig\_peptide
- (B) LOCATION: 201..296

## (ix) FEATURE:

- (A) NAME/KEY: mat\_peptide
- (B) LOCATION: 297..854

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

ATAATCAGCT CACCGTTGGG ACCGACCTCG ACCAGGGGTC CTTTGTGACT GCCGGGCTTG 60  
 ACGCGGACGA CCACAGAGTC GGTTCATCGCC TAAGGCTACC GTTCTGACCT GGGGCTGCGT 120  
 GGGCGCCGAC GACGTGAGGC ACGTCATGTC TCAGCGGCCC ACCGCCACCT CGGTCGCCGG 180

CAGTATGTCA GCATGTGCAG ATG ACT CCA CGC AGC CTT GTT CGC ATC GTT	230
Met Thr Pro Arg Ser Leu Val Arg Ile Val	
-32 -30 -25	
GGT GTC GTG GTT GCG ACG ACC TTG GCG CTG GTG AGC GCA CCC GCC GGC	278
Gly Val Val Val Ala Thr Thr Leu Ala Leu Val Ser Ala Pro Ala Gly	
-20 -15 -10	
GGT CGT GCC GCG CAT GCG GAT CCG TGT TCG GAC ATC GCG GTC GTT TTC	326
Gly Arg Ala Ala His Ala Asp Pro Cys Ser Asp Ile Ala Val Val Phe	
-5 1 5 10	
GCT CGC GGC ACG CAT CAG GCT TCT GGT CTT GGC GAC GTC GGT GAG GCG	374
Ala Arg Gly Thr His Gln Ala Ser Gly Leu Gly Asp Val Gly Glu Ala	
15 20 25	
TTC GTC GAC TCG CTT ACC TCG CAA GTT GGC GGG CGG TCG ATT GGG GTC	422
Phe Val Asp Ser Leu Thr Ser Gln Val Gly Gly Arg Ser Ile Gly Val	
30 35 40	
TAC GCG GTG AAC TAC CCA GCA AGC GAC GAC TAC CGC GCG AGC GCG TCA	470
Tyr Ala Val Asn Tyr Pro Ala Ser Asp Asp Tyr Arg Ala Ser Ala Ser	
45 50 55	
AAC GGT TCC GAT GAT GCG AGC GCC CAC ATC CAG CGC ACC GTC GCC AGC	518
Asn Gly Ser Asp Asp Ala Ser Ala His Ile Gln Arg Thr Val Ala Ser	
60 65 70	
TGC CCG AAC ACC AGG ATT GTG CTT GGT GGC TAT TCG CAG GGT GCG ACG	566
Cys Pro Asn Thr Arg Ile Val Leu Gly Gly Tyr Ser Gln Gly Ala Thr	
75 80 85 90	
GTC ATC GAT TTG TCC ACC TCG GCG ATG CCG CCC GCG GTG GCA GAT CAT	614
Val Ile Asp Leu Ser Thr Ser Ala Met Pro Pro Ala Val Ala Asp His	
95 100 105	
GTC GCC GCT GTC GCC CTT TTC GGC GAG CCA TCC AGT GGT TTC TCC AGC	662
Val Ala Ala Val Ala Leu Phe Gly Glu Pro Ser Ser Gly Phe Ser Ser	
110 115 120	
ATG TTG TGG GGC GGC GGG TCG TTG CCG ACA ATC GGT CCG CTG TAT AGC	710
Met Leu Trp Gly Gly Gly Ser Leu Pro Thr Ile Gly Pro Leu Tyr Ser	
125 130 135	
TCT AAG ACC ATA AAC TTG TGT GCT CCC GAC GAT CCA ATA TGC ACC GGA	758
Ser Lys Thr Ile Asn Leu Cys Ala Pro Asp Asp Pro Ile Cys Thr Gly	
140 145 150	
GGC GGC AAT ATT ATG GCG CAT GTT TCG TAT GTT CAG TCG GGG ATG ACA	806
Gly Gly Asn Ile Met Ala His Val Ser Tyr Val Gln Ser Gly Met Thr	
155 160 165 170	
AGC CAG GCG GCG ACA TTC GCG GCG AAC AGG CTC GAT CAC GCC GGA TGA	854
Ser Gln Ala Ala Thr Phe Ala Ala Asn Arg Leu Asp His Ala Gly	
175 180 185	
TCAAAGACTG TTGTCCCTAT ACCGCTGGGG CTGTAGTCGA TGTACACCGG CTGGAATCTG	914

AAGGGCAAGA ACCCGGTATT CATCAGGCCG GATGAAATGA CGGTCGGGCG GTAATCGTTT 974  
 GTGTTGAACG CGTAGAGCCG ATCACCGCCG GGGCTGGTGT AGACCTCAAT GTTTGTGTTC 1034  
 GCCGGCAGGG TTCCGGATCC 1054

## (2) INFORMATION FOR SEQ ID NO: 10:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 217 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

Met Thr Pro Arg Ser Leu Val Arg Ile Val Gly Val Val Val Ala Thr  
 -32 -30 -25 -20

Thr Leu Ala Leu Val Ser Ala Pro Ala Gly Gly Arg Ala Ala His Ala  
 -15 -10 -5

Asp Pro Cys Ser Asp Ile Ala Val Val Phe Ala Arg Gly Thr His Gln  
 1 5 10 15

Ala Ser Gly Leu Gly Asp Val Gly Glu Ala Phe Val Asp Ser Leu Thr  
 20 25 30

Ser Gln Val Gly Gly Arg Ser Ile Gly Val Tyr Ala Val Asn Tyr Pro  
 35 40 45

Ala Ser Asp Asp Tyr Arg Ala Ser Ala Ser Asn Gly Ser Asp Asp Ala  
 50 55 60

Ser Ala His Ile Gln Arg Thr Val Ala Ser Cys Pro Asn Thr Arg Ile  
 65 70 75 80

Val Leu Gly Gly Tyr Ser Gln Gly Ala Thr Val Ile Asp Leu Ser Thr  
 85 90 95

Ser Ala Met Pro Pro Ala Val Ala Asp His Val Ala Ala Val Ala Leu  
 100 105 110

Phe Gly Glu Pro Ser Ser Gly Phe Ser Ser Met Leu Trp Gly Gly Gly  
 115 120 125

Ser Leu Pro Thr Ile Gly Pro Leu Tyr Ser Ser Lys Thr Ile Asn Leu  
 130 135 140

Cys Ala Pro Asp Asp Pro Ile Cys Thr Gly Gly Gly Asn Ile Met Ala  
 145 150 155 160

His Val Ser Tyr Val Gln Ser Gly Met Thr Ser Gln Ala Ala Thr Phe  
 165 170 175

Ala Ala Asn Arg Leu Asp His Ala Gly  
 180 185



## (2) INFORMATION FOR SEQ ID NO: 11:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 949 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: circular

## (ii) MOLECULE TYPE: DNA (genomic)

## (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 201..749

## (ix) FEATURE:

- (A) NAME/KEY: mat\_peptide
- (B) LOCATION: 224..749

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

```

AGCCGCTCGC GTGGGGTCAA CCGGGTTTCC ACCTGCTCAC TCATTTTGCC GCCTTTCTGT      60
GTCCGGGCCG AGGCTTGCGC TCAATAACTC GGTCAAGTTC CTTACAGAC TGCCATCACT      120
GGCCCGTCGG CGGGCTCGTT GCGGGTGCGC CGCGTGCGGG TTTGTGTTCC GGGCACC GGG      180
TGGGGGCCCG CCCGGGCGTA ATG GCA GAC TGT GAT TCC GTG ACT AAC AGC      230
          Met Ala Asp Cys Asp Ser Val Thr Asn Ser
          -7      -5              1

CCC CTT GCG ACC GCT ACC GCC ACG CTG CAC ACT AAC CGC GGC GAC ATC      278
Pro Leu Ala Thr Ala Thr Ala Thr Leu His Thr Asn Arg Gly Asp Ile
      5              10              15

AAG ATC GCC CTG TTC GGA AAC CAT GCG CCC AAG ACC GTC GCC AAT TTT      326
Lys Ile Ala Leu Phe Gly Asn His Ala Pro Lys Thr Val Ala Asn Phe
      20              25              30              35

GTG GGC CTT GCG CAG GGC ACC AAG GAC TAT TCG ACC CAA AAC GCA TCA      374
Val Gly Leu Ala Gln Gly Thr Lys Asp Tyr Ser Thr Gln Asn Ala Ser
          40              45              50

GGT GGC CCG TCC GGC CCG TTC TAC GAC GGC GCG GTC TTT CAC CGG GTG      422
Gly Gly Pro Ser Gly Pro Phe Tyr Asp Gly Ala Val Phe His Arg Val
          55              60              65

ATC CAG GGC TTC ATG ATC CAG GGT GGC GAT CCA ACC GGG ACG GGT CGC      470
Ile Gln Gly Phe Met Ile Gln Gly Gly Asp Pro Thr Gly Thr Gly Arg
          70              75              80

GGC GGA CCC GGC TAC AAG TTC GCC GAC GAG TTC CAC CCC GAG CTG CAA      518
Gly Gly Pro Gly Tyr Lys Phe Ala Asp Glu Phe His Pro Glu Leu Gln
      85              90              95

```

TTC GAC AAG CCC TAT CTG CTC GCG ATG GCC AAC GCC GGT CCG GGC ACC	566
Phe Asp Lys Pro Tyr Leu Leu Ala Met Ala Asn Ala Gly Pro Gly Thr	
100 105 110 115	
AAC GGC TCA CAG TTT TTC ATC ACC GTC GGC AAG ACT CCG CAC CTG AAC	614
Asn Gly Ser Gln Phe Phe Ile Thr Val Gly Lys Thr Pro His Leu Asn	
120 125 130	
CGG CGC CAC ACC ATT TTC GGT GAA GTG ATC GAC GCG GAG TCA CAG CGG	662
Arg Arg His Thr Ile Phe Gly Glu Val Ile Asp Ala Glu Ser Gln Arg	
135 140 145	
GTT GTG GAG GCG ATC TCC AAG ACG GCC ACC GAC GGC AAC GAT CGG CCG	710
Val Val Glu Ala Ile Ser Lys Thr Ala Thr Asp Gly Asn Asp Arg Pro	
150 155 160	
ACG GAC CCG GTG GTG ATC GAG TCG ATC ACC ATC TCC TGA CCCGAAGCTA	759
Thr Asp Pro Val Val Ile Glu Ser Ile Thr Ile Ser	
165 170 175	
CGTCGGCTCG TCGCTCGAAT ACACCTTGTC GACCCGCCAG GGCACGTGGC GGTACACCGA	819
CACGCCGTTG GGGCCGTTCA ACCGGACGCC CTCACGCCAA GTCCGCTCAC CTTTGCCCGC	879
GACCGGCGTA ACCGGCAGCG GTAAGCGCAT CGAGCACCTC CACTGGGTCG GTGCCGAGAT	939
CCCAGCGGGA	949

## (2) INFORMATION FOR SEQ ID NO: 12:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 182 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

Met Ala Asp Cys Asp Ser Val Thr Asn Ser Pro Leu Ala Thr Ala Thr	
-7 -5 1 5	
Ala Thr Leu His Thr Asn Arg Gly Asp Ile Lys Ile Ala Leu Phe Gly	
10 15 20 25	
Asn His Ala Pro Lys Thr Val Ala Asn Phe Val Gly Leu Ala Gln Gly	
30 35 40	
Thr Lys Asp Tyr Ser Thr Gln Asn Ala Ser Gly Gly Pro Ser Gly Pro	
45 50 55	
Phe Tyr Asp Gly Ala Val Phe His Arg Val Ile Gln Gly Phe Met Ile	
60 65 70	
Gln Gly Gly Asp Pro Thr Gly Thr Gly Arg Gly Gly Pro Gly Tyr Lys	
75 80 85	

Phe Ala Asp Glu Phe His Pro Glu Leu Gln Phe Asp Lys Pro Tyr Leu  
 90 95 100 105  
 Leu Ala Met Ala Asn Ala Gly Pro Gly Thr Asn Gly Ser Gln Phe Phe  
 110 115 120  
 Ile Thr Val Gly Lys Thr Pro His Leu Asn Arg Arg His Thr Ile Phe  
 125 130 135  
 Gly Glu Val Ile Asp Ala Glu Ser Gln Arg Val Val Glu Ala Ile Ser  
 140 145 150  
 Lys Thr Ala Thr Asp Gly Asn Asp Arg Pro Thr Asp Pro Val Val Ile  
 155 160 165  
 Glu Ser Ile Thr Ile Ser  
 170 175

## (2) INFORMATION FOR SEQ ID NO: 13:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1060 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: circular

## (ii) MOLECULE TYPE: DNA (genomic)

## (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 201..860

## (ix) FEATURE:

- (A) NAME/KEY: sig\_peptide
- (B) LOCATION: 201..296

## (ix) FEATURE:

- (A) NAME/KEY: mat\_peptide
- (B) LOCATION: 297..860

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

TGGACCTTCA CCGGCGGTCC CTTGCTTCG GGGGCGACAC CTAACATACT GGTGTC AAC 60  
 CTACCGCGAC ACCGCTGGGA CTTTGTGCCA TTGCCGGCCA CTCGGGGCCG CTGCGGCCTG 120  
 GAAAAATTGG TCGGGCACGG GCGGCCGCGG GTCGCTACCA TCCCACTGTG AATGATTAC 180  
 TGACCCGCCG ACTGCTCACC ATG GGC GCG GCC GCC GCA ATG CTG GCC GCG 230  
 Met Gly Ala Ala Ala Ala Met Leu Ala Ala  
 -32 -30 -25

GTG CTT CTG CTT ACT CCC ATC ACC GTT CCC GCC GGC TAC CCC GGT GCC	278
Val Leu Leu Leu Thr Pro Ile Thr Val Pro Ala Gly Tyr Pro Gly Ala	
-20 -15 -10	
GTT GCA CCG GCC ACT GCA GCC TGC CCC GAC GCC GAA GTG GTG TTC GCC	326
Val Ala Pro Ala Thr Ala Ala Cys Pro Asp Ala Glu Val Val Phe Ala	
-5 1 5 10	
CGC GGC CGC TTC GAA CCG CCC GGG ATT GGC ACG GTC GGC AAC GCA TTC	374
Arg Gly Arg Phe Glu Pro Pro Gly Ile Gly Thr Val Gly Asn Ala Phe	
15 20 25	
GTC AGC GCG CTG CGC TCG AAG GTC AAC AAG AAT GTC GGG GTC TAC GCG	422
Val Ser Ala Leu Arg Ser Lys Val Asn Lys Asn Val Gly Val Tyr Ala	
30 35 40	
GTG AAA TAC CCC GCC GAC AAT CAG ATC GAT GTG GGC GCC AAC GAC ATG	470
Val Lys Tyr Pro Ala Asp Asn Gln Ile Asp Val Gly Ala Asn Asp Met	
45 50 55	
AGC GCC CAC ATT CAG AGC ATG GCC AAC AGC TGT CCG AAT ACC CGC CTG	518
Ser Ala His Ile Gln Ser Met Ala Asn Ser Cys Pro Asn Thr Arg Leu	
60 65 70	
GTG CCC GGC GGT TAC TCG CTG GGC GCG GCC GTC ACC GAC GTG GTA CTC	566
Val Pro Gly Gly Tyr Ser Leu Gly Ala Ala Val Thr Asp Val Val Leu	
75 80 85 90	
GCG GTG CCC ACC CAG ATG TGG GGC TTC ACC AAT CCC CTG CCT CCC GGC	614
Ala Val Pro Thr Gln Met Trp Gly Phe Thr Asn Pro Leu Pro Pro Gly	
95 100 105	
AGT GAT GAG CAC ATC GCC GCG GTC GCG CTG TTC GGC AAT GGC AGT CAG	662
Ser Asp Glu His Ile Ala Ala Val Ala Leu Phe Gly Asn Gly Ser Gln	
110 115 120	
TGG GTC GGC CCC ATC ACC AAC TTC AGC CCC GCC TAC AAC GAT CGG ACC	710
Trp Val Gly Pro Ile Thr Asn Phe Ser Pro Ala Tyr Asn Asp Arg Thr	
125 130 135	
ATC GAG TTG TGT CAC GGC GAC GAC CCC GTC TGC CAC CCT GCC GAC CCC	758
Ile Glu Leu Cys His Gly Asp Asp Pro Val Cys His Pro Ala Asp Pro	
140 145 150	
AAC ACC TGG GAG GCC AAC TGG CCC CAG CAC CTC GCC GGG GCC TAT GTC	806
Asn Thr Trp Glu Ala Asn Trp Pro Gln His Leu Ala Gly Ala Tyr Val	
155 160 165 170	
TCG TCG GGC ATG GTC AAC CAG GCG GCT GAC TTC GTT GCC GGA AAG CTG	854
Ser Ser Gly Met Val Asn Gln Ala Ala Asp Phe Val Ala Gly Lys Leu	
175 180 185	
CAA TAG CCACCTAGCC CGTGCGCGAG TCTTTGCTTC ACGCTTTCGC TAACCGACCA	910
Gln	
ACGCGCGCAC GATGGAGGGG TCCGTGGTCA TATCAAGACA AGAAGGGAGT AGGCGATGCA	970

CGCAAAAGTC GGC GACTACC TCGTGGTGAA GGGCACAACC AC GGAACGGC ATGATCAACA 1030  
 TGCTGAGATC ATCGAGGTGC GCTCCGCAGA 1060

## (2) INFORMATION FOR SEQ ID NO: 14:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 219 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

Met Gly Ala Ala Ala Ala Met Leu Ala Ala Val Leu Leu Leu Thr Pro  
 -32 -30 -25 -20  
 Ile Thr Val Pro Ala Gly Tyr Pro Gly Ala Val Ala Pro Ala Thr Ala  
 -15 -10 -5  
 Ala Cys Pro Asp Ala Glu Val Val Phe Ala Arg Gly Arg Phe Glu Pro  
 1 5 10 15  
 Pro Gly Ile Gly Thr Val Gly Asn Ala Phe Val Ser Ala Leu Arg Ser  
 20 25 30  
 Lys Val Asn Lys Asn Val Gly Val Tyr Ala Val Lys Tyr Pro Ala Asp  
 35 40 45  
 Asn Gln Ile Asp Val Gly Ala Asn Asp Met Ser Ala His Ile Gln Ser  
 50 55 60  
 Met Ala Asn Ser Cys Pro Asn Thr Arg Leu Val Pro Gly Gly Tyr Ser  
 65 70 75 80  
 Leu Gly Ala Ala Val Thr Asp Val Val Leu Ala Val Pro Thr Gln Met  
 85 90 95  
 Trp Gly Phe Thr Asn Pro Leu Pro Pro Gly Ser Asp Glu His Ile Ala  
 100 105 110  
 Ala Val Ala Leu Phe Gly Asn Gly Ser Gln Trp Val Gly Pro Ile Thr  
 115 120 125  
 Asn Phe Ser Pro Ala Tyr Asn Asp Arg Thr Ile Glu Leu Cys His Gly  
 130 135 140  
 Asp Asp Pro Val Cys His Pro Ala Asp Pro Asn Thr Trp Glu Ala Asn  
 145 150 155 160  
 Trp Pro Gln His Leu Ala Gly Ala Tyr Val Ser Ser Gly Met Val Asn  
 165 170 175  
 Gln Ala Ala Asp Phe Val Ala Gly Lys Leu Gln  
 180 185

## (2) INFORMATION FOR SEQ ID NO: 15:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1198 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: circular

## (ii) MOLECULE TYPE: DNA (genomic)

## (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 201..998

## (ix) FEATURE:

- (A) NAME/KEY: mat\_peptide
- (B) LOCATION: 201..998

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

```

CAGATGCTGC GCAACATGTT TCTCGGCGAT CCGGCAGGCA ACACCGATCG AGTGCTTGAC      60
TTTTCACCG CGGTGACCGG CGGACTGTTC TTCTACCCA CCATCGACTT TCTCGACCAT      120
CCACCGCCCC TACCGCAGGC GCGGACGCCA ACTCTGGCAG CCGGGTCGCT ATCGATCGGC      180
AGCTTGAAAG GAAGCCCCCG ATG AAC AAT CTC TAC CGC GAT TTG GCA CCG      230
          Met Asn Asn Leu Tyr Arg Asp Leu Ala Pro
              1              5              10

GTC ACC GAA GCC GCT TGG GCG GAA ATC GAA TTG GAG GCG GCG CGG ACG      278
Val Thr Glu Ala Ala Trp Ala Glu Ile Glu Leu Glu Ala Ala Arg Thr
              15              20              25

TTC AAG CGA CAC ATC GCC GGG CGC CGG GTG GTC GAT GTC AGT GAT CCC      326
Phe Lys Arg His Ile Ala Gly Arg Arg Val Val Asp Val Ser Asp Pro
              30              35              40

GGG GGG CCC GTC ACC GCG GCG GTC AGC ACC GGC CGG CTG ATC GAT GTT      374
Gly Gly Pro Val Thr Ala Ala Val Ser Thr Gly Arg Leu Ile Asp Val
              45              50              55

AAG GCA CCA ACC AAC GGC GTG ATC GCC CAC CTG CGG GCC AGC AAA CCC      422
Lys Ala Pro Thr Asn Gly Val Ile Ala His Leu Arg Ala Ser Lys Pro
              60              65              70

CTT GTC CGG CTA CGG GTT CCG TTT ACC CTG TCG CGC AAC GAG ATC GAC      470
Leu Val Arg Leu Arg Val Pro Phe Thr Leu Ser Arg Asn Glu Ile Asp
              75              80              85              90

GAC GTG GAA CGT GGC TCT AAG GAC TCC GAT TGG GAA CCG GTA AAG GAG      518
Asp Val Glu Arg Gly Ser Lys Asp Ser Asp Trp Glu Pro Val Lys Glu
              95              100              105

```

GCG GCC AAG AAG CTG GCC TTC GTC GAG GAC CGC ACA ATA TTC GAA GGC Ala Ala Lys Lys Leu Ala Phe Val Glu Asp Arg Thr Ile Phe Glu Gly 110 115 120	566
TAC AGC GCC GCA TCA ATC GAA GGG ATC CGC AGC GCG AGT TCG AAC CCG Tyr Ser Ala Ala Ser Ile Glu Gly Ile Arg Ser Ala Ser Ser Asn Pro 125 130 135	614
GCG CTG ACG TTG CCC GAG GAT CCC CGT GAA ATC CCT GAT GTC ATC TCC Ala Leu Thr Leu Pro Glu Asp Pro Arg Glu Ile Pro Asp Val Ile Ser 140 145 150	662
CAG GCA TTG TCC GAA CTG CGG TTG GCC GGT GTG GAC GGA CCG TAT TCG Gln Ala Leu Ser Glu Leu Arg Leu Ala Gly Val Asp Gly Pro Tyr Ser 155 160 165 170	710
GTG TTG CTC TCT GCT GAC GTC TAC ACC AAG GTT AGC GAG ACT TCC GAT Val Leu Leu Ser Ala Asp Val Tyr Thr Lys Val Ser Glu Thr Ser Asp 175 180 185	758
CAC GGC TAT CCC ATC CGT GAG CAT CTG AAC CGG CTG GTG GAC GGG GAC His Gly Tyr Pro Ile Arg Glu His Leu Asn Arg Leu Val Asp Gly Asp 190 195 200	806
ATC ATT TGG GCC CCG GCC ATC GAC GGC GCG TTC GTG CTG ACC ACT CGA Ile Ile Trp Ala Pro Ala Ile Asp Gly Ala Phe Val Leu Thr Thr Arg 205 210 215	854
GGC GGC GAC TTC GAC CTA CAG CTG GGC ACC GAC GTT GCA ATC GGG TAC Gly Gly Asp Phe Asp Leu Gln Leu Gly Thr Asp Val Ala Ile Gly Tyr 220 225 230	902
GCC AGC CAC GAC ACG GAC ACC GAG CGC CTC TAC CTG CAG GAG ACG CTG Ala Ser His Asp Thr Asp Thr Glu Arg Leu Tyr Leu Gln Glu Thr Leu 235 240 245 250	950
ACG TTC CTT TGC TAC ACC GCC GAG GCG TCG GTC GCG CTC AGC CAC TAA Thr Phe Leu Cys Tyr Thr Ala Glu Ala Ser Val Ala Leu Ser His 255 260 265	998
GGCACGAGCG CGAGCAATAG CTCCTATGGC AAGCGGCCGC GGGTTGGGTG TGTTCGGAGC	1058
TGGGCTGGTG GACGGTGCGC AGGGCCTGGA AGACGGTGCG GGCTAGGCGG CGTTTGAGGC	1118
AGCGTAGTGC TGCGCGTTTG GTTTTCCCGG CGTCTTGACAG CCTTTGGTAG TAGGCCCTGGC	1178
CCCGGCTGTC GGTCAATCCGG	1198

## (2) INFORMATION FOR SEQ ID NO: 16:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 265 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

Met Asn Asn Leu Tyr Arg Asp Leu Ala Pro Val Thr Glu Ala Ala Trp  
 1 5 10 15  
 Ala Glu Ile Glu Leu Glu Ala Ala Arg Thr Phe Lys Arg His Ile Ala  
 20 25 30  
 Gly Arg Arg Val Val Asp Val Ser Asp Pro Gly Gly Pro Val Thr Ala  
 35 40 45  
 Ala Val Ser Thr Gly Arg Leu Ile Asp Val Lys Ala Pro Thr Asn Gly  
 50 55 60  
 Val Ile Ala His Leu Arg Ala Ser Lys Pro Leu Val Arg Leu Arg Val  
 65 70 75 80  
 Pro Phe Thr Leu Ser Arg Asn Glu Ile Asp Asp Val Glu Arg Gly Ser  
 85 90 95  
 Lys Asp Ser Asp Trp Glu Pro Val Lys Glu Ala Ala Lys Lys Leu Ala  
 100 105 110  
 Phe Val Glu Asp Arg Thr Ile Phe Glu Gly Tyr Ser Ala Ala Ser Ile  
 115 120 125  
 Glu Gly Ile Arg Ser Ala Ser Ser Asn Pro Ala Leu Thr Leu Pro Glu  
 130 135 140  
 Asp Pro Arg Glu Ile Pro Asp Val Ile Ser Gln Ala Leu Ser Glu Leu  
 145 150 155 160  
 Arg Leu Ala Gly Val Asp Gly Pro Tyr Ser Val Leu Leu Ser Ala Asp  
 165 170 175  
 Val Tyr Thr Lys Val Ser Glu Thr Ser Asp His Gly Tyr Pro Ile Arg  
 180 185 190  
 Glu His Leu Asn Arg Leu Val Asp Gly Asp Ile Ile Trp Ala Pro Ala  
 195 200 205  
 Ile Asp Gly Ala Phe Val Leu Thr Thr Arg Gly Gly Asp Phe Asp Leu  
 210 215 220  
 Gln Leu Gly Thr Asp Val Ala Ile Gly Tyr Ala Ser His Asp Thr Asp  
 225 230 235 240  
 Thr Glu Arg Leu Tyr Leu Gln Glu Thr Leu Thr Phe Leu Cys Tyr Thr  
 245 250 255  
 Ala Glu Ala Ser Val Ala Leu Ser His  
 260 265

## (2) INFORMATION FOR SEQ ID NO: 17:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear



(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Mycobacterium tuberculosis

(B) STRAIN: H37Rv

(ix) FEATURE:

(A) NAME/KEY: Duplication

(B) LOCATION: 1

(D) OTHER INFORMATION: Ala is Ala or Ser

(ix) FEATURE:

(A) NAME/KEY: Duplication

(B) LOCATION: 13

(D) OTHER INFORMATION: Xaa is unknown

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

Ala	Glu	Leu	Asp	Ala	Pro	Ala	Gln	Ala	Gly	Thr	Glu	Xaa	Ala	Val
1				5					10					15

(2) INFORMATION FOR SEQ ID NO: 18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Mycobacterium tuberculosis

(B) STRAIN: H37Rv

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

Ala	Gln	Ile	Thr	Leu	Arg	Gly	Asn	Ala	Ile	Asn	Thr	Val	Gly	Glu
1				5				10						15

(2) INFORMATION FOR SEQ ID NO: 19:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

(ix) Feature:

- (A) NAME/KEY: Other
- (B) LOCATION: 3
- (C) OTHER INFORMATION: Xaa is unknown

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

Asp	Pro	Xaa	Ser	Asp	Ile	Ala	Val	Val	Phe	Ala	Arg	Gly	Thr	His
1				5					10					15

(2) INFORMATION FOR SEQ ID NO: 20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

Thr	Asn	Ser	Pro	Leu	Ala	Thr	Ala	Thr	Ala	Thr	Leu	His	Thr	Asn
1				5				10						15

(2) INFORMATION FOR SEQ ID NO: 21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

(ix) Feature:

- (A) NAME/KEY: Other
- (B) LOCATION: 2

(C) OTHER INFORMATION: Xaa is unknown

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

Ala	Xaa	Pro	Asp	Ala	Glu	Val	Val	Phe	Ala	Arg	Gly	Arg	Phe	Glu
1				5				10					15	

(2) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

(ix) Feature:

- (A) NAME/KEY: Other
- (B) LOCATION: 1
- (C) OTHER INFORMATION: Xaa is unknown

(ix) FEATURE:

- (A) NAME/KEY: Duplication
- (B) LOCATION: 2
- (D) OTHER INFORMATION: Ile is Ile or Val

(ix) FEATURE:

- (A) NAME/KEY: Duplication
- (B) LOCATION: 10
- (D) OTHER INFORMATION: Val is Val or Thr

(ix) FEATURE:

- (A) NAME/KEY: Duplication
- (B) LOCATION: 11
- (D) OTHER INFORMATION: Val is Val or Phe

(ix) FEATURE:

- (A) NAME/KEY: Duplication
- (B) LOCATION: 14
- (D) OTHER INFORMATION: Asp is Asp or Gln

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

Xaa	Ile	Gln	Lys	Ser	Leu	Glu	Leu	Ile	Val	Val	Thr	Ala	Asp	Glu
1				5				10					15	

(2) INFORMATION FOR SEQ ID NO: 23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 19 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

Met Asn Asn Leu Tyr Arg Asp Leu Ala Pro Val Thr Glu Ala Ala Trp  
 1                      5                      10                      15

Ala Glu Ile

(2) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 34 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

CCCGGCTCGA GAACCTSTAC CGCGACCTSG CSCC

34

(2) INFORMATION FOR SEQ ID NO: 25:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 37 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 25:

GGGCCGATC CGASGCSGCG TCCTTSACSG GYTGCCA

37

(2) INFORMATION FOR SEQ ID NO: 26:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 28 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26:

GGAAGCCCCA TATGAACAAT CTCTACCG

28

(2) INFORMATION FOR SEQ ID NO: 27:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 32 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:

CGCGCTCAGC CCTTAGTGAC TGAGCGCGAC CG

32

(2) INFORMATION FOR SEQ ID NO: 28:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 24 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:

CTCGAATTCG CCGGGTGCAC ACAG

24

(2) INFORMATION FOR SEQ ID NO: 29:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 25 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:

CTCGAATTCG CCCCCATACG AGAAC

25

(2) INFORMATION FOR SEQ ID NO: 30:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 30:

GTGTATCTGC TGGAC

15

(2) INFORMATION FOR SEQ ID NO: 31:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 31:

CCGACTGGCT GGCCG

15

(2) INFORMATION FOR SEQ ID NO: 32:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 24 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 32:

GAGGAATTCG CTTAGCGGAT CGCA

24

(2) INFORMATION FOR SEQ ID NO: 33:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 33:

CCCACATTCC GTTGG

15

(2) INFORMATION FOR SEQ ID NO: 34:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 34:

GTCCAGCAGA TACAC

15

(2) INFORMATION FOR SEQ ID NO: 35:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 35:

GTACGAGAAT TCATGTCGCA AATCATG

27

(2) INFORMATION FOR SEQ ID NO: 36:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 36:

GTACGAGAAT TCGAGCTTGG GGTGCCG

27

(2) INFORMATION FOR SEQ ID NO: 37:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 28 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 37:

CGATTCCAAG CTTGTGGCCG CCGACCCG

28

(2) INFORMATION FOR SEQ ID NO: 38:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 30 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: YES



(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 38:

CGTTAGGGAT CCTCATCGCC ATGGTGTGG

30

(2) INFORMATION FOR SEQ ID NO: 39:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 26 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 39:

CGTTAGGGAT CCGGTTCCAC TGTGCC

26

(2) INFORMATION FOR SEQ ID NO: 40:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 28 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 40:

CGTTAGGGAT CCTCAGGTCT TTTCGATG

28

(2) INFORMATION FOR SEQ ID NO: 41:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 952 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: circular

(ii) MOLECULE TYPE: DNA (genomic)

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mycobacterium tuberculosis
- (B) STRAIN: H37Rv

## (ix) FEATURE:

(A) NAME/KEY: CDS  
(B) LOCATION: 45..944

## (ix) FEATURE:

(A) NAME/KEY: sig\_peptide  
(B) LOCATION: 45..143

## (ix) FEATURE:

(A) NAME/KEY: mat\_peptide  
(B) LOCATION: 144..941

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 41:

GAATTCGCCG GGTGCACACA GCCTTACACG ACGGAGGTGG ACAC ATG AAG GGT CGG	56
Met Lys Gly Arg	
-33 -30	
TCG GCG CTG CTG CGG GCG CTC TGG ATT GCC GCA CTG TCA TTC GGG TTG	104
Ser Ala Leu Leu Arg Ala Leu Trp Ile Ala Ala Leu Ser Phe Gly Leu	
-25 -20 -15	
GGC GGT GTC GCG GTA GCC GCG GAA CCC ACC GCC AAG GCC GCC CCA TAC	152
Gly Gly Val Ala Val Ala Ala Glu Pro Thr Ala Lys Ala Ala Pro Tyr	
-10 -5 1	
GAG AAC CTG ATG GTG CCG TCG CCC TCG ATG GGC CGG GAC ATC CCG GTG	200
Glu Asn Leu Met Val Pro Ser Pro Ser Met Gly Arg Asp Ile Pro Val	
5 10 15	
GCC TTC CTA GCC GGT GGG CCG CAC GCG GTG TAT CTG CTG GAC GCC TTC	248
Ala Phe Leu Ala Gly Gly Pro His Ala Val Tyr Leu Leu Asp Ala Phe	
20 25 30 35	
AAC GCC GGC CCG GAT GTC AGT AAC TGG GTC ACC GCG GGT AAC GCG ATG	296
Asn Ala Gly Pro Asp Val Ser Asn Trp Val Thr Ala Gly Asn Ala Met	
40 45 50	
AAC ACG TTG GCG GGC AAG GGG ATT TCG GTG GTG GCA CCG GCC GGT GGT	344
Asn Thr Leu Ala Gly Lys Gly Ile Ser Val Val Ala Pro Ala Gly Gly	
55 60 65	
GCG TAC AGC ATG TAC ACC AAC TGG GAG CAG GAT GGC AGC AAG CAG TGG	392
Ala Tyr Ser Met Tyr Thr Asn Trp Glu Gln Asp Gly Ser Lys Gln Trp	
70 75 80	
GAC ACC TTC TTG TCC GCT GAG CTG CCC GAC TGG CTG GCC GCT AAC CGG	440
Asp Thr Phe Leu Ser Ala Glu Leu Pro Asp Trp Leu Ala Ala Asn Arg	
85 90 95	
GGC TTG GCC CCC GGT GGC CAT GCG GCC GTT GGC GCC GCT CAG GGC GGT	488
Gly Leu Ala Pro Gly Gly His Ala Ala Val Gly Ala Ala Gln Gly Gly	
100 105 110 115	
TAC GGG GCG ATG GCG CTG GCG GCC TTC CAC CCC GAC CGC TTC GGC TTC	536
Tyr Gly Ala Met Ala Leu Ala Ala Phe His Pro Asp Arg Phe Gly Phe	
120 125 130	

150

GCT GGC TCG ATG TCG GGC TTT TTG TAC CCG TCG AAC ACC ACC ACC AAC	584
Ala Gly Ser Met Ser Gly Phe Leu Tyr Pro Ser Asn Thr Thr Thr Asn	
135 140 145	
GGT GCG ATC GCG GCG GGC ATG CAG CAA TTC GGC GGT GTG GAC ACC AAC	632
Gly Ala Ile Ala Ala Gly Met Gln Gln Phe Gly Gly Val Asp Thr Asn	
150 155 160	
GGA ATG TGG GGA GCA CCA CAG CTG GGT CGG TGG AAG TGG CAC GAC CCG	680
Gly Met Trp Gly Ala Pro Gln Leu Gly Arg Trp Lys Trp His Asp Pro	
165 170 175	
TGG GTG CAT GCC AGC CTG CTG GCG CAA AAC AAC ACC CGG GTG TGG GTG	728
Trp Val His Ala Ser Leu Leu Ala Gln Asn Asn Thr Arg Val Trp Val	
180 185 190 195	
TGG AGC CCG ACC AAC CCG GGA GCC AGC GAT CCC GCC GCC ATG ATC GGC	776
Trp Ser Pro Thr Asn Pro Gly Ala Ser Asp Pro Ala Ala Met Ile Gly	
200 205 210	
CAA ACC GCC GAG GCG ATG GGT AAC AGC CGC ATG TTC TAC AAC CAG TAT	824
Gln Thr Ala Glu Ala Met Gly Asn Ser Arg Met Phe Tyr Asn Gln Tyr	
215 220 225	
CGC AGC GTC GGC GGG CAC AAC GGA CAC TTC GAC TTC CCA GCC AGC GGT	872
Arg Ser Val Gly Gly His Asn Gly His Phe Asp Phe Pro Ala Ser Gly	
230 235 240	
GAC AAC GGC TGG GGC TCG TGG GCG CCC CAG CTG GGC GCT ATG TCG GGC	920
Asp Asn Gly Trp Gly Ser Trp Ala Pro Gln Leu Gly Ala Met Ser Gly	
245 250 255	
GAT ATC GTC GGT GCG ATC CGC TAA GCGAATTC	952
Asp Ile Val Gly Ala Ile Arg	
260 265	

(2) INFORMATION FOR SEQ ID NO: 42:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 299 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 42:

Met Lys Gly Arg Ser Ala Leu Leu Arg Ala Leu Trp Ile Ala Ala Leu	
-33 -30 -25 -20	
Ser Phe Gly Leu Gly Gly Val Ala Val Ala Ala Glu Pro Thr Ala Lys	
-15 -10 -5	
Ala Ala Pro Tyr Glu Asn Leu Met Val Pro Ser Pro Ser Met Gly Arg	
1 5 10 15	
Asp Ile Pro Val Ala Phe Leu Ala Gly Gly Pro His Ala Val Tyr Leu	
20 25 30	

151

Leu	Asp	Ala	Phe	Asn	Ala	Gly	Pro	Asp	Val	Ser	Asn	Trp	Val	Thr	Ala
		35						40					45		
Gly	Asn	Ala	Met	Asn	Thr	Leu	Ala	Gly	Lys	Gly	Ile	Ser	Val	Val	Ala
		50					55					60			
Pro	Ala	Gly	Gly	Ala	Tyr	Ser	Met	Tyr	Thr	Asn	Trp	Glu	Gln	Asp	Gly
		65				70					75				
Ser	Lys	Gln	Trp	Asp	Thr	Phe	Leu	Ser	Ala	Glu	Leu	Pro	Asp	Trp	Leu
	80				85					90					95
Ala	Ala	Asn	Arg	Gly	Leu	Ala	Pro	Gly	Gly	His	Ala	Ala	Val	Gly	Ala
			100					105					110		
Ala	Gln	Gly	Gly	Tyr	Gly	Ala	Met	Ala	Leu	Ala	Ala	Phe	His	Pro	Asp
		115					120						125		
Arg	Phe	Gly	Phe	Ala	Gly	Ser	Met	Ser	Gly	Phe	Leu	Tyr	Pro	Ser	Asn
		130					135					140			
Thr	Thr	Thr	Asn	Gly	Ala	Ile	Ala	Ala	Gly	Met	Gln	Gln	Phe	Gly	Gly
		145				150					155				
Val	Asp	Thr	Asn	Gly	Met	Trp	Gly	Ala	Pro	Gln	Leu	Gly	Arg	Trp	Lys
	160				165					170					175
Trp	His	Asp	Pro	Trp	Val	His	Ala	Ser	Leu	Leu	Ala	Gln	Asn	Asn	Thr
			180						185					190	
Arg	Val	Trp	Val	Trp	Ser	Pro	Thr	Asn	Pro	Gly	Ala	Ser	Asp	Pro	Ala
			195					200					205		
Ala	Met	Ile	Gly	Gln	Thr	Ala	Glu	Ala	Met	Gly	Asn	Ser	Arg	Met	Phe
		210					215					220			
Tyr	Asn	Gln	Tyr	Arg	Ser	Val	Gly	Gly	His	Asn	Gly	His	Phe	Asp	Phe
	225					230					235				
Pro	Ala	Ser	Gly	Asp	Asn	Gly	Trp	Gly	Ser	Trp	Ala	Pro	Gln	Leu	Gly
	240				245					250					255
Ala	Met	Ser	Gly	Asp	Ile	Val	Gly	Ala	Ile	Arg					
				260					265						

(2) INFORMATION FOR SEQ ID NO: 43:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 43:

GCAACACCCG GGATGTCGCA AATCATG

27

(2) INFORMATION FOR SEQ ID NO: 44:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 44:

GTAACACCCG GGGTGGCCGC CGACCCG

27

(2) INFORMATION FOR SEQ ID NO: 45:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 37 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 45:

CTACTAAGCT TGGATCCCTA GCCGCCCCAT TTGGCGG

37

(2) INFORMATION FOR SEQ ID NO: 46:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 38 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(iv) ANTI-SENSE: YES

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 46:

CTACTAAGCT TCCATGGTCA GGTCTTTTCG ATGCTTAC

38

## (2) INFORMATION FOR SEQ ID NO: 47:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 450 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 105...320

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 47:

GTGCCGCGCT CCCAGGGTT CTTATGGTTC GATATACCTG AGTTTGATGG AAGTCCGATG 60  
 ACCAGCAGTC AGCATACGGC ATGGCCGAAA AGAGTGGGGT GATG ATG GCC GAG GAT 116  
 Met Ala Glu Asp  
 1

GTT CGC GCC GAG ATC GTG GCC AGC GTT CTC GAA GTC GTT GTC AAC GAA 164  
 Val Arg Ala Glu Ile Val Ala Ser Val Leu Glu Val Val Val Asn Glu  
 5 10 15 20

GGC GAT CAG ATC GAC AAG GGC GAC GTC GTG GTG CTG CTG GAG TCG ATG 212  
 Gly Asp Gln Ile Asp Lys Gly Asp Val Val Val Leu Leu Glu Ser Met  
 25 30 35

AAG ATG GAG ATC CCC GTC CTG GCC GAA GCT GCC GGA ACG GTC AGC AAG 260  
 Lys Met Glu Ile Pro Val Leu Ala Glu Ala Ala Gly Thr Val Ser Lys  
 40 45 50

GTG GCG GTA TCG GTG GGC GAT GTC ATT CAG GCC GGC GAC CTT ATC GCG 308  
 Val Ala Val Ser Val Gly Asp Val Ile Gln Ala Gly Asp Leu Ile Ala  
 55 60 65

GTG ATC AGC TAGTCGTTGA TAGTCACTCA TGTCCACACT CGGTGATCTG CTCGCCGAA 366  
 Val Ile Ser  
 70

CACACGGTGC TGCCGGGCAG CGCGGTGGAC CACCTGCATG CGGTGGTCGG GGAGTGGCAG 426

CTCCTTGCCG ACTTGTCGTT TGCC 450

## (2) INFORMATION FOR SEQ ID NO: 48:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 71 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein  
(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 48:

```

Met Ala Glu Asp Val Arg Ala Glu Ile Val Ala Ser Val Leu Glu Val
 1             5             10             15
Val Val Asn Glu Gly Asp Gln Ile Asp Lys Gly Asp Val Val Val Leu
             20             25             30
Leu Glu Ser Met Lys Met Glu Ile Pro Val Leu Ala Glu Ala Ala Gly
             35             40             45
Thr Val Ser Lys Val Ala Val Ser Val Gly Asp Val Ile Gln Ala Gly
             50             55             60
Asp Leu Ile Ala Val Ile Ser
65             70

```

(2) INFORMATION FOR SEQ ID NO: 49:

(i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 750 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence  
 (B) LOCATION: 113...640  
 (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 49:

```

GGGTACCCAT CGATGGGTTG CGGTTTCGGCA CCGAGGTGCT AACGCACTTG CTGACACACT      60
GCTAGTCGAA AACGAGGCTA GTCGCAACGT CGATCACACG AGAGGACTGA CC ATG ACA      118
                               Met Thr
                               1
ACT TCA CCC GAC CCG TAT GCC GCG CTG CCC AAG CTG CCG TCC TTC AGC      166
Thr Ser Pro Asp Pro Tyr Ala Ala Leu Pro Lys Leu Pro Ser Phe Ser
      5             10             15
CTG ACG TCA ACC TCG ATC ACC GAT GGG CAG CCG CTG GCT ACA CCC CAG      214
Leu Thr Ser Thr Ser Ile Thr Asp Gly Gln Pro Leu Ala Thr Pro Gln
      20             25             30
GTC AGC GGG ATC ATG GGT GCG GGC GGG GCG GAT GCC AGT CCG CAG CTG      262
Val Ser Gly Ile Met Gly Ala Gly Gly Ala Asp Ala Ser Pro Gln Leu
      35             40             45             50

```

155

AGG TGG TCG GGA TTT CCC AGC GAG ACC CGC AGC TTC GCG GTA ACC GTC	310
Arg Trp Ser Gly Phe Pro Ser Glu Thr Arg Ser Phe Ala Val Thr Val	
55 60 65	
TAC GAC CCT GAT GCC CCC ACC CTG TCC GGG TTC TGG CAC TGG GCG GTG	358
Tyr Asp Pro Asp Ala Pro Thr Leu Ser Gly Phe Trp His Trp Ala Val	
70 75 80	
GCC AAC CTG CCT GCC AAC GTC ACC GAG TTG CCC GAG GGT GTC GGC GAT	406
Ala Asn Leu Pro Ala Asn Val Thr Glu Leu Pro Glu Gly Val Gly Asp	
85 90 95	
GGC CGC GAA CTG CCG GGC GGG GCA CTG ACA TTG GTC AAC GAC GCC GGT	454
Gly Arg Glu Leu Pro Gly Gly Ala Leu Thr Leu Val Asn Asp Ala Gly	
100 105 110	
ATG CGC CGG TAT GTG GGT GCG GCG CCG CCT CCC GGT CAT GGG GTG CAT	502
Met Arg Arg Tyr Val Gly Ala Ala Pro Pro Pro Gly His Gly Val His	
115 120 125 130	
CGC TAC TAC GTC GCG GTA CAC GCG GTG AAG GTC GAA AAG CTC GAC CTC	550
Arg Tyr Tyr Val Ala Val His Ala Val Lys Val Glu Lys Leu Asp Leu	
135 140 145	
CCC GAG GAC GCG AGT CCT GCA TAT CTG GGA TTC AAC CTG TTC CAG CAC	598
Pro Glu Asp Ala Ser Pro Ala Tyr Leu Gly Phe Asn Leu Phe Gln His	
150 155 160	
GCG ATT GCA CGA GCG GTC ATC TTC GGC ACC TAC GAG CAG CGT TAGCGCTTT	649
Ala Ile Ala Arg Ala Val Ile Phe Gly Thr Tyr Glu Gln Arg	
165 170 175	
AGCTGGGTTG CCGACGTCTT GCCGAGCCGA CCGCTTCGTG CAGCGAGCCG AACCCGCCGT	709
CATGCAGCCT GCGGGCAATG CCTTCATGGA TGTCTTGGC C	750

(2) INFORMATION FOR SEQ ID NO: 50:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 176 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 50:

Met Thr Thr Ser Pro Asp Pro Tyr Ala Ala Leu Pro Lys Leu Pro Ser	
1 5 10 15	
Phe Ser Leu Thr Ser Thr Ser Ile Thr Asp Gly Gln Pro Leu Ala Thr	
20 25 30	
Pro Gln Val Ser Gly Ile Met Gly Ala Gly Gly Ala Asp Ala Ser Pro	
35 40 45	



Gln Leu Arg Trp Ser Gly Phe Pro Ser Glu Thr Arg Ser Phe Ala Val  
 50 55 60  
 Thr Val Tyr Asp Pro Asp Ala Pro Thr Leu Ser Gly Phe Trp His Trp  
 65 70 75 80  
 Ala Val Ala Asn Leu Pro Ala Asn Val Thr Glu Leu Pro Glu Gly Val  
 85 90 95  
 Gly Asp Gly Arg Glu Leu Pro Gly Gly Ala Leu Thr Leu Val Asn Asp  
 100 105 110  
 Ala Gly Met Arg Arg Tyr Val Gly Ala Ala Pro Pro Pro Gly His Gly  
 115 120 125  
 Val His Arg Tyr Tyr Val Ala Val His Ala Val Lys Val Glu Lys Leu  
 130 135 140  
 Asp Leu Pro Glu Asp Ala Ser Pro Ala Tyr Leu Gly Phe Asn Leu Phe  
 145 150 155 160  
 Gln His Ala Ile Ala Arg Ala Val Ile Phe Gly Thr Tyr Glu Gln Arg  
 165 170 175

## (2) INFORMATION FOR SEQ ID NO: 51:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 800 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 18...695
- (D) OTHER INFORMATION:

- (A) NAME/KEY: Signal Sequence
- (B) LOCATION: 18...134
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 51:

TCATGAGGTT CATCGGG GTG ATC CCA CGC CCG CAG CCG CAT TCG GGC CGC 50  
 Met Ile Pro Arg Pro Gln Pro His Ser Gly Arg  
 -35 -30  
 TGG CGA GCC GGT GCC GCA CGC CGC CTC ACC AGC CTG GTG GCC GCC GCC 98  
 Trp Arg Ala Gly Ala Ala Arg Arg Leu Thr Ser Leu Val Ala Ala Ala  
 -25 -20 -15  
 TTT GCG GCG GCC ACA CTG TTG CTT ACC CCC GCG CTG GCA CCA CCG GCA 146  
 Phe Ala Ala Ala Thr Leu Leu Leu Thr Pro Ala Leu Ala Pro Pro Ala  
 -10 -5 1 5

TCG GCG GGC TGC CCG GAT GCC GAG GTG GTG TTC GCC CGC GGA ACC GGC Ser Ala Gly Cys Pro Asp Ala Glu Val Val Phe Ala Arg Gly Thr Gly 10 15 20	194
GAA CCA CCT GGC CTC GGT CGG GTA GGC CAA GCT TTC GTC AGT TCA TTG Glu Pro Pro Gly Leu Gly Arg Val Gly Gln Ala Phe Val Ser Ser Leu 25 30 35	242
CGC CAG CAG ACC AAC AAG AGC ATC GGG ACA TAC GGA GTC AAC TAC CCG Arg Gln Gln Thr Asn Lys Ser Ile Gly Thr Tyr Gly Val Asn Tyr Pro 40 45 50	290
GCC AAC GGT GAT TTC TTG GCC GCC GCT GAC GGC GCG AAC GAC GCC AGC Ala Asn Gly Asp Phe Leu Ala Ala Ala Asp Gly Ala Asn Asp Ala Ser 55 60 65	338
GAC CAC ATT CAG CAG ATG GCC AGC GCG TGC CGG GCC ACG AGG TTG GTG Asp His Ile Gln Gln Met Ala Ser Ala Cys Arg Ala Thr Arg Leu Val 70 75 80 85	386
CTC GGC GGC TAC TCC CAG GGT GCG GCC GTG ATC GAC ATC GTC ACC GCC Leu Gly Gly Tyr Ser Gln Gly Ala Ala Val Ile Asp Ile Val Thr Ala 90 95 100	434
GCA CCA CTG CCG GGC CTC GGG TTC ACG CAG CCG TTG CCG CCC GCA GCG Ala Pro Leu Pro Gly Leu Gly Phe Thr Gln Pro Leu Pro Pro Ala Ala 105 110 115	482
GAC GAT CAC ATC GCC GCG ATC GCC CTG TTC GGG AAT CCC TCG GGC CGC Asp Asp His Ile Ala Ala Ile Ala Leu Phe Gly Asn Pro Ser Gly Arg 120 125 130	530
GCT GGC GGG CTG ATG AGC GCC CTG ACC CCT CAA TTC GGG TCC AAG ACC Ala Gly Gly Leu Met Ser Ala Leu Thr Pro Gln Phe Gly Ser Lys Thr 135 140 145	578
ATC AAC CTC TGC AAC AAC GGC GAC CCG ATT TGT TCG GAC GGC AAC CGG Ile Asn Leu Cys Asn Asn Gly Asp Pro Ile Cys Ser Asp Gly Asn Arg 150 155 160 165	626
TGG CGA GCG CAC CTA GGC TAC GTG CCC GGG ATG ACC AAC CAG GCG GCG Trp Arg Ala His Leu Gly Tyr Val Pro Gly Met Thr Asn Gln Ala Ala 170 175 180	674
CGT TTC GTC GCG AGC AGG ATC TAACGCGAGC CGCCCCATAG ATTCCGGCTA AGCA Arg Phe Val Ala Ser Arg Ile 185	729
ACGGCTGCGC CGCCGCCCGG CCACGAGTGA CCGCCGCCGA CTGGCACACC GCTTACCACG	789
GCCTTATGCT G	800

## (2) INFORMATION FOR SEQ ID NO: 52:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 226 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein  
(v) FRAGMENT TYPE: internal  
(ix) FEATURE:

(A) NAME/KEY: Signal Sequence  
(B) LOCATION: 1...38  
(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 52:

```

Met Ile Pro Arg Pro Gln Pro His Ser Gly Arg Trp Arg Ala Gly Ala
      -35              -30              -25

Ala Arg Arg Leu Thr Ser Leu Val Ala Ala Ala Phe Ala Ala Ala Thr
      -20              -15              -10

Leu Leu Leu Thr Pro Ala Leu Ala Pro Pro Ala Ser Ala Gly Cys Pro
      -5              1              5              10

Asp Ala Glu Val Val Phe Ala Arg Gly Thr Gly Glu Pro Pro Gly Leu
      15              20              25

Gly Arg Val Gly Gln Ala Phe Val Ser Ser Leu Arg Gln Gln Thr Asn
      30              35              40

Lys Ser Ile Gly Thr Tyr Gly Val Asn Tyr Pro Ala Asn Gly Asp Phe
      45              50              55

Leu Ala Ala Ala Asp Gly Ala Asn Asp Ala Ser Asp His Ile Gln Gln
      60              65              70

Met Ala Ser Ala Cys Arg Ala Thr Arg Leu Val Leu Gly Gly Tyr Ser
      75              80              85              90

Gln Gly Ala Ala Val Ile Asp Ile Val Thr Ala Ala Pro Leu Pro Gly
      95              100             105

Leu Gly Phe Thr Gln Pro Leu Pro Pro Ala Ala Asp Asp His Ile Ala
      110             115             120

Ala Ile Ala Leu Phe Gly Asn Pro Ser Gly Arg Ala Gly Gly Leu Met
      125             130             135

Ser Ala Leu Thr Pro Gln Phe Gly Ser Lys Thr Ile Asn Leu Cys Asn
      140             145             150

Asn Gly Asp Pro Ile Cys Ser Asp Gly Asn Arg Trp Arg Ala His Leu
      155             160             165             170

Gly Tyr Val Pro Gly Met Thr Asn Gln Ala Ala Arg Phe Val Ala Ser
      175             180             185

Arg Ile

```

## (2) INFORMATION FOR SEQ ID NO: 53:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 700 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 73...615
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 53:

```

CTAGGAAAGC CTTTCCTGAG TAAGTATTGC CTCGTTGCA TACCGCCCTT TACCTGCGTT      60
AATCTGCATT TT ATG ACA GAA TAC GAA GGG CCT AAG ACA AAA TTC CAC GCG      111
      Met Thr Glu Tyr Glu Gly Pro Lys Thr Lys Phe His Ala
          1              5              10

TTA ATG CAG GAA CAG ATT CAT AAC GAA TTC ACA GCG GCA CAA CAA TAT      159
Leu Met Gln Glu Gln Ile His Asn Glu Phe Thr Ala Ala Gln Gln Tyr
      15              20              25

GTC GCG ATC GCG GTT TAT TTC GAC AGC GAA GAC CTG CCG CAG TTG GCG      207
Val Ala Ile Ala Val Tyr Phe Asp Ser Glu Asp Leu Pro Gln Leu Ala
      30              35              40              45

AAG CAT TTT TAC AGC CAA GCG GTC GAG GAA CGA AAC CAT GCA ATG ATG      255
Lys His Phe Tyr Ser Gln Ala Val Glu Glu Arg Asn His Ala Met Met
          50              55              60

CTC GTG CAA CAC CTG CTC GAC CGC GAC CTT CGT GTC GAA ATT CCC GGC      303
Leu Val Gln His Leu Leu Asp Arg Asp Leu Arg Val Glu Ile Pro Gly
          65              70              75

GTA GAC ACG GTG CGA AAC CAG TTC GAC AGA CCC CGC GAG GCA CTG GCG      351
Val Asp Thr Val Arg Asn Gln Phe Asp Arg Pro Arg Glu Ala Leu Ala
          80              85              90

CTG GCG CTC GAT CAG GAA CGC ACA GTC ACC GAC CAG GTC GGT CGG CTG      399
Leu Ala Leu Asp Gln Glu Arg Thr Val Thr Asp Gln Val Gly Arg Leu
          95              100             105

ACA GCG GTG GCC CGC GAC GAG GGC GAT TTC CTC GGC GAG CAG TTC ATG      447
Thr Ala Val Ala Arg Asp Glu Gly Asp Phe Leu Gly Glu Gln Phe Met
      110              115              120              125

CAG TGG TTC TTG CAG GAA CAG ATC GAA GAG GTG GCC TTG ATG GCA ACC      495
Gln Trp Phe Leu Gln Glu Gln Ile Glu Glu Val Ala Leu Met Ala Thr
          130              135              140

CTG GTG CGG GTT GCC GAT CGG GCC GGG GCC AAC CTG TTC GAG CTA GAG      543
Leu Val Arg Val Ala Asp Arg Ala Gly Ala Asn Leu Phe Glu Leu Glu
          145              150              155

```

160

AAC TTC GTC GCA CGT GAA GTG GAT GTG GCG CCG GCC GCA TCA GGC GCC 591  
Asn Phe Val Ala Arg Glu Val Asp Val Ala Pro Ala Ala Ser Gly Ala  
160 165 170

CCG CAC GCT GCC GGG GGC CGC CTC TAGATCCCTG GCGGGGATCA GCGAGTGGTC 645  
Pro His Ala Ala Gly Gly Arg Leu  
175 180

CCGTTGCCCC GCCCGTCTTC CAGCCAGGCC TTGGTGCGGC CGGGGTGGTG AGTAC 700

(2) INFORMATION FOR SEQ ID NO: 54:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 181 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 54:

Met Thr Glu Tyr Glu Gly Pro Lys Thr Lys Phe His Ala Leu Met Gln  
1 5 10 15  
Glu Gln Ile His Asn Glu Phe Thr Ala Ala Gln Gln Tyr Val Ala Ile  
20 25 30  
Ala Val Tyr Phe Asp Ser Glu Asp Leu Pro Gln Leu Ala Lys His Phe  
35 40 45  
Tyr Ser Gln Ala Val Glu Glu Arg Asn His Ala Met Met Leu Val Gln  
50 55 60  
His Leu Leu Asp Arg Asp Leu Arg Val Glu Ile Pro Gly Val Asp Thr  
65 70 75 80  
Val Arg Asn Gln Phe Asp Arg Pro Arg Glu Ala Leu Ala Leu Ala Leu  
85 90 95  
Asp Gln Glu Arg Thr Val Thr Asp Gln Val Gly Arg Leu Thr Ala Val  
100 105 110  
Ala Arg Asp Glu Gly Asp Phe Leu Gly Glu Gln Phe Met Gln Trp Phe  
115 120 125  
Leu Gln Glu Gln Ile Glu Glu Val Ala Leu Met Ala Thr Leu Val Arg  
130 135 140  
Val Ala Asp Arg Ala Gly Ala Asn Leu Phe Glu Leu Glu Asn Phe Val  
145 150 155 160  
Ala Arg Glu Val Asp Val Ala Pro Ala Ala Ser Gly Ala Pro His Ala  
165 170 175

Ala Gly Gly Arg Leu  
180

(2) INFORMATION FOR SEQ ID NO: 55:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 950 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 133...918
- (D) OTHER INFORMATION:

- (A) NAME/KEY: Signal Sequence
- (B) LOCATION: 133...233
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 55:

TGGGCTCGGC ACTGGCTCTC CCACGGTGGC GCGCTGATTT CTCCCCACGG TAGGCGTTGC	60
GACGCATGTT CTTACCGTC TATCCACAGC TACCGACATT TGCTCCGGCT GGATCGCGGG	120
TAAAATTCCG TC GTG AAC AAT CGA CCC ATC CGC CTG CTG ACA TCC GGC AGG	171
Met Asn Asn Arg Pro Ile Arg Leu Leu Thr Ser Gly Arg	
-30 -25	
GCT GGT TTG GGT GCG GGC GCA TTG ATC ACC GCC GTC GTC CTG CTC ATC	219
Ala Gly Leu Gly Ala Gly Ala Leu Ile Thr Ala Val Val Leu Leu Ile	
-20 -15 -10 -5	
GCC TTG GGC GCT GTT TGG ACC CCG GTT GCC TTC GCC GAT GGA TGC CCG	267
Ala Leu Gly Ala Val Trp Thr Pro Val Ala Phe Ala Asp Gly Cys Pro	
1 5 10	
GAC GCC GAA GTC ACG TTC GCC CGC GGC ACC GGC GAG CCG CCC GGA ATC	315
Asp Ala Glu Val Thr Phe Ala Arg Gly Thr Gly Glu Pro Pro Gly Ile	
15 20 25	
GGG CGC GTT GGC CAG GCG TTC GTC GAC TCG CTG CGC CAG CAG ACT GGC	363
Gly Arg Val Gly Gln Ala Phe Val Asp Ser Leu Arg Gln Gln Thr Gly	
30 35 40	
ATG GAG ATC GGA GTA TAC CCG GTG AAT TAC GCC GCC AGC CGC CTA CAG	411
Met Glu Ile Gly Val Tyr Pro Val Asn Tyr Ala Ala Ser Arg Leu Gln	
45 50 55 60	
CTG CAC GGG GGA GAC GGC GCC AAC GAC GCC ATA TCG CAC ATT AAG TCC	459
Leu His Gly Gly Asp Gly Ala Asn Asp Ala Ile Ser His Ile Lys Ser	
65 70 75	

ATG GCC TCG TCA TGC CCG AAC ACC AAG CTG GTC TTG GGC GGC TAT TCG	507
Met Ala Ser Ser Cys Pro Asn Thr Lys Leu Val Leu Gly Gly Tyr Ser	
80 85 90	
CAG GGC GCA ACC GTG ATC GAT ATC GTG GCC GGG GTT CCG TTG GGC AGC	555
Gln Gly Ala Thr Val Ile Asp Ile Val Ala Gly Val Pro Leu Gly Ser	
95 100 105	
ATC AGC TTT GGC AGT CCG CTA CCT GCG GCA TAC GCA GAC AAC GTC GCA	603
Ile Ser Phe Gly Ser Pro Leu Pro Ala Ala Tyr Ala Asp Asn Val Ala	
110 115 120	
GCG GTC GCG GTC TTC GGC AAT CCG TCC AAC CGC GCC GGC GGA TCG CTG	651
Ala Val Ala Val Phe Gly Asn Pro Ser Asn Arg Ala Gly Gly Ser Leu	
125 130 135 140	
TCG AGC CTG AGC CCG CTA TTC GGT TCC AAG GCG ATT GAC CTG TGC AAT	699
Ser Ser Leu Ser Pro Leu Phe Gly Ser Lys Ala Ile Asp Leu Cys Asn	
145 150 155	
CCC ACC GAT CCG ATC TGC CAT GTG GGC CCC GGC AAC GAA TTC AGC GGA	747
Pro Thr Asp Pro Ile Cys His Val Gly Pro Gly Asn Glu Phe Ser Gly	
160 165 170	
CAC ATC GAC GGC TAC ATA CCC ACC TAC ACC ACC CAG GCG GCT AGT TTC	795
His Ile Asp Gly Tyr Ile Pro Thr Tyr Thr Thr Gln Ala Ala Ser Phe	
175 180 185	
GTC GTG CAG AGG CTC CGC GCC GGG TCG GTG CCA CAT CTG CCT GGA TCC	843
Val Val Gln Arg Leu Arg Ala Gly Ser Val Pro His Leu Pro Gly Ser	
190 195 200	
GTC CCG CAG CTG CCC GGG TCT GTC CTT CAG ATG CCC GGC ACT GCC GCA	891
Val Pro Gln Leu Pro Gly Ser Val Leu Gln Met Pro Gly Thr Ala Ala	
205 210 215 220	
CCG GCT CCC GAA TCG CTG CAC GGT CGC TGACGCTTTG TCAGTAAGCC CATAAAA	945
Pro Ala Pro Glu Ser Leu His Gly Arg	
225	
TCGCG	950

## (2) INFORMATION FOR SEQ ID NO: 56:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 262 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (ix) FEATURE:

- (A) NAME/KEY: Signal Sequence
- (B) LOCATION: 1...33

## (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 57:

Met Asn Asn Arg Pro Ile Arg Leu Leu Thr Ser Gly Arg Ala Gly Leu  
                     -30                    -25                    -20

Gly Ala Gly Ala Leu Ile Thr Ala Val Val Leu Leu Ile Ala Leu Gly  
                     -15                    -10                    -5

Ala Val Trp Thr Pro Val Ala Phe Ala Asp Gly Cys Pro Asp Ala Glu  
           1                    5                    10                    15

Val Thr Phe Ala Arg Gly Thr Gly Glu Pro Pro Gly Ile Gly Arg Val  
                     20                    25                    30

Gly Gln Ala Phe Val Asp Ser Leu Arg Gln Gln Thr Gly Met Glu Ile  
                     35                    40                    45

Gly Val Tyr Pro Val Asn Tyr Ala Ala Ser Arg Leu Gln Leu His Gly  
           50                    55                    60

Gly Asp Gly Ala Asn Asp Ala Ile Ser His Ile Lys Ser Met Ala Ser  
           65                    70                    75

Ser Cys Pro Asn Thr Lys Leu Val Leu Gly Gly Tyr Ser Gln Gly Ala  
           80                    85                    90                    95

Thr Val Ile Asp Ile Val Ala Gly Val Pro Leu Gly Ser Ile Ser Phe  
                     100                    105                    110

Gly Ser Pro Leu Pro Ala Ala Tyr Ala Asp Asn Val Ala Ala Val Ala  
           115                    120                    125

Val Phe Gly Asn Pro Ser Asn Arg Ala Gly Gly Ser Leu Ser Ser Leu  
           130                    135                    140

Ser Pro Leu Phe Gly Ser Lys Ala Ile Asp Leu Cys Asn Pro Thr Asp  
           145                    150                    155

Pro Ile Cys His Val Gly Pro Gly Asn Glu Phe Ser Gly His Ile Asp  
           160                    165                    170                    175

Gly Tyr Ile Pro Thr Tyr Thr Thr Gln Ala Ala Ser Phe Val Val Gln  
                     180                    185                    190

Arg Leu Arg Ala Gly Ser Val Pro His Leu Pro Gly Ser Val Pro Gln  
           195                    200                    205

Leu Pro Gly Ser Val Leu Gln Met Pro Gly Thr Ala Ala Pro Ala Pro  
           210                    215                    220

Glu Ser Leu His Gly Arg  
           225

(2) INFORMATION FOR SEQ ID NO: 57:



## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1000 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 94...966
- (D) OTHER INFORMATION:

- (A) NAME/KEY: Signal Sequence
- (B) LOCATION: 94...264
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 57:

```

CGAGGAGACC GACGATCTGC TCGACGAAAT CGACGACGTC CTCGAGGAGA ACGCCGAGGA      60
CTTCGTCCGC GCATACGTCC AAAAGGGCGG ACA GTG ACC TGG CCG TTG CCC GAT      114
                               Met Thr Trp Pro Leu Pro Asp
                               -55                               -50

CGC CTG TCC ATT AAT TCA CTC TCT GGA ACA CCC GCT GTA GAC CTA TCT      162
Arg Leu Ser Ile Asn Ser Leu Ser Gly Thr Pro Ala Val Asp Leu Ser
                               -45                               -40                               -35

TCT TTC ACT GAC TTC CTG CGC CGC CAG GCG CCG GAG TTG CTG CCG GCA      210
Ser Phe Thr Asp Phe Leu Arg Arg Gln Ala Pro Glu Leu Leu Pro Ala
                               -30                               -25                               -20

AGC ATC AGC GGC GGT GCG CCA CTC GCA GGC GGC GAT GCG CAA CTG CCG      258
Ser Ile Ser Gly Gly Ala Pro Leu Ala Gly Gly Asp Ala Gln Leu Pro
                               -15                               -10                               -5

CAC GGC ACC ACC ATT GTC GCG CTG AAA TAC CCC GGC GGT GTT GTC ATG      306
His Gly Thr Thr Ile Val Ala Leu Lys Tyr Pro Gly Gly Val Val Met
                               1                               5                               10                               15

GCG GGT GAC CGG CGT TCG ACG CAG GGC AAC ATG ATT TCT GGG CGT GAT      354
Ala Gly Asp Arg Arg Ser Thr Gln Gly Asn Met Ile Ser Gly Arg Asp
                               20                               25                               30

GTG CGC AAG GTG TAT ATC ACC GAT GAC TAC ACC GCT ACC GGC ATC GCT      402
Val Arg Lys Val Tyr Ile Thr Asp Asp Tyr Thr Ala Thr Gly Ile Ala
                               35                               40                               45

GGC ACG GCT GCG GTC GCG GTT GAG TTT GCC CGG CTG TAT GCC GTG GAA      450
Gly Thr Ala Ala Val Ala Val Glu Phe Ala Arg Leu Tyr Ala Val Glu
                               50                               55                               60

CTT GAG CAC TAC GAG AAG CTC GAG GGT GTG CCG CTG ACG TTT GCC GGC      498
Leu Glu His Tyr Glu Lys Leu Glu Gly Val Pro Leu Thr Phe Ala Gly
                               65                               70                               75

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165

AAA ATC AAC CGG CTG GCG ATT ATG GTG CGT GGC AAT CTG GCG GCC GCG	546
Lys Ile Asn Arg Leu Ala Ile Met Val Arg Gly Asn Leu Ala Ala Ala	
80 85 90 95	
ATG CAG GGT CTG CTG GCG TTG CCG TTG CTG GCG GGC TAC GAC ATT CAT	594
Met Gln Gly Leu Leu Ala Leu Pro Leu Leu Ala Gly Tyr Asp Ile His	
100 105 110	
GCG TCT GAC CCG CAG AGC GCG GGT CGT ATC GTT TCG TTC GAC GCC GCC	642
Ala Ser Asp Pro Gln Ser Ala Gly Arg Ile Val Ser Phe Asp Ala Ala	
115 120 125	
GGC GGT TGG AAC ATC GAG GAA GAG GGC TAT CAG GCG GTG GGC TCG GGT	690
Gly Gly Trp Asn Ile Glu Glu Glu Gly Tyr Gln Ala Val Gly Ser Gly	
130 135 140	
TCG CTG TTC GCG AAG TCG TCG ATG AAG AAG TTG TAT TCG CAG GTT ACC	738
Ser Leu Phe Ala Lys Ser Ser Met Lys Lys Leu Tyr Ser Gln Val Thr	
145 150 155	
GAC GGT GAT TCG GGG CTG CGG GTG GCG GTC GAG GCG CTC TAC GAC GCC	786
Asp Gly Asp Ser Gly Leu Arg Val Ala Val Glu Ala Leu Tyr Asp Ala	
160 165 170 175	
GCC GAC GAC GAC TCC GCC ACC GGC GGT CCG GAC CTG GTG CGG GGC ATC	834
Ala Asp Asp Asp Ser Ala Thr Gly Gly Pro Asp Leu Val Arg Gly Ile	
180 185 190	
TTT CCG ACG GCG GTG ATC ATC GAC GCC GAC GGG GCG GTT GAC GTG CCG	882
Phe Pro Thr Ala Val Ile Ile Asp Ala Asp Gly Ala Val Asp Val Pro	
195 200 205	
GAG AGC CGG ATT GCC GAA TTG GCC CGC GCG ATC ATC GAA AGC CGT TCG	930
Glu Ser Arg Ile Ala Glu Leu Ala Arg Ala Ile Ile Glu Ser Arg Ser	
210 215 220	
GGT GCG GAT ACT TTC GGC TCC GAT GGC GGT GAG AAG TGAGTTTTCC GTATTT	982
Gly Ala Asp Thr Phe Gly Ser Asp Gly Gly Glu Lys	
225 230 235	
CATCTCGCCT GAGCAGGC	1000

(2) INFORMATION FOR SEQ ID NO: 58:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 291 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(ix) FEATURE:

- (A) NAME/KEY: Signal Sequence
- (B) LOCATION: 1...56

## (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 58:

Met	Thr	Trp	Pro	Leu	Pro	Asp	Arg	Leu	Ser	Ile	Asn	Ser	Leu	Ser	Gly	-55	-50	-45	
Thr	Pro	Ala	Val	Asp	Leu	Ser	Ser	Phe	Thr	Asp	Phe	Leu	Arg	Arg	Gln	-40	-35	-30	-25
Ala	Pro	Glu	Leu	Leu	Pro	Ala	Ser	Ile	Ser	Gly	Gly	Ala	Pro	Leu	Ala	-20	-15	-10	
Gly	Gly	Asp	Ala	Gln	Leu	Pro	His	Gly	Thr	Thr	Ile	Val	Ala	Leu	Lys	-5	1	5	
Tyr	Pro	Gly	Gly	Val	Val	Met	Ala	Gly	Asp	Arg	Arg	Ser	Thr	Gln	Gly	10	15	20	
Asn	Met	Ile	Ser	Gly	Arg	Asp	Val	Arg	Lys	Val	Tyr	Ile	Thr	Asp	Asp	25	30	35	40
Tyr	Thr	Ala	Thr	Gly	Ile	Ala	Gly	Thr	Ala	Ala	Val	Ala	Val	Glu	Phe	45	50	55	
Ala	Arg	Leu	Tyr	Ala	Val	Glu	Leu	Glu	His	Tyr	Glu	Lys	Leu	Glu	Gly	60	65	70	
Val	Pro	Leu	Thr	Phe	Ala	Gly	Lys	Ile	Asn	Arg	Leu	Ala	Ile	Met	Val	75	80	85	
Arg	Gly	Asn	Leu	Ala	Ala	Ala	Met	Gln	Gly	Leu	Leu	Ala	Leu	Pro	Leu	90	95	100	
Leu	Ala	Gly	Tyr	Asp	Ile	His	Ala	Ser	Asp	Pro	Gln	Ser	Ala	Gly	Arg	105	110	115	120
Ile	Val	Ser	Phe	Asp	Ala	Ala	Gly	Gly	Trp	Asn	Ile	Glu	Glu	Glu	Gly	125	130	135	
Tyr	Gln	Ala	Val	Gly	Ser	Gly	Ser	Leu	Phe	Ala	Lys	Ser	Ser	Met	Lys	140	145	150	
Lys	Leu	Tyr	Ser	Gln	Val	Thr	Asp	Gly	Asp	Ser	Gly	Leu	Arg	Val	Ala	155	160	165	
Val	Glu	Ala	Leu	Tyr	Asp	Ala	Ala	Asp	Asp	Asp	Ser	Ala	Thr	Gly	Gly	170	175	180	
Pro	Asp	Leu	Val	Arg	Gly	Ile	Phe	Pro	Thr	Ala	Val	Ile	Ile	Asp	Ala	185	190	195	200
Asp	Gly	Ala	Val	Asp	Val	Pro	Glu	Ser	Arg	Ile	Ala	Glu	Leu	Ala	Arg	205	210	215	
Ala	Ile	Ile	Glu	Ser	Arg	Ser	Gly	Ala	Asp	Thr	Phe	Gly	Ser	Asp	Gly	220	225	230	

Gly Glu Lys  
235

(2) INFORMATION FOR SEQ ID NO: 59:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 900 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 66...808
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 59:

TTGGCCCGCG CGATCATCGA AAGCCGTTTCG GGTGCGGATA CTTTCGGCTC CGATGGCGGT	60
GAGAA GTG AGT TTT CCG TAT TTC ATC TCG CCT GAG CAG GCG ATG CGC GAG	110
Met Ser Phe Pro Tyr Phe Ile Ser Pro Glu Gln Ala Met Arg Glu	
1 5 10 15	
CGC AGC GAG TTG GCG CGT AAG GGC ATT GCG CGG GCC AAA AGC GTG GTG	158
Arg Ser Glu Leu Ala Arg Lys Gly Ile Ala Arg Ala Lys Ser Val Val	
20 25 30	
GCG CTG GCC TAT GCC GGT GGT GTG CTG TTC GTC GCG GAG AAT CCG TCG	206
Ala Leu Ala Tyr Ala Gly Gly Val Leu Phe Val Ala Glu Asn Pro Ser	
35 40 45	
CGG TCG CTG CAG AAG ATC AGT GAG CTC TAC GAT CGG GTG GGT TTT GCG	254
Arg Ser Leu Gln Lys Ile Ser Glu Leu Tyr Asp Arg Val Gly Phe Ala	
50 55 60	
GCT GCG GGC AAG TTC AAC GAG TTC GAC AAT TTG CGC CGC GGC GGG ATC	302
Ala Ala Gly Lys Phe Asn Glu Phe Asp Asn Leu Arg Arg Gly Gly Ile	
65 70 75	
CAG TTC GCC GAC ACC CGC GGT TAC GCC TAT GAC CGT CGT GAC GTC ACG	350
Gln Phe Ala Asp Thr Arg Gly Tyr Ala Tyr Asp Arg Arg Asp Val Thr	
80 85 90 95	
GGT CGG CAG TTG GCC AAT GTC TAC GCG CAG ACT CTA GGC ACC ATC TTC	398
Gly Arg Gln Leu Ala Asn Val Tyr Ala Gln Thr Leu Gly Thr Ile Phe	
100 105 110	
ACC GAA CAG GCC AAG CCC TAC GAG GTT GAG TTG TGT GTG GCC GAG GTG	446
Thr Glu Gln Ala Lys Pro Tyr Glu Val Glu Leu Cys Val Ala Glu Val	
115 120 125	
GCG CAT TAC GGC GAG ACG AAA CGC CCT GAG TTG TAT CGT ATT ACC TAC	494
Ala His Tyr Gly Glu Thr Lys Arg Pro Glu Leu Tyr Arg Ile Thr Tyr	
130 135 140	

GAC GGG TCG ATC GCC GAC GAG CCG CAT TTC GTG GTG ATG GGC GGC ACC Asp Gly Ser Ile Ala Asp Glu Pro His Phe Val Val Met Gly Gly Thr 145 150 155	542
ACG GAG CCG ATC GCC AAC GCG CTC AAA GAG TCG TAT GCC GAG AAC GCC Thr Glu Pro Ile Ala Asn Ala Leu Lys Glu Ser Tyr Ala Glu Asn Ala 160 165 170 175	590
AGC CTG ACC GAC GCC CTG CGT ATC GCG GTC GCT GCA TTG CGG GCC GGC Ser Leu Thr Asp Ala Leu Arg Ile Ala Val Ala Ala Leu Arg Ala Gly 180 185 190	638
AGT GCC GAC ACC TCG GGT GGT GAT CAA CCC ACC CTT GGC GTG GCC AGC Ser Ala Asp Thr Ser Gly Gly Asp Gln Pro Thr Leu Gly Val Ala Ser 195 200 205	686
TTA GAG GTG GCC GTT CTC GAT GCC AAC CGG CCA CGG CGC GCG TTC CGG Leu Glu Val Ala Val Leu Asp Ala Asn Arg Pro Arg Arg Ala Phe Arg 210 215 220	734
CGC ATC ACC GGC TCC GCC CTG CAA GCG TTG CTG GTA GAC CAG GAA AGC Arg Ile Thr Gly Ser Ala Leu Gln Ala Leu Leu Val Asp Gln Glu Ser 225 230 235	782
CCG CAG TCT GAC GGC GAA TCG TCG GG CTGAGTCCGA AAGTCCGACG CGTGTCTG Pro Gln Ser Asp Gly Glu Ser Ser Gly 240 245	836
GGACCCCGCT GCGACGTTAA CTGCGCCTAA CCCC GGCTCG ACGCGTCGCC GGCCGTCCTG	896
ACTT	900

## (2) INFORMATION FOR SEQ ID NO: 60:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 248 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 60:

Met Ser Phe Pro Tyr Phe Ile Ser Pro Glu Gln Ala Met Arg Glu Arg 1 5 10 15
Ser Glu Leu Ala Arg Lys Gly Ile Ala Arg Ala Lys Ser Val Val Ala 20 25 30
Leu Ala Tyr Ala Gly Gly Val Leu Phe Val Ala Glu Asn Pro Ser Arg 35 40 45
Ser Leu Gln Lys Ile Ser Glu Leu Tyr Asp Arg Val Gly Phe Ala Ala 50 55 60

169

Ala Gly Lys Phe Asn Glu Phe Asp Asn Leu Arg Arg Gly Gly Ile Gln	
65	70 75 80
Phe Ala Asp Thr Arg Gly Tyr Ala Tyr Asp Arg Arg Asp Val Thr Gly	
	85 90 95
Arg Gln Leu Ala Asn Val Tyr Ala Gln Thr Leu Gly Thr Ile Phe Thr	
	100 105 110
Glu Gln Ala Lys Pro Tyr Glu Val Glu Leu Cys Val Ala Glu Val Ala	
	115 120 125
His Tyr Gly Glu Thr Lys Arg Pro Glu Leu Tyr Arg Ile Thr Tyr Asp	
	130 135 140
Gly Ser Ile Ala Asp Glu Pro His Phe Val Val Met Gly Gly Thr Thr	
	145 150 155 160
Glu Pro Ile Ala Asn Ala Leu Lys Glu Ser Tyr Ala Glu Asn Ala Ser	
	165 170 175
Leu Thr Asp Ala Leu Arg Ile Ala Val Ala Ala Leu Arg Ala Gly Ser	
	180 185 190
Ala Asp Thr Ser Gly Gly Asp Gln Pro Thr Leu Gly Val Ala Ser Leu	
	195 200 205
Glu Val Ala Val Leu Asp Ala Asn Arg Pro Arg Arg Ala Phe Arg Arg	
	210 215 220
Ile Thr Gly Ser Ala Leu Gln Ala Leu Leu Val Asp Gln Glu Ser Pro	
	225 230 235 240
Gln Ser Asp Gly Glu Ser Ser Gly	
	245

(2) INFORMATION FOR SEQ ID NO:61:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1560 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 98...1487
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 61:

GAGTCATTGC CTGGTCGGCG TCATTCCGTA CTAGTCGGTT GTCGGACTTG ACCTACTGGG	60
TCAGGCCGAC GAGCACTCGA CCATTAGGGT AGGGGCC GTG ACC CAC TAT GAC GTC	115
Met Thr His Tyr Asp Val	
1 5	

GTC GTT CTC GGA GCC GGT CCC GGC GGG TAT GTC GCG GCG ATT CGC GCC	163
Val Val Leu Gly Ala Gly Pro Gly Gly Tyr Val Ala Ala Ile Arg Ala	
10 15 20	
GCA CAG CTC GGC CTG AGC ACT GCA ATC GTC GAA CCC AAG TAC TGG GGC	211
Ala Gln Leu Gly Leu Ser Thr Ala Ile Val Glu Pro Lys Tyr Trp Gly	
25 30 35	
GGA GTA TGC CTC AAT GTC GGC TGT ATC CCA TCC AAG GCG CTG TTG CGC	259
Gly Val Cys Leu Asn Val Gly Cys Ile Pro Ser Lys Ala Leu Leu Arg	
40 45 50	
AAC GCC GAA CTG GTC CAC ATC TTC ACC AAG GAC GCC AAA GCA TTT GGC	307
Asn Ala Glu Leu Val His Ile Phe Thr Lys Asp Ala Lys Ala Phe Gly	
55 60 65 70	
ATC AGC GGC GAG GTG ACC TTC GAC TAC GGC ATC GCC TAT GAC CGC AGC	355
Ile Ser Gly Glu Val Thr Phe Asp Tyr Gly Ile Ala Tyr Asp Arg Ser	
75 80 85	
CGA AAG GTA GCC GAG GGC AGG GTG GCC GGT GTG CAC TTC CTG ATG AAG	403
Arg Lys Val Ala Glu Gly Arg Val Ala Gly Val His Phe Leu Met Lys	
90 95 100	
AAG AAC AAG ATC ACC GAG ATC CAC GGG TAC GGC ACA TTT GCC GAC GCC	451
Lys Asn Lys Ile Thr Glu Ile His Gly Tyr Gly Thr Phe Ala Asp Ala	
105 110 115	
AAC ACG TTG TTG GTT GAT CTC AAC GAC GGC GGT ACA GAA TCG GTC ACG	499
Asn Thr Leu Leu Val Asp Leu Asn Asp Gly Gly Thr Glu Ser Val Thr	
120 125 130	
TTC GAC AAC GCC ATC ATC GCG ACC GGC AGT AGC ACC CGG CTG GTT CCC	547
Phe Asp Asn Ala Ile Ile Ala Thr Gly Ser Ser Thr Arg Leu Val Pro	
135 140 145 150	
GGC ACC TCA CTG TCG GCC AAC GTA GTC ACC TAC GAG GAA CAG ATC CTG	595
Gly Thr Ser Leu Ser Ala Asn Val Val Thr Tyr Glu Glu Gln Ile Leu	
155 160 165	
TCC CGA GAG CTG CCG AAA TCG ATC ATT ATT GCC GGA GCT GGT GCC ATT	643
Ser Arg Glu Leu Pro Lys Ser Ile Ile Ile Ala Gly Ala Gly Ala Ile	
170 175 180	
GGC ATG GAG TTC GGC TAC GTG CTG AAG AAC TAC GGC GTT GAC GTG ACC	691
Gly Met Glu Phe Gly Tyr Val Leu Lys Asn Tyr Gly Val Asp Val Thr	
185 190 195	
ATC GTG GAA TTC CTT CCG CGG GCG CTG CCC AAC GAG GAC GCC GAT GTG	739
Ile Val Glu Phe Leu Pro Arg Ala Leu Pro Asn Glu Asp Ala Asp Val	
200 205 210	
TCC AAG GAG ATC GAG AAG CAG TTC AAA AAG CTG GGT GTC ACG ATC CTG	787
Ser Lys Glu Ile Glu Lys Gln Phe Lys Lys Leu Gly Val Thr Ile Leu	
215 220 225 230	
ACC GCC ACG AAG GTC GAG TCC ATC GCC GAT GGC GGG TCG CAG GTC ACC	835
Thr Ala Thr Lys Val Glu Ser Ile Ala Asp Gly Gly Ser Gln Val Thr	

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	235	240	245	
GTG ACC GTC ACC AAG GAC GGC GTG GCG CAA GAG CTT AAG GCG GAA AAG				883
Val Thr Val Thr Lys Asp Gly Val Ala Gln Glu Leu Lys Ala Glu Lys	250	255	260	
GTG TTG CAG GCC ATC GGA TTT GCG CCC AAC GTC GAA GGG TAC GGG CTG				931
Val Leu Gln Ala Ile Gly Phe Ala Pro Asn Val Glu Gly Tyr Gly Leu	265	270	275	
GAC AAG GCA GGC GTC GCG CTG ACC GAC CGC AAG GCT ATC GGT GTC GAC				979
Asp Lys Ala Gly Val Ala Leu Thr Asp Arg Lys Ala Ile Gly Val Asp	280	285	290	
GAC TAC ATG CGT ACC AAC GTG GGC CAC ATC TAC GCT ATC GGC GAT GTC				1027
Asp Tyr Met Arg Thr Asn Val Gly His Ile Tyr Ala Ile Gly Asp Val	295	300	305	310
AAT GGA TTA CTG CAG CTG GCG CAC GTC GCC GAG GCA CAA GGC GTG GTA				1075
Asn Gly Leu Leu Gln Leu Ala His Val Ala Glu Ala Gln Gly Val Val	315	320	325	
GCC GCC GAA ACC ATT GCC GGT GCA GAG ACT TTG ACG CTG GGC GAC CAT				1123
Ala Ala Glu Thr Ile Ala Gly Ala Glu Thr Leu Thr Leu Gly Asp His	330	335	340	
CGG ATG TTG CCG CGC GCG ACG TTC TGT CAG CCA AAC GTT GCC AGC TTC				1171
Arg Met Leu Pro Arg Ala Thr Phe Cys Gln Pro Asn Val Ala Ser Phe	345	350	355	
GGG CTC ACC GAG CAG CAA GCC CGC AAC GAA GGT TAC GAC GTG GTG GTG				1219
Gly Leu Thr Glu Gln Gln Ala Arg Asn Glu Gly Tyr Asp Val Val Val	360	365	370	
GCC AAG TTC CCG TTC ACG GCC AAC GCC AAG GCG CAC GGC GTG GGT GAC				1267
Ala Lys Phe Pro Phe Thr Ala Asn Ala Lys Ala His Gly Val Gly Asp	375	380	385	390
CCC AGT GGG TTC GTC AAG CTG GTG GCC GAC GCC AAG CAC GGC GAG CTA				1315
Pro Ser Gly Phe Val Lys Leu Val Ala Asp Ala Lys His Gly Glu Leu	395	400	405	
CTG GGT GGG CAC CTG GTC GGC CAC GAC GTG GCC GAG CTG CTG CCG GAG				1363
Leu Gly Gly His Leu Val Gly His Asp Val Ala Glu Leu Leu Pro Glu	410	415	420	
CTC ACG CTG GCG CAG AGG TGG GAC CTG ACC GCC AGC GAG CTG GCT CGC				1411
Leu Thr Leu Ala Gln Arg Trp Asp Leu Thr Ala Ser Glu Leu Ala Arg	425	430	435	
AAC GTC CAC ACC CAC CCA ACG ATG TCT GAG GCG CTG CAG GAG TGC TTC				1459
Asn Val His Thr His Pro Thr Met Ser Glu Ala Leu Gln Glu Cys Phe	440	445	450	
CAC GGC CTG GTT GGC CAC ATG ATC AAT T TCTGAGCGGC TCATGACGAG GCGCG				1512
His Gly Leu Val Gly His Met Ile Asn Phe	455	460		



CGAGCACTGA CACCCCCCAG ATCATCATGG GTGCCATCGG TGGTGTGG

1560

## (2) INFORMATION FOR SEQ ID NO: 62:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 464 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 62:

```

Met Thr His Tyr Asp Val Val Val Leu Gly Ala Gly Pro Gly Gly Tyr
 1          5          10          15
Val Ala Ala Ile Arg Ala Ala Gln Leu Gly Leu Ser Thr Ala Ile Val
          20          25          30
Glu Pro Lys Tyr Trp Gly Gly Val Cys Leu Asn Val Gly Cys Ile Pro
          35          40          45
Ser Lys Ala Leu Leu Arg Asn Ala Glu Leu Val His Ile Phe Thr Lys
          50          55          60
Asp Ala Lys Ala Phe Gly Ile Ser Gly Glu Val Thr Phe Asp Tyr Gly
65          70          75          80
Ile Ala Tyr Asp Arg Ser Arg Lys Val Ala Glu Gly Arg Val Ala Gly
          85          90          95
Val His Phe Leu Met Lys Lys Asn Lys Ile Thr Glu Ile His Gly Tyr
          100          105          110
Gly Thr Phe Ala Asp Ala Asn Thr Leu Leu Val Asp Leu Asn Asp Gly
          115          120          125
Gly Thr Glu Ser Val Thr Phe Asp Asn Ala Ile Ile Ala Thr Gly Ser
          130          135          140
Ser Thr Arg Leu Val Pro Gly Thr Ser Leu Ser Ala Asn Val Val Thr
          145          150          155          160
Tyr Glu Glu Gln Ile Leu Ser Arg Glu Leu Pro Lys Ser Ile Ile Ile
          165          170          175
Ala Gly Ala Gly Ala Ile Gly Met Glu Phe Gly Tyr Val Leu Lys Asn
          180          185          190
Tyr Gly Val Asp Val Thr Ile Val Glu Phe Leu Pro Arg Ala Leu Pro
          195          200          205
Asn Glu Asp Ala Asp Val Ser Lys Glu Ile Glu Lys Gln Phe Lys Lys
          210          215          220
Leu Gly Val Thr Ile Leu Thr Ala Thr Lys Val Glu Ser Ile Ala Asp
          225          230          235          240
Gly Gly Ser Gln Val Thr Val Thr Val Thr Lys Asp Gly Val Ala Gln
          245          250          255
Glu Leu Lys Ala Glu Lys Val Leu Gln Ala Ile Gly Phe Ala Pro Asn
          260          265          270
Val Glu Gly Tyr Gly Leu Asp Lys Ala Gly Val Ala Leu Thr Asp Arg
          275          280          285
Lys Ala Ile Gly Val Asp Asp Tyr Met Arg Thr Asn Val Gly His Ile
          290          295          300
Tyr Ala Ile Gly Asp Val Asn Gly Leu Leu Gln Leu Ala His Val Ala
          305          310          315          320
Glu Ala Gln Gly Val Val Ala Ala Glu Thr Ile Ala Gly Ala Glu Thr
          325          330          335
Leu Thr Leu Gly Asp His Arg Met Leu Pro Arg Ala Thr Phe Cys Gln

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340	345	350
Pro Asn Val Ala Ser Phe Gly Leu Thr Glu Gln Gln Ala Arg Asn Glu		
355	360	365
Gly Tyr Asp Val Val Val Ala Lys Phe Pro Phe Thr Ala Asn Ala Lys		
370	375	380
Ala His Gly Val Gly Asp Pro Ser Gly Phe Val Lys Leu Val Ala Asp		
385	390	395
Ala Lys His Gly Glu Leu Leu Gly Gly His Leu Val Gly His Asp Val		
405	410	415
Ala Glu Leu Leu Pro Glu Leu Thr Leu Ala Gln Arg Trp Asp Leu Thr		
420	425	430
Ala Ser Glu Leu Ala Arg Asn Val His Thr His Pro Thr Met Ser Glu		
435	440	445
Ala Leu Gln Glu Cys Phe His Gly Leu Val Gly His Met Ile Asn Phe		
450	455	460

## (2) INFORMATION FOR SEQ ID NO: 63:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 550 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 101...490
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 63:

GGCCCGGCTC GCGGCCGCC TGCAGGAAAA GAAGGCCTGC CCAGGCCCAG ACTCAGCCGA	60
GTAGTCACCC AGTACCCAC ACCAGGAAGG ACCGCCCATC ATG GCA AAG CTC TCC	115
Met Ala Lys Leu Ser	
1 5	
ACC GAC GAA CTG CTG GAC GCG TTC AAG GAA ATG ACC CTG TTG GAG CTC	163
Thr Asp Glu Leu Leu Asp Ala Phe Lys Glu Met Thr Leu Leu Glu Leu	
10 15 20	
TCC GAC TTC GTC AAG AAG TTC GAG GAG ACC TTC GAG GTC ACC GCC GCC	211
Ser Asp Phe Val Lys Lys Phe Glu Glu Thr Phe Glu Val Thr Ala Ala	
25 30 35	
GCT CCA GTC GCC GTC GCC GCC GCC GGT GCC GCC CCG GCC GGT GCC GCC	259
Ala Pro Val Ala Val Ala Ala Ala Gly Ala Ala Pro Ala Gly Ala Ala	
40 45 50	
GTC GAG GCT GCC GAG GAG CAG TCC GAG TTC GAC GTG ATC CTT GAG GCC	307
Val Glu Ala Ala Glu Glu Gln Ser Glu Phe Asp Val Ile Leu Glu Ala	
55 60 65	
GCC GGC GAC AAG AAG ATC GGC GTC ATC AAG GTG GTC CGG GAG ATC GTT	355
Ala Gly Asp Lys Lys Ile Gly Val Ile Lys Val Val Arg Glu Ile Val	
70 75 80 85	

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TCC GGC CTG GGC CTC AAG GAG GCC AAG GAC CTG GTC GAC GGC GCG CCC      403
Ser Gly Leu Gly Leu Lys Glu Ala Lys Asp Leu Val Asp Gly Ala Pro
          90                      95                      100

AAG CCG CTG CTG GAG AAG GTC GCC AAG GAG GCC GCC GAC GAG GCC AAG      451
Lys Pro Leu Leu Glu Lys Val Ala Lys Glu Ala Ala Asp Glu Ala Lys
          105                      110                      115

GCC AAG CTG GAG GCC GCC GGC GCC ACC GTC ACC GTC AAG TAGCTCTGCC CA    502
Ala Lys Leu Glu Ala Ala Gly Ala Thr Val Thr Val Lys
          120                      125                      130

GCGTGTCTCTT TTGCGTCTGC TCGGCCCGTA GCGAACACTG CGCCCGCT              550

```

(2) INFORMATION FOR SEQ ID NO: 64:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 130 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 64:

```

Met Ala Lys Leu Ser Thr Asp Glu Leu Leu Asp Ala Phe Lys Glu Met
  1             5             10             15

Thr Leu Leu Glu Leu Ser Asp Phe Val Lys Lys Phe Glu Glu Thr Phe
          20             25             30

Glu Val Thr Ala Ala Ala Pro Val Ala Val Ala Ala Ala Gly Ala Ala
          35             40             45

Pro Ala Gly Ala Ala Val Glu Ala Ala Glu Glu Gln Ser Glu Phe Asp
          50             55             60

Val Ile Leu Glu Ala Ala Gly Asp Lys Lys Ile Gly Val Ile Lys Val
          65             70             75             80

Val Arg Glu Ile Val Ser Gly Leu Gly Leu Lys Glu Ala Lys Asp Leu
          85             90             95

Val Asp Gly Ala Pro Lys Pro Leu Leu Glu Lys Val Ala Lys Glu Ala
          100            105            110

Ala Asp Glu Ala Lys Ala Lys Leu Glu Ala Ala Gly Ala Thr Val Thr
          115            120            125

Val Lys
          130

```

(2) INFORMATION FOR SEQ ID NO: 65:

- (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 900 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence  
 (B) LOCATION: 87...770  
 (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 65:

TGAACGCCAT CGGGTCCAAC GAACGCAGCG CTACCTGATC ACCACCGGGT CTGTTAGGGC	60
TCTTCCCCAG GTCGTACAGT CGGGCC ATG GCC ATT GAG GTT TCG GTG TTG CGG	113
Met Ala Ile Glu Val Ser Val Leu Arg	
1 5	
GTT TTC ACC GAT TCA GAC GGG AAT TTC GGT AAT CCG CTG GGG GTG ATC	161
Val Phe Thr Asp Ser Asp Gly Asn Phe Gly Asn Pro Leu Gly Val Ile	
10 15 20 25	
AAC GCC AGC AAG GTC GAA CAC CGC GAC AGG CAG CAG CTG GCA GCC CAA	209
Asn Ala Ser Lys Val Glu His Arg Asp Arg Gln Gln Leu Ala Ala Gln	
30 35 40	
TCG GGC TAC AGC GAA ACC ATA TTC GTC GAT CTT CCC AGC CCC GGC TCA	257
Ser Gly Tyr Ser Glu Thr Ile Phe Val Asp Leu Pro Ser Pro Gly Ser	
45 50 55	
ACC ACC GCA CAC GCC ACC ATC CAT ACT CCC CGC ACC GAA ATT CCG TTC	305
Thr Thr Ala His Ala Thr Ile His Thr Pro Arg Thr Glu Ile Pro Phe	
60 65 70	
GCC GGA CAC CCG ACC GTG GGA GCG TCC TGG TGG CTG CGC GAG AGG GGG	353
Ala Gly His Pro Thr Val Gly Ala Ser Trp Trp Leu Arg Glu Arg Gly	
75 80 85	
ACG CCA ATT AAC ACG CTG CAG GTG CCG GCC GGC ATC GTC CAG GTG AGC	401
Thr Pro Ile Asn Thr Leu Gln Val Pro Ala Gly Ile Val Gln Val Ser	
90 95 100 105	
TAC CAC GGT GAT CTC ACC GCC ATC AGC GCC CGC TCG GAA TGG GCA CCC	449
Tyr His Gly Asp Leu Thr Ala Ile Ser Ala Arg Ser Glu Trp Ala Pro	
110 115 120	
GAG TTC GCC ATC CAC GAC CTG GAT TCA CTT GAT GCG CTT GCC GCC GCC	497
Glu Phe Ala Ile His Asp Leu Asp Ser Leu Asp Ala Leu Ala Ala Ala	
125 130 135	
GAC CCC GCC GAC TTT CCG GAC GAC ATC GCG CAC TAC CTC TGG ACC TGG	545
Asp Pro Ala Asp Phe Pro Asp Asp Ile Ala His Tyr Leu Trp Thr Trp	
140 145 150	
ACC GAC CGC TCC GCT GGC TCG CTG CGC GCC CGC ATG TTT GCC GCC AAC	593
Thr Asp Arg Ser Ala Gly Ser Leu Arg Ala Arg Met Phe Ala Ala Asn	

155	160	165	
TTG GGC GTC ACC GAA GAC GAA GCG ACC GGT GCC GCG GCC ATC CGG ATT			641
Leu Gly Val Thr Glu Asp Glu Ala Thr Gly Ala Ala Ala Ile Arg Il			
170	175	180	185
ACC GAT TAC CTC AGC CGT GAC CTC ACC ATC ACC CAG GGC AAA GGA TCG			689
Thr Asp Tyr Leu Ser Arg Asp Leu Thr Ile Thr Gln Gly Lys Gly Ser			
	190	195	200
TTG ATC CAC ACC ACC TGG AGT CCC GAG GGC TGG GTT CGG GTA GCC GGC			737
Leu Ile His Thr Thr Trp Ser Pro Glu Gly Trp Val Arg Val Ala Gly			
	205	210	215
CGA GTT GTC AGC GAC GGT GTG GCA CAA CTC GAC TGACGTAGAG CTCAGCGCTG			790
Arg Val Val Ser Asp Gly Val Ala Gln Leu Asp			
	220	225	
CCGATGCAAC ACGGCGGCAA GGTGATCCTG CAGGGGTTGC CCGACCGCGC GCATCTGCAA			850
CGAGTACGAA AGCTCGTCGC CGTCGATGCG GTAGGAACGG TCAAGGGCGG			900

## (2) INFORMATION FOR SEQ ID NO: 66:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 228 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 66:

Met	Ala	Ile	Glu	Val	Ser	Val	Leu	Arg	Val	Phe	Thr	Asp	Ser	Asp	Gly
1				5					10						15
Asn	Phe	Gly	Asn	Pro	Leu	Gly	Val	Ile	Asn	Ala	Ser	Lys	Val	Glu	His
		20					25						30		
Arg	Asp	Arg	Gln	Gln	Leu	Ala	Ala	Gln	Ser	Gly	Tyr	Ser	Glu	Thr	Ile
		35					40						45		
Phe	Val	Asp	Leu	Pro	Ser	Pro	Gly	Ser	Thr	Thr	Ala	His	Ala	Thr	Ile
	50					55					60				
His	Thr	Pro	Arg	Thr	Glu	Ile	Pro	Phe	Ala	Gly	His	Pro	Thr	Val	Gly
65					70					75					80
Ala	Ser	Trp	Trp	Leu	Arg	Glu	Arg	Gly	Thr	Pro	Ile	Asn	Thr	Leu	Gln
				85					90					95	
Val	Pro	Ala	Gly	Ile	Val	Gln	Val	Ser	Tyr	His	Gly	Asp	Leu	Thr	Ala
		100						105					110		
Ile	Ser	Ala	Arg	Ser	Glu	Trp	Ala	Pro	Glu	Phe	Ala	Ile	His	Asp	Leu

115	120	125
Asp Ser Leu Asp Ala Leu	Ala Ala Ala Asp Pro	Ala Asp Phe Pro Asp
130	135	140
Asp Ile Ala His Tyr Leu	Trp Thr Trp Thr Asp	Arg Ser Ala Gly Ser
145	150	155 160
Leu Arg Ala Arg Met Phe	Ala Ala Asn Leu Gly	Val Thr Glu Asp Glu
	165	170 175
Ala Thr Gly Ala Ala Ala	Ile Arg Ile Thr Asp	Tyr Leu Ser Arg Asp
	180	185 190
Leu Thr Ile Thr Gln Gly	Lys Gly Ser Leu Ile	His Thr Thr Trp Ser
	195	200 205
Pro Glu Gly Trp Val Arg	Val Ala Gly Arg Val	Val Ser Asp Gly Val
	210	215 220
Ala Gln Leu Asp		
225		

## (2) INFORMATION FOR SEQ ID NO: 67:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 500 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 49...465
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 67:

GTTTGTGGTG TCGGTGGTCT GGGGGGCGCC AACTGGGATT CGGTTGGG GTG GGT GCA	57
Met Gly Ala	
1	
GGT CCG GCG ATG GGC ATC GGA GGT GTG GGT GGT TTG GGT GGG GCC GGT	105
Gly Pro Ala Met Gly Ile Gly Gly Val Gly Gly Leu Gly Gly Ala Gly	
5 10 15	
TCG GGT CCG GCG ATG GGC ATG GGG GGT GTG GGT GGT TTG GGT GGG GCC	153
Ser Gly Pro Ala Met Gly Met Gly Gly Val Gly Gly Leu Gly Gly Ala	
20 25 30 35	
GGT TCG GGT CCG GCG ATG GGC ATG GGG GGT GTG GGT GGT TTA GAT GCG	201
Gly Ser Gly Pro Ala Met Gly Met Gly Gly Val Gly Gly Leu Asp Ala	
40 45 50	
GCC GGT TCC GGC GAG GGC GGC TCT CCT GCG GCG ATC GGC ATC GGA GTT	249

Ala Gly Ser Gly Glu Gly Gly Ser Pro Ala Ala Ile Gly Ile Gly Val	
55 60 65	
GGC GGA GGC GGA GGT GGG GGT GGG GGT GGC GGC GGC GGG GCC GAC ACG	297
Gly Gly Gly Gly Gly Gly Gly Gly Gly Gly Gly Gly Gly Gly Ala Asp Thr	
70 75 80	
AAC CGC TCC GAC AGG TCG TCG GAC GTC GGG GGC GGA GTC TGG CCG TTG	345
Asn Arg Ser Asp Arg Ser Ser Asp Val Gly Gly Gly Val Trp Pro Leu	
85 90 95	
GGC TTC GGT AGG TTT GCC GAT GCG GGC GCC GGC GGA AAC GAA GCA CTG	393
Gly Phe Gly Arg Phe Ala Asp Ala Gly Ala Gly Gly Asn Glu Ala Leu	
100 105 110 115	
GGG TCG AAG AAC GGC TGC GCT GCC ATA TCG TCC GGA GCT TCC ATA CCT	441
Gly Ser Lys Asn Gly Cys Ala Ala Ile Ser Ser Gly Ala Ser Ile Pro	
120 125 130	
TCG TGC GGC CGG AAG AGC TTG TCG TAGTCGGCCG CCATGACAAC CTCTCAGAGT	495
Ser Cys Gly Arg Lys Ser Leu Ser	
135	
GCGCT	500

## (2) INFORMATION FOR SEQ ID NO: 68:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 139 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 68:

Met Gly Ala Gly Pro Ala Met Gly Ile Gly Gly Val Gly Gly Leu Gly	
1 5 10 15	
Gly Ala Gly Ser Gly Pro Ala Met Gly Met Gly Gly Val Gly Gly Leu	
20 25 30	
Gly Gly Ala Gly Ser Gly Pro Ala Met Gly Met Gly Gly Val Gly Gly	
35 40 45	
Leu Asp Ala Ala Gly Ser Gly Glu Gly Gly Ser Pro Ala Ala Ile Gly	
50 55 60	
Ile Gly Val Gly Gly Gly Gly Gly Gly Gly Gly Gly Gly Gly Gly Gly	
65 70 75 80	
Ala Asp Thr Asn Arg Ser Asp Arg Ser Ser Asp Val Gly Gly Gly Val	
85 90 95	
Trp Pro Leu Gly Phe Gly Arg Phe Ala Asp Ala Gly Ala Gly Gly Asn	

100	105	110
Glu Ala Leu Gly Ser Lys Asn Gly Cys Ala Ala Ile Ser Ser Gly Ala		
115	120	125
Ser Ile Pro Ser Cys Gly Arg Lys Ser Leu Ser		
130	135	

## (2) INFORMATION FOR SEQ ID NO: 69:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2050 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 22...2019
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 69:

AGCGCACTCT GAGAGGTTGT C ATG GCG GCC GAC TAC GAC AAG CTC TTC CGG	51
Met Ala Ala Asp Tyr Asp Lys Leu Phe Arg	
1 5 10	
CCG CAC GAA GGT ATG GAA GCT CCG GAC GAT ATG GCA GCG CAG CCG TTC	99
Pro His Glu Gly Met Glu Ala Pro Asp Asp Met Ala Ala Gln Pro Phe	
15 20 25	
TTC GAC CCC AGT GCT TCG TTT CCG CCG GCG CCC GCA TCG GCA AAC CTA	147
Phe Asp Pro Ser Ala Ser Phe Pro Pro Ala Pro Ala Ser Ala Asn Leu	
30 35 40	
CCG AAG CCC AAC GGC CAG ACT CCG CCC CCG ACG TCC GAC GAC CTG TCG	195
Pro Lys Pro Asn Gly Gln Thr Pro Pro Pro Thr Ser Asp Asp Leu Ser	
45 50 55	
GAG CGG TTC GTG TCG GCC CCG CCG CCG CCA CCC CCA CCC CCA CCT CCG	243
Glu Arg Phe Val Ser Ala Pro Pro Pro Pro Pro Pro Pro Pro Pro	
60 65 70	
CCT CCG CCA ACT CCG ATG CCG ATC GCC GCA GGA GAG CCG CCC TCG CCG	291
Pro Pro Pro Thr Pro Met Pro Ile Ala Ala Gly Glu Pro Pro Ser Pro	
75 80 85 90	
GAA CCG GCC GCA TCT AAA CCA CCC ACA CCC CCC ATG CCC ATC GCC GGA	339
Glu Pro Ala Ala Ser Lys Pro Pro Thr Pro Pro Met Pro Ile Ala Gly	
95 100 105	
CCC GAA CCG GCC CCA CCC AAA CCA CCC ACA CCC CCC ATG CCC ATC GCC	387
Pro Glu Pro Ala Pro Pro Lys Pro Pro Thr Pro Pro Met Pro Ile Ala	
110 115 120	



GGA CCC GAA CCG GCC CCA CCC AAA CCA CCC ACA CCT CCG ATG CCC ATC	435
Gly Pro Glu Pro Ala Pro Pro Lys Pro Pro Thr Pro Pro Met Pro Ile	
125 130 135	
GCC GGA CCT GCA CCC ACC CCA ACC GAA TCC CAG TTG GCG CCC CCC AGA	483
Ala Gly Pro Ala Pro Thr Thr Glu Ser Gln Leu Ala Pro Pro Arg	
140 145 150	
CCA CCG ACA CCA CAA ACG CCA ACC GGA GCG CCG CAG CAA CCG GAA TCA	531
Pro Pro Thr Pro Gln Thr Pro Thr Gly Ala Pro Gln Gln Pro Glu Ser	
155 160 165 170	
CCG GCG CCC CAC GTA CCC TCG CAC GGG CCA CAT CAA CCC CGG CGC ACC	579
Pro Ala Pro His Val Pro Ser His Gly Pro His Gln Pro Arg Arg Thr	
175 180 185	
GCA CCA GCA CCG CCC TGG GCA AAG ATG CCA ATC GGC GAA CCC CCG CCC	627
Ala Pro Ala Pro Pro Trp Ala Lys Met Pro Ile Gly Glu Pro Pro Pro	
190 195 200	
GCT CCG TCC AGA CCG TCT GCG TCC CCG GCC GAA CCA CCG ACC CGG CCT	675
Ala Pro Ser Arg Pro Ser Ala Ser Pro Ala Glu Pro Pro Thr Arg Pro	
205 210 215	
GCC CCC CAA CAC TCC CGA CGT GCG CGC CGG GGT CAC CGC TAT CGC ACA	723
Ala Pro Gln His Ser Arg Arg Ala Arg Arg Gly His Arg Tyr Arg Thr	
220 225 230	
GAC ACC GAA CGA AAC GTC GGG AAG GTA GCA ACT GGT CCA TCC ATC CAG	771
Asp Thr Glu Arg Asn Val Gly Lys Val Ala Thr Gly Pro Ser Ile Gln	
235 240 245 250	
GCG CGG CTG CGG GCA GAG GAA GCA TCC GGC GCG CAG CTC GCC CCC GGA	819
Ala Arg Leu Arg Ala Glu Glu Ala Ser Gly Ala Gln Leu Ala Pro Gly	
255 260 265	
ACG GAG CCC TCG CCA GCG CCG TTG GGC CAA CCG AGA TCG TAT CTG GCT	867
Thr Glu Pro Ser Pro Ala Pro Leu Gly Gln Pro Arg Ser Tyr Leu Ala	
270 275 280	
CCG CCC ACC CGC CCC GCG CCG ACA GAA CCT CCC CCC AGC CCC TCG CCG	915
Pro Pro Thr Arg Pro Ala Pro Thr Glu Pro Pro Pro Ser Pro Ser Pro	
285 290 295	
CAG CGC AAC TCC GGT CGG CGT GCC GAG CGA CGC GTC CAC CCC GAT TTA	963
Gln Arg Asn Ser Gly Arg Arg Ala Glu Arg Arg Val His Pro Asp Leu	
300 305 310	
GCC GCC CAA CAT GCC GCG GCG CAA CCT GAT TCA ATT ACG GCC GCA ACC	1011
Ala Ala Gln His Ala Ala Ala Gln Pro Asp Ser Ile Thr Ala Ala Thr	
315 320 325 330	
ACT GGC GGT CGT CGC CGC AAG CGT GCA GCG CCG GAT CTC GAC GCG ACA	1059
Thr Gly Gly Arg Arg Lys Arg Ala Ala Pro Asp Leu Asp Ala Thr	
335 340 345	
CAG AAA TCC TTA AGG CCG GCG GCC AAG GGG CCG AAG GTG AAG AAG GTG	1107
Gln Lys Ser Leu Arg Pro Ala Ala Lys Gly Pro Lys Val Lys Lys Val	

350	355	360	
AAG CCC CAG AAA CCG AAG GCC ACG AAG CCG CCC AAA GTG GTG TCG CAG Lys Pro Gln Lys Pro Lys Ala Thr Lys Pro Pro Lys Val Val Ser Gln 365 370 375			1155
CGC GGC TGG CGA CAT TGG GTG CAT GCG TTG ACG CGA ATC AAC CTG GGC Arg Gly Trp Arg His Trp Val His Ala Leu Thr Arg Ile Asn Leu Gly 380 385 390			1203
CTG TCA CCC GAC GAG AAG TAC GAG CTG GAC CTG CAC GCT CGA GTC CGC Leu Ser Pro Asp Glu Lys Tyr Glu Leu Asp Leu His Ala Arg Val Arg 395 400 405 410			1251
CGC AAT CCC CGC GGG TCG TAT CAG ATC GCC GTC GTC GGT CTC AAA GGT Arg Asn Pro Arg Gly Ser Tyr Gln Ile Ala Val Val Gly Leu Lys Gly 415 420 425			1299
GGG GCT GGC AAA ACC ACG CTG ACA GCA GCG TTG GGG TCG ACG TTG GCT Gly Ala Gly Lys Thr Thr Leu Thr Ala Ala Leu Gly Ser Thr Leu Ala 430 435 440			1347
CAG GTG CGG GCC GAC CGG ATC CTG GCT CTA GAC GCG GAT CCA GGC GCC Gln Val Arg Ala Asp Arg Ile Leu Ala Leu Asp Ala Asp Pro Gly Ala 445 450 455			1395
GGA AAC CTC GCC GAT CGG GTA GGG CGA CAA TCG GGC GCG ACC ATC GCT Gly Asn Leu Ala Asp Arg Val Gly Arg Gln Ser Gly Ala Thr Ile Ala 460 465 470			1443
GAT GTG CTT GCA GAA AAA GAG CTG TCG CAC TAC AAC GAC ATC CGC GCA Asp Val Leu Ala Glu Lys Glu Leu Ser His Tyr Asn Asp Ile Arg Ala 475 480 485 490			1491
CAC ACT AGC GTC AAT GCG GTC AAT CTG GAA GTG CTG CCG GCA CCG GAA His Thr Ser Val Asn Ala Val Asn Leu Glu Val Leu Pro Ala Pro Glu 495 500 505			1539
TAC AGC TCG GCG CAG CGC GCG CTC AGC GAC GCC GAC TGG CAT TTC ATC Tyr Ser Ser Ala Gln Arg Ala Leu Ser Asp Ala Asp Trp His Phe Ile 510 515 520			1587
GCC GAT CCT GCG TCG AGG TTT TAC AAC CTC GTC TTG GCT GAT TGT GGG Ala Asp Pro Ala Ser Arg Phe Tyr Asn Leu Val Leu Ala Asp Cys Gly 525 530 535			1635
GCC GGC TTC TTC GAC CCG CTG ACC CGC GGC GTG CTG TCC ACG GTG TCC Ala Gly Phe Phe Asp Pro Leu Thr Arg Gly Val Leu Ser Thr Val Ser 540 545 550			1683
GGT GTC GTG GTC GTG GCA AGT GTC TCA ATC GAC GGC GCA CAA CAG GCG Gly Val Val Val Val Ala Ser Val Ser Ile Asp Gly Ala Gln Gln Ala 555 560 565 570			1731
TCG GTC GCG TTG GAC TGG TTG CGC AAC AAC GGT TAC CAA GAT TTG GCG Ser Val Ala Leu Asp Trp Leu Arg Asn Asn Gly Tyr Gln Asp Leu Ala 575 580 585			1779

AGC CGC GCA TGC GTG GTC ATC AAT CAC ATC ATG CCG GGA GAA CCC AAT	1827
Ser Arg Ala Cys Val Val Ile Asn His Ile Met Pro Gly Glu Pro Asn	
590 595 600	
GTC GCA GTT AAA GAC CTG GTG CGG CAT TTC GAA CAG CAA GTT CAA CCC	1875
Val Ala Val Lys Asp Leu Val Arg His Phe Glu Gln Gln Val Gln Pro	
605 610 615	
GGC CGG GTC GTG GTC ATG CCG TGG GAC AGG CAC ATT GCG GCC GGA ACC	1923
Gly Arg Val Val Val Met Pro Trp Asp Arg His Ile Ala Ala Gly Thr	
620 625 630	
GAG ATT TCA CTC GAC TTG CTC GAC CCT ATC TAC AAG CGC AAG GTC CTC	1971
Glu Ile Ser Leu Asp Leu Leu Asp Pro Ile Tyr Lys Arg Lys Val Leu	
635 640 645 650	
GAA TTG GCC GCA GCG CTA TCC GAC GAT TTC GAG AGG GCT GGA CGT CGT T	2020
Glu Leu Ala Ala Ala Leu Ser Asp Asp Phe Glu Arg Ala Gly Arg Arg	
655 660 665	
GAGCGCACCT GCTGTTGCTG CTGGTCCTAC	2050

## (2) INFORMATION FOR SEQ ID NO: 70:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 666 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 70:

Met	Ala	Ala	Asp	Tyr	Asp	Lys	Leu	Phe	Arg	Pro	His	Glu	Gly	Met	Glu
1				5					10					15	
Ala	Pro	Asp	Asp	Met	Ala	Ala	Gln	Pro	Phe	Phe	Asp	Pro	Ser	Ala	Ser
		20						25					30		
Phe	Pro	Pro	Ala	Pro	Ala	Ser	Ala	Asn	Leu	Pro	Lys	Pro	Asn	Gly	Gln
		35					40					45			
Thr	Pro	Pro	Pro	Thr	Ser	Asp	Asp	Leu	Ser	Glu	Arg	Phe	Val	Ser	Ala
	50					55				60					
Pro	Pro	Pro	Pro	Pro	Pro	Pro	Pro	Pro	Pro	Pro	Pro	Pro	Thr	Pro	Met
	65					70				75				80	
Pro	Ile	Ala	Ala	Gly	Glu	Pro	Pro	Ser	Pro	Glu	Pro	Ala	Ala	Ser	Lys
				85					90					95	
Pro	Pro	Thr	Pro	Pro	Met	Pro	Ile	Ala	Gly	Pro	Glu	Pro	Ala	Pro	Pro
		100						105					110		
Lys	Pro	Pro	Thr	Pro	Pro	Met	Pro	Ile	Ala	Gly	Pro	Glu	Pro	Ala	Pro

115	120	125
Pro Lys Pro Pro Thr Pro Pro Met Pro Ile Ala Gly Pro Ala Pro Thr 130 135 140		
Pro Thr Glu Ser Gln Leu Ala Pro Pro Arg Pro Pro Thr Pro Gln Thr 145 150 155 160		
Pro Thr Gly Ala Pro Gln Gln Pro Glu Ser Pro Ala Pro His Val Pro 165 170 175		
Ser His Gly Pro His Gln Pro Arg Arg Thr Ala Pro Ala Pro Pro Trp 180 185 190		
Ala Lys Met Pro Ile Gly Glu Pro Pro Pro Ala Pro Ser Arg Pro Ser 195 200 205		
Ala Ser Pro Ala Glu Pro Pro Thr Arg Pro Ala Pro Gln His Ser Arg 210 215 220		
Arg Ala Arg Arg Gly His Arg Tyr Arg Thr Asp Thr Glu Arg Asn Val 225 230 235 240		
Gly Lys Val Ala Thr Gly Pro Ser Ile Gln Ala Arg Leu Arg Ala Glu 245 250 255		
Glu Ala Ser Gly Ala Gln Leu Ala Pro Gly Thr Glu Pro Ser Pro Ala 260 265 270		
Pro Leu Gly Gln Pro Arg Ser Tyr Leu Ala Pro Pro Thr Arg Pro Ala 275 280 285		
Pro Thr Glu Pro Pro Pro Ser Pro Ser Pro Gln Arg Asn Ser Gly Arg 290 295 300		
Arg Ala Glu Arg Arg Val His Pro Asp Leu Ala Ala Gln His Ala Ala 305 310 315 320		
Ala Gln Pro Asp Ser Ile Thr Ala Ala Thr Thr Gly Gly Arg Arg Arg 325 330 335		
Lys Arg Ala Ala Pro Asp Leu Asp Ala Thr Gln Lys Ser Leu Arg Pro 340 345 350		
Ala Ala Lys Gly Pro Lys Val Lys Lys Val Lys Pro Gln Lys Pro Lys 355 360 365		
Ala Thr Lys Pro Pro Lys Val Val Ser Gln Arg Gly Trp Arg His Trp 370 375 380		
Val His Ala Leu Thr Arg Ile Asn Leu Gly Leu Ser Pro Asp Glu Lys 385 390 395 400		
Tyr Glu Leu Asp Leu His Ala Arg Val Arg Arg Asn Pro Arg Gly Ser 405 410 415		
Tyr Gln Ile Ala Val Val Gly Leu Lys Gly Gly Ala Gly Lys Thr Thr 420 425 430		

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Leu Thr Ala Ala Leu Gly Ser Thr Leu Ala Gln Val Arg Ala Asp Arg
    435                      440                      445

Ile Leu Ala Leu Asp Ala Asp Pro Gly Ala Gly Asn Leu Ala Asp Arg
    450                      455                      460

Val Gly Arg Gln Ser Gly Ala Thr Ile Ala Asp Val Leu Ala Glu Lys
    465                      470                      475                      480

Glu Leu Ser His Tyr Asn Asp Ile Arg Ala His Thr Ser Val Asn Ala
                485                      490                      495

Val Asn Leu Glu Val Leu Pro Ala Pro Glu Tyr Ser Ser Ala Gln Arg
                500                      505                      510

Ala Leu Ser Asp Ala Asp Trp His Phe Ile Ala Asp Pro Ala Ser Arg
    515                      520                      525

Phe Tyr Asn Leu Val Leu Ala Asp Cys Gly Ala Gly Phe Phe Asp Pro
    530                      535                      540

Leu Thr Arg Gly Val Leu Ser Thr Val Ser Gly Val Val Val Val Ala
    545                      550                      555                      560

Ser Val Ser Ile Asp Gly Ala Gln Gln Ala Ser Val Ala Leu Asp Trp
                565                      570                      575

Leu Arg Asn Asn Gly Tyr Gln Asp Leu Ala Ser Arg Ala Cys Val Val
                580                      585                      590

Ile Asn His Ile Met Pro Gly Glu Pro Asn Val Ala Val Lys Asp Leu
    595                      600                      605

Val Arg His Phe Glu Gln Gln Val Gln Pro Gly Arg Val Val Val Met
    610                      615                      620

Pro Trp Asp Arg His Ile Ala Ala Gly Thr Glu Ile Ser Leu Asp Leu
    625                      630                      635                      640

Leu Asp Pro Ile Tyr Lys Arg Lys Val Leu Glu Leu Ala Ala Ala Leu
                645                      650                      655

Ser Asp Asp Phe Glu Arg Ala Gly Arg Arg
    660                      665

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## (2) INFORMATION FOR SEQ ID NO: 71:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1890 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 79...1851

## (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 71:

GCAGCGATGA GGAGGAGCGG CGCCAACGGC CCGCGCCGGC GACGATGCAA AGCGCAGCGA	60
TGAGGAGGAG CGGCGCGC ATG ACT GCT GAA CCG GAA GTA CGG ACG CTG CGC	111
Met Thr Ala Glu Pro Glu Val Arg Thr Leu Arg	
1 5 10	
GAG GTT GTG CTG GAC CAG CTC GGC ACT GCT GAA TCG CGT GCG TAC AAG	159
Glu Val Val Leu Asp Gln Leu Gly Thr Ala Glu Ser Arg Ala Tyr Lys	
15 20 25	
ATG TGG CTG CCG CCG TTG ACC AAT CCG GTC CCG CTC AAC GAG CTC ATC	207
Met Trp Leu Pro Pro Leu Thr Asn Pro Val Pro Leu Asn Glu Leu Ile	
30 35 40	
GCC CGT GAT CGG CGA CAA CCC CTG CGA TTT GCC CTG GGG ATC ATG GAT	255
Ala Arg Asp Arg Arg Gln Pro Leu Arg Phe Ala Leu Gly Ile Met Asp	
45 50 55	
GAA CCG CGC CGC CAT CTA CAG GAT GTG TGG GGC GTA GAC GTT TCC GGG	303
Glu Pro Arg Arg His Leu Gln Asp Val Trp Gly Val Asp Val Ser Gly	
60 65 70 75	
GCC GGC GGC AAC ATC GGT ATT GGG GGC GCA CCT CAA ACC GGG AAG TCG	351
Ala Gly Gly Asn Ile Gly Ile Gly Gly Ala Pro Gln Thr Gly Lys Ser	
80 85 90	
ACG CTA CTG CAG ACG ATG GTG ATG TCG GCC GCC GCC ACA CAC TCA CCG	399
Thr Leu Leu Gln Thr Met Val Met Ser Ala Ala Ala Thr His Ser Pro	
95 100 105	
CGC AAC GTT CAG TTC TAT TGC ATC GAC CTA GGT GGC GGC GGG CTG ATC	447
Arg Asn Val Gln Phe Tyr Cys Ile Asp Leu Gly Gly Gly Gly Leu Ile	
110 115 120	
TAT CTC GAA AAC CTT CCA CAC GTC GGT GGC GTA GCC AAT CGG TCC GAG	495
Tyr Leu Glu Asn Leu Pro His Val Gly Gly Val Ala Asn Arg Ser Glu	
125 130 135	
CCC GAC AAG GTC AAC CGG GTG GTC GCA GAG ATG CAA GCC GTC ATG CGG	543
Pro Asp Lys Val Asn Arg Val Val Ala Glu Met Gln Ala Val Met Arg	
140 145 150 155	
CAA CGG GAA ACC ACC TTC AAG GAA CAC CGA GTG GGC TCG ATC GGG ATG	591
Gln Arg Glu Thr Thr Phe Lys Glu His Arg Val Gly Ser Ile Gly Met	
160 165 170	
TAC CGG CAG CTG CGT GAC GAT CCA AGT CAA CCC GTT GCG TCC GAT CCA	639
Tyr Arg Gln Leu Arg Asp Asp Pro Ser Gln Pro Val Ala Ser Asp Pro	
175 180 185	
TAC GGC GAC GTC TTT CTG ATC ATC GAC GGA TGG CCC GGT TTT GTC GGC	687
Tyr Gly Asp Val Phe Leu Ile Ile Asp Gly Trp Pro Gly Phe Val Gly	
190 195 200	

GAG TTC CCC GAC CTT GAG GGG CAG GTT CAA GAT CTG GCC GCC CAG GGG Glu Phe Pro Asp Leu Glu Gly Gln Val Gln Asp Leu Ala Ala Gln Gly 205 210 215	735
CTG GGG TTC GGC GTC CAC GTC ATC ATC TCC ACC CCA CGC TGG ACA GAG Leu Gly Phe Gly Val His Val Ile Ile Ser Thr Pro Arg Trp Thr Glu 220 225 230 235	783
CTG AAG TCG CGT GTT CGC GAC TAC CTC GGC ACC AAG ATC GAG TTC CGG Leu Lys Ser Arg Val Arg Asp Tyr Leu Gly Thr Lys Ile Glu Phe Arg 240 245 250	831
CTT GGT GAC GTC AAT GAA ACC CAG ATC GAC CGG ATT ACC CGC GAG ATC Leu Gly Asp Val Asn Glu Thr Gln Ile Asp Arg Ile Thr Arg Glu Ile 255 260 265	879
CCG GCG AAT CGT CCG GGT CGG GCA GTG TCG ATG GAA AAG CAC CAT CTG Pro Ala Asn Arg Pro Gly Arg Ala Val Ser Met Glu Lys His His Leu 270 275 280	927
ATG ATC GGC GTG CCC AGG TTC GAC GGC GTG CAC AGC GCC GAT AAC CTG Met Ile Gly Val Pro Arg Phe Asp Gly Val His Ser Ala Asp Asn Leu 285 290 295	975
GTG GAG GCG ATC ACC GCG GGG GTG ACG CAG ATC GCT TCC CAG CAC ACC Val Glu Ala Ile Thr Ala Gly Val Thr Gln Ile Ala Ser Gln His Thr 300 305 310 315	1023
GAA CAG GCA CCT CCG GTG CGG GTC CTG CCG GAG CGT ATC CAC CTG CAC Glu Gln Ala Pro Pro Val Arg Val Leu Pro Glu Arg Ile His Leu His 320 325 330	1071
GAA CTC GAC CCG AAC CCG CCG GGA CCA GAG TCC GAC TAC CGC ACT CGC Glu Leu Asp Pro Asn Pro Pro Gly Pro Glu Ser Asp Tyr Arg Thr Arg 335 340 345	1119
TGG GAG ATT CCG ATC GGC TTG CGC GAG ACG GAC CTG ACG CCG GCT CAC Trp Glu Ile Pro Ile Gly Leu Arg Glu Thr Asp Leu Thr Pro Ala His 350 355 360	1167
TGC CAC ATG CAC ACG AAC CCG CAC CTA CTG ATC TTC GGT GCG GCC AAA Cys His Met His Thr Asn Pro His Leu Leu Ile Phe Gly Ala Ala Lys 365 370 375	1215
TCG GGC AAG ACG ACC ATT GCC CAC GCG ATC GCG CGC GCC ATT TGT GCC Ser Gly Lys Thr Thr Ile Ala His Ala Ile Ala Arg Ala Ile Cys Ala 380 385 390 395	1263
CGA AAC AGT CCC CAG CAG GTG CGG TTC ATG CTC GCG GAC TAC CGC TCG Arg Asn Ser Pro Gln Gln Val Arg Phe Met Leu Ala Asp Tyr Arg Ser 400 405 410	1311
GGC CTG CTG GAC GCG GTG CCG GAC ACC CAT CTG CTG GGC GCC GGC GCG Gly Leu Leu Asp Ala Val Pro Asp Thr His Leu Leu Gly Ala Gly Ala 415 420 425	1359
ATC AAC CGC AAC AGC GCG TCG CTA GAC GAG GCC GCT CAA GCA CTG GCG Ile Asn Arg Asn Ser Ala Ser Leu Asp Glu Ala Ala Gln Ala Leu Ala	1407

430	435	440	
GTC AAC CTG AAG AAG CGG TTG CCG CCG ACC GAC CTG ACG ACG GCG CAG Val Asn Leu Lys Lys Arg Leu Pro Pro Thr Asp Leu Thr Thr Ala Gln 445 450 455			1455
CTA CGC TCG CGT TCG TGG TGG AGC GGA TTT GAC GTC GTG CTT CTG GTC Leu Arg Ser Arg Ser Trp Trp Ser Gly Phe Asp Val Val Leu Leu Val 460 465 470 475			1503
GAC GAT TGG CAC ATG ATC GTG GGT GCC GCC GGG GGG ATG CCG CCG ATG Asp Asp Trp His Met Ile Val Gly Ala Ala Gly Gly Met Pro Pro Met 480 485 490			1551
GCA CCG CTG GCC CCG TTA TTG CCG GCG GCG GCA GAT ATC GGG TTG CAC Ala Pro Leu Ala Pro Leu Leu Pro Ala Ala Ala Asp Ile Gly Leu His 495 500 505			1599
ATC ATT GTC ACC TGT CAG ATG AGC CAG GCT TAC AAG GCA ACC ATG GAC Ile Ile Val Thr Cys Gln Met Ser Gln Ala Tyr Lys Ala Thr Met Asp 510 515 520			1647
AAG TTC GTC GGC GCC GCA TTC GGG TCG GGC GCT CCG ACA ATG TTC CTT Lys Phe Val Gly Ala Ala Phe Gly Ser Gly Ala Pro Thr Met Phe Leu 525 530 535			1695
TCG GGC GAG AAG CAG GAA TTC CCA TCC AGT GAG TTC AAG GTC AAG CGG Ser Gly Glu Lys Gln Glu Phe Pro Ser Ser Glu Phe Lys Val Lys Arg 540 545 550 555			1743
CGC CCC CCT GGC CAG GCA TTT CTC GTC TCG CCA GAC GGC AAA GAG GTC Arg Pro Pro Gly Gln Ala Phe Leu Val Ser Pro Asp Gly Lys Glu Val 560 565 570			1791
ATC CAG GCC CCC TAC ATC GAG CCT CCA GAA GAA GTG TTC GCA GCA CCC Ile Gln Ala Pro Tyr Ile Glu Pro Pro Glu Glu Val Phe Ala Ala Pro 575 580 585			1839
CCA AGC GCC GGT TAAGATTATT TCATTGCCGG TGTAGCAGGA CCCGAGCTC Pro Ser Ala Gly 590			1890

## (2) INFORMATION FOR SEQ ID NO: 72:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 591 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 72:

Met Thr Ala Glu Pro Glu Val Arg Thr Leu Arg Glu Val Val Leu Asp  
1 5 10 15



Gln Leu Gly Thr Ala Glu Ser Arg Ala Tyr Lys Met Trp Leu Pro Pro  
 20 25 30  
 Leu Thr Asn Pro Val Pro Leu Asn Glu Leu Ile Ala Arg Asp Arg Arg  
 35 40 45  
 Gln Pro Leu Arg Phe Ala Leu Gly Ile Met Asp Glu Pro Arg Arg His  
 50 55 60  
 Leu Gln Asp Val Trp Gly Val Asp Val Ser Gly Ala Gly Gly Asn Ile  
 65 70 75 80  
 Gly Ile Gly Gly Ala Pro Gln Thr Gly Lys Ser Thr Leu Leu Gln Thr  
 85 90 95  
 Met Val Met Ser Ala Ala Ala Thr His Ser Pro Arg Asn Val Gln Phe  
 100 105 110  
 Tyr Cys Ile Asp Leu Gly Gly Gly Gly Leu Ile Tyr Leu Glu Asn Leu  
 115 120 125  
 Pro His Val Gly Gly Val Ala Asn Arg Ser Glu Pro Asp Lys Val Asn  
 130 135 140  
 Arg Val Val Ala Glu Met Gln Ala Val Met Arg Gln Arg Glu Thr Thr  
 145 150 155 160  
 Phe Lys Glu His Arg Val Gly Ser Ile Gly Met Tyr Arg Gln Leu Arg  
 165 170 175  
 Asp Asp Pro Ser Gln Pro Val Ala Ser Asp Pro Tyr Gly Asp Val Phe  
 180 185 190  
 Leu Ile Ile Asp Gly Trp Pro Gly Phe Val Gly Glu Phe Pro Asp Leu  
 195 200 205  
 Glu Gly Gln Val Gln Asp Leu Ala Ala Gln Gly Leu Gly Phe Gly Val  
 210 215 220  
 His Val Ile Ile Ser Thr Pro Arg Trp Thr Glu Leu Lys Ser Arg Val  
 225 230 235 240  
 Arg Asp Tyr Leu Gly Thr Lys Ile Glu Phe Arg Leu Gly Asp Val Asn  
 245 250 255  
 Glu Thr Gln Ile Asp Arg Ile Thr Arg Glu Ile Pro Ala Asn Arg Pro  
 260 265 270  
 Gly Arg Ala Val Ser Met Glu Lys His His Leu Met Ile Gly Val Pro  
 275 280 285  
 Arg Phe Asp Gly Val His Ser Ala Asp Asn Leu Val Glu Ala Ile Thr  
 290 295 300  
 Ala Gly Val Thr Gln Ile Ala Ser Gln His Thr Glu Gln Ala Pro Pro  
 305 310 315 320  
 Val Arg Val Leu Pro Glu Arg Ile His Leu His Glu Leu Asp Pro Asn

189

325	330	335
Pro Pro Gly Pro Glu Ser Asp Tyr Arg Thr Arg Trp Glu Ile Pro Ile		
340	345	350
Gly Leu Arg Glu Thr Asp Leu Thr Pro Ala His Cys His Met His Thr		
355	360	365
Asn Pro His Leu Leu Ile Phe Gly Ala Ala Lys Ser Gly Lys Thr Thr		
370	375	380
Ile Ala His Ala Ile Ala Arg Ala Ile Cys Ala Arg Asn Ser Pro Gln		
385	390	395
Gln Val Arg Phe Met Leu Ala Asp Tyr Arg Ser Gly Leu Leu Asp Ala		
405	410	415
Val Pro Asp Thr His Leu Leu Gly Ala Gly Ala Ile Asn Arg Asn Ser		
420	425	430
Ala Ser Leu Asp Glu Ala Ala Gln Ala Leu Ala Val Asn Leu Lys Lys		
435	440	445
Arg Leu Pro Pro Thr Asp Leu Thr Thr Ala Gln Leu Arg Ser Arg Ser		
450	455	460
Trp Trp Ser Gly Phe Asp Val Val Leu Leu Val Asp Asp Trp His Met		
465	470	475
Ile Val Gly Ala Ala Gly Gly Met Pro Pro Met Ala Pro Leu Ala Pro		
485	490	495
Leu Leu Pro Ala Ala Ala Asp Ile Gly Leu His Ile Ile Val Thr Cys		
500	505	510
Gln Met Ser Gln Ala Tyr Lys Ala Thr Met Asp Lys Phe Val Gly Ala		
515	520	525
Ala Phe Gly Ser Gly Ala Pro Thr Met Phe Leu Ser Gly Glu Lys Gln		
530	535	540
Glu Phe Pro Ser Ser Glu Phe Lys Val Lys Arg Arg Pro Pro Gly Gln		
545	550	555
Ala Phe Leu Val Ser Pro Asp Gly Lys Glu Val Ile Gln Ala Pro Tyr		
565	570	575
Ile Glu Pro Pro Glu Glu Val Phe Ala Ala Pro Pro Ser Ala Gly		
580	585	590

(2) INFORMATION FOR SEQ ID NO: 73:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 73:

Asp Pro Val Asp Asp Ala Phe Ile Ala Lys Leu Asn Thr Ala Gly  
1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 74:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 14 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(ix) Feature:

(A) NAME/KEY: Other

(B) LOCATION: 14

(C) OTHER INFORMATION: Xaa is unknown

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 74:

Asp Pro Val Asp Ala Ile Ile Asn Leu Asp Asn Tyr Gly Xaa  
1 5 10

(2) INFORMATION FOR SEQ ID NO: 75:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(ix) Feature:

(A) NAME/KEY: Other

(B) LOCATION: 5

(C) OTHER INFORMATION: Xaa is unknown

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 75:

Ala Glu Met Lys Xaa Phe Lys Asn Ala Ile Val Gln Glu Ile Asp  
1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 76:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 14 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(ix) FEATURE:

(A) NAME/KEY: Other

(B) LOCATION: 3...3

(D) OTHER INFORMATION: Ala is Ala or Gln

(A) NAME/KEY: Other

(B) LOCATION: 7...7

(D) OTHER INFORMATION: Thr is Gly or Thr

(ix) Feature:

(A) NAME/KEY: Other

(B) LOCATION: 11

(C) OTHER INFORMATION: Xaa is unknown

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 76:

Val Ile Ala Gly Met Val Thr His Ile His Xaa Val Ala Gly  
1 5 10

(2) INFORMATION FOR SEQ ID NO: 77:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 77:

Thr Asn Ile Val Val Leu Ile Lys Gln Val Pro Asp Thr Trp Ser  
1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 78:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 78:

Ala Ile Glu Val Ser Val Leu Arg Val Phe Thr Asp Ser Asp Gly  
1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 79:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 15 amino acids  
 (B) TYPE: amino acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide  
 (v) FRAGMENT TYPE: N-terminal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 79:

Ala Lys Leu Ser Thr Asp Glu Leu Leu Asp Ala Phe Lys Glu Met  
 1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 80:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 15 amino acids  
 (B) TYPE: amino acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide  
 (v) FRAGMENT TYPE: N-terminal

(ix) FEATURE:

- (A) NAME/KEY: Other  
 (B) LOCATION: 4...4  
 (D) OTHER INFORMATION: Asp is Asp or Glu

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 80:

Asp Pro Ala Asp Ala Pro Asp Val Pro Thr Ala Ala Gln Leu Thr  
 1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 81:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 50 amino acids  
 (B) TYPE: amino acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide  
 (v) FRAGMENT TYPE: N-terminal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 81:

Ala Glu Asp Val Arg Ala Glu Ile Val Ala Ser Val Leu Glu Val Val  
 1 5 10 15

Val Asn Glu Gly Asp Gln Ile Asp Lys Gly Asp Val Val Val Leu Leu  
 20 25 30

Glu Ser Met Tyr Met Glu Ile Pro Val Leu Ala Glu Ala Ala Gly Thr  
 35 40 45  
 Val Ser

50

## (2) INFORMATION FOR SEQ ID NO: 82:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 15 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide
- (v) FRAGMENT TYPE: N-terminal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 82:

Thr	Thr	Ser	Pro	Asp	Pro	Tyr	Ala	Ala	Leu	Pro	Lys	Leu	Pro	Ser
1				5					10					15

## (2) INFORMATION FOR SEQ ID NO: 83:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 15 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide
- (v) FRAGMENT TYPE: N-terminal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 83:

Thr	Glu	Tyr	Glu	Gly	Pro	Lys	Thr	Lys	Phe	His	Ala	Leu	Met	Gln
1				5					10					15

## (2) INFORMATION FOR SEQ ID NO: 84:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 15 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide
- (v) FRAGMENT TYPE: N-terminal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 84:

Thr	Thr	Ile	Val	Ala	Leu	Lys	Tyr	Pro	Gly	Gly	Val	Val	Met	Ala
1				5					10					15

## (2) INFORMATION FOR SEQ ID NO: 85:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 15 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide
- (v) FRAGMENT TYPE: N-terminal

- (ix) FEATURE:
  - (A) NAME/KEY: Other
  - (B) LOCATION: 10
  - (D) OTHER INFORMATION: Xaa is unknown

- (ix) FEATURE:
  - (A) NAME/KEY: Other
  - (B) LOCATION: 15
  - (D) OTHER INFORMATION: Xaa is unknown

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 85:

Ser Phe Pro Tyr Phe Ile Ser Pro Glu Xaa Ala Met Arg Glu Xaa  
 1                      5                      10                      15

- (2) INFORMATION FOR SEQ ID NO: 86:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 15 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide
- (v) FRAGMENT TYPE: N-terminal

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 86:

Thr His Tyr Asp Val Val Val Leu Gly Ala Gly Pro Gly Gly Tyr  
 1                      5                      10                      15

- (2) INFORMATION FOR SEQ ID NO: 87:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 450 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: Other

- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 107...400
  - (D) OTHER INFORMATION:

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 87:

AGCCCGGTAA TCGAGTTCGG GCAATGCTGA CCATCGGGTT TGTTCCTCGGC TATAACCGAA 60

CGGTTTGTGT ACGGGATACA AATACAGGGA GGGAAGAAGT AGGCAA ATG GAA AAA 115  
 Met Glu Lys

1

ATG TCA CAT GAT CCG ATC GCT GCC GAC ATT GGC ACG CAA GTG AGC GAC 163  
 Met Ser His Asp Pro Ile Ala Ala Asp Ile Gly Thr Gln Val Ser Asp  
     5                    10                    15  
  
 AAC GCT CTG CAC GGC GTG ACG GCC GGC TCG ACG GCG CTG ACG TCG GTG 211  
 Asn Ala Leu His Gly Val Thr Ala Gly Ser Thr Ala Leu Thr Ser Val  
     20                    25                    30                    35  
  
 ACC GGG CTG GTT CCC GCG GGG GCC GAT GAG GTC TCC GCC CAA GCG GCG 259  
 Thr Gly Leu Val Pro Ala Gly Ala Asp Glu Val Ser Ala Gln Ala Ala  
                     40                    45                    50  
  
 ACG GCG TTC ACA TCG GAG GGC ATC CAA TTG CTG GCT TCC AAT GCA TCG 307  
 Thr Ala Phe Thr Ser Glu Gly Ile Gln Leu Leu Ala Ser Asn Ala Ser  
                     55                    60                    65  
  
 GCC CAA GAC CAG CTC CAC CGT GCG GGC GAA GCG GTC CAG GAC GTC GCC 355  
 Ala Gln Asp Gln Leu His Arg Ala Gly Glu Ala Val Gln Asp Val Ala  
                     70                    75                    80  
  
 CGC ACC TAT TCG CAA ATC GAC GAC GGC GCC GCC GGC GTC TTC GCC TAATA 405  
 Arg Thr Tyr Ser Gln Ile Asp Asp Gly Ala Ala Gly Val Phe Ala  
     85                    90                    95  
  
 GGCCCCAAC ACATCGGAGG GAGTGATCAC CATGCTGTGG CACGC 450

## (2) INFORMATION FOR SEQ ID NO: 88:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 98 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 88:

Met Glu Lys Met Ser His Asp Pro Ile Ala Ala Asp Ile Gly Thr Gln  
   1                    5                    10                    15  
  
 Val Ser Asp Asn Ala Leu His Gly Val Thr Ala Gly Ser Thr Ala Leu  
                     20                    25                    30  
  
 Thr Ser Val Thr Gly Leu Val Pro Ala Gly Ala Asp Glu Val Ser Ala  
                     35                    40                    45  
  
 Gln Ala Ala Thr Ala Phe Thr Ser Glu Gly Ile Gln Leu Leu Ala Ser  
                     50                    55                    60  
  
 Asn Ala Ser Ala Gln Asp Gln Leu His Arg Ala Gly Glu Ala Val Gln  
   65                    70                    75                    80  
  
 Asp Val Ala Arg Thr Tyr Ser Gln Ile Asp Asp Gly Ala Ala Gly Val  
                     85                    90                    95  
  
 Phe Ala



## (2) INFORMATION FOR SEQ ID NO: 89:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 460 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 37...453
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 89:

GCAACCGGCT TTTGATCAG CTGAGACATC AGCGGC GTG CGG GTC AAC GAC CCA	54
Met Arg Val Asn Asp Pro	
1 5	
CCT GCG CCA GGT AGC GAC TCC GCG CGC AGC AGG CCC GCG CCC GCG CTG	102
Pro Ala Pro Gly Ser Asp Ser Ala Arg Ser Arg Pro Ala Pro Ala Leu	
10 15 20	
GGG CCT GAT CCA CCA GCC AGC GGA TGG TTC GAC AGC GGA CTG GTG CCG	150
Gly Pro Asp Pro Pro Ala Ser Gly Trp Phe Asp Ser Gly Leu Val Pro	
25 30 35	
AGC AGG CCC ATC TGC GCG GCT TCC TCG TCG GCT GGG TTG CCG CCG CCG	198
Ser Arg Pro Ile Cys Ala Ala Ser Ser Ser Ala Gly Leu Pro Pro Pro	
40 45 50	
GTG CCG CCC ACC TGG CTG AAC AAC GAC GTC ACC TGC TGC AGC GGC TGG	246
Val Pro Pro Thr Trp Leu Asn Asn Asp Val Thr Cys Cys Ser Gly Trp	
55 60 65 70	
GTC AGC TGC TGC ATC GGG CCG CTC ATC TCA CCC AGT TGG CCG AGG GTC	294
Val Ser Cys Cys Ile Gly Pro Leu Ile Ser Pro Ser Trp Pro Arg Val	
75 80 85	
TGG GTA GCC GCC GGC GGC AAC TGG CCA ACC GGT GTT GAG CTG CCA GGG	342
Trp Val Ala Ala Gly Gly Asn Trp Pro Thr Gly Val Glu Leu Pro Gly	
90 95 100	
GAG GGC ATT CCG AAG ATC GGG TTC GTC GTG CTC TGG CTC GCG CCG GGA	390
Glu Gly Ile Pro Lys Ile Gly Phe Val Val Leu Trp Leu Ala Pro Gly	
105 110 115	
TCA AGG ATC GAC GCC ATC GGC TCG AGC TTC TCG AAA AGC GTG TTA ACC	438
Ser Arg Ile Asp Ala Ile Gly Ser Ser Phe Ser Lys Ser Val Leu Thr	
120 125 130	
GCG GTC TCG GCC TGG TAGACCT	460
Ala Val Ser Ala Trp	
135	

## (2) INFORMATION FOR SEQ ID NO: 90:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 139 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 90:

```

Met Arg Val Asn Asp Pro Pro Ala Pro Gly Ser Asp Ser Ala Arg Ser
 1             5             10             15

Arg Pro Ala Pro Ala Leu Gly Pro Asp Pro Pro Ala Ser Gly Trp Phe
          20             25             30

Asp Ser Gly Leu Val Pro Ser Arg Pro Ile Cys Ala Ala Ser Ser Ser
          35             40             45

Ala Gly Leu Pro Pro Pro Val Pro Pro Thr Trp Leu Asn Asn Asp Val
 50             55             60

Thr Cys Cys Ser Gly Trp Val Ser Cys Cys Ile Gly Pro Leu Ile Ser
 65             70             75             80

Pro Ser Trp Pro Arg Val Trp Val Ala Ala Gly Gly Asn Trp Pro Thr
          85             90             95

Gly Val Glu Leu Pro Gly Glu Gly Ile Pro Lys Ile Gly Phe Val Val
          100            105            110

Leu Trp Leu Ala Pro Gly Ser Arg Ile Asp Ala Ile Gly Ser Ser Phe
          115            120            125

Ser Lys Ser Val Leu Thr Ala Val Ser Ala Trp
          130            135

```

## (2) INFORMATION FOR SEQ ID NO: 91:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1200 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 28...1140
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 91:

TAATAGGCCC CCAACACATC GGAGGGA GTG ATC ACC ATG CTG TGG CAC GCA ATG	54
Met Ile Thr Met Leu Trp His Ala Met	
1 5	
CCA CCG GAG CTA AAT ACC GCA CGG CTG ATG GCC GGC GCG GGT CCG GCT	102
Pro Pro Glu Leu Asn Thr Ala Arg Leu Met Ala Gly Ala Gly Pro Ala	
10 15 20 25	
CCA ATG CTT GCG GCG GCC GCG GGA TGG CAG ACG CTT TCG GCG GCT CTG	150
Pro Met Leu Ala Ala Ala Gly Trp Gln Thr Leu Ser Ala Ala Leu	
30 35 40	
GAC GCT CAG GCC GTC GAG TTG ACC GCG CGC CTG AAC TCT CTG GGA GAA	198
Asp Ala Gln Ala Val Glu Leu Thr Ala Arg Leu Asn Ser Leu Gly Glu	
45 50 55	
GCC TGG ACT GGA GGT GGC AGC GAC AAG GCG CTT GCG GCT GCA ACG CCG	246
Ala Trp Thr Gly Gly Gly Ser Asp Lys Ala Leu Ala Ala Thr Pro	
60 65 70	
ATG GTG GTC TGG CTA CAA ACC GCG TCA ACA CAG GCC AAG ACC CGT GCG	294
Met Val Val Trp Leu Gln Thr Ala Ser Thr Gln Ala Lys Thr Arg Ala	
75 80 85	
ATG CAG GCG ACG GCG CAA GCC GCG GCA TAC ACC CAG GCC ATG GCC ACG	342
Met Gln Ala Thr Ala Gln Ala Ala Ala Tyr Thr Gln Ala Met Ala Thr	
90 95 100 105	
ACG CCG TCG CTG CCG GAG ATC GCC GCC AAC CAC ATC ACC CAG GCC GTC	390
Thr Pro Ser Leu Pro Glu Ile Ala Ala Asn His Ile Thr Gln Ala Val	
110 115 120	
CTT ACG GCC ACC AAC TTC TTC GGT ATC AAC ACG ATC CCG ATC GCG TTG	438
Leu Thr Ala Thr Asn Phe Phe Gly Ile Asn Thr Ile Pro Ile Ala Leu	
125 130 135	
ACC GAG ATG GAT TAT TTC ATC CGT ATG TGG AAC CAG GCA GCC CTG GCA	486
Thr Glu Met Asp Tyr Phe Ile Arg Met Trp Asn Gln Ala Ala Leu Ala	
140 145 150	
ATG GAG GTC TAC CAG GCC GAG ACC GCG GTT AAC ACG CTT TTC GAG AAG	534
Met Glu Val Tyr Gln Ala Glu Thr Ala Val Asn Thr Leu Phe Glu Lys	
155 160 165	
CTC GAG CCG ATG GCG TCG ATC CTT GAT CCC GGC GCG AGC CAG AGC ACG	582
Leu Glu Pro Met Ala Ser Ile Leu Asp Pro Gly Ala Ser Gln Ser Thr	
170 175 180 185	
ACG AAC CCG ATC TTC GGA ATG CCC TCC CCT GGC AGC TCA ACA CCG GTT	630
Thr Asn Pro Ile Phe Gly Met Pro Ser Pro Gly Ser Ser Thr Pro Val	
190 195 200	
GGC CAG TTG CCG CCG GCG GCT ACC CAG ACC CTC GGC CAA CTG GGT GAG	678
Gly Gln Leu Pro Pro Ala Ala Thr Gln Thr Leu Gly Gln Leu Gly Glu	
205 210 215	
ATG AGC GGC CCG ATG CAG CAG CTG ACC CAG CCG CTG CAG CAG GTG ACG	726
Met Ser Gly Pro Met Gln Gln Leu Thr Gln Pro Leu Gln Gln Val Thr	

220	225	230	
TCG TTG TTC AGC CAG GTG GGC GGC ACC GGC GGC GGC AAC CCA GCC GAC			774
Ser Leu Phe Ser Gln Val Gly Gly Thr Gly Gly Gly Asn Pro Ala Asp			
235	240	245	
GAG GAA GCC GCG CAG ATG GGC CTG CTC GGC ACC AGT CCG CTG TCG AAC			822
Glu Glu Ala Ala Gln Met Gly Leu Leu Gly Thr Ser Pro Leu Ser Asn			
250	255	260	265
CAT CCG CTG GCT GGT GGA TCA GGC CCC AGC GCG GGC GCG GGC CTG CTG			870
His Pro Leu Ala Gly Gly Ser Gly Pro Ser Ala Gly Ala Gly Leu Leu			
270	275	280	
CGC GCG GAG TCG CTA CCT GGC GCA GGT GGG TCG TTG ACC CGC ACG CCG			918
Arg Ala Glu Ser Leu Pro Gly Ala Gly Gly Ser Leu Thr Arg Thr Pro			
285	290	295	
CTG ATG TCT CAG CTG ATC GAA AAG CCG GTT GCC CCC TCG GTG ATG CCG			966
Leu Met Ser Gln Leu Ile Glu Lys Pro Val Ala Pro Ser Val Met Pro			
300	305	310	
GCG GCT GCT GCC GGA TCG TCG GCG ACG GGT GGC GCC GCT CCG GTG GGT			1014
Ala Ala Ala Ala Gly Ser Ser Ala Thr Gly Gly Ala Ala Pro Val Gly			
315	320	325	
GCG GGA GCG ATG GGC CAG GGT GCG CAA TCC GGC GGC TCC ACC AGG CCG			1062
Ala Gly Ala Met Gly Gln Gly Ala Gln Ser Gly Gly Ser Thr Arg Pro			
330	335	340	345
GGT CTG GTC GCG CCG GCA CCG CTC GCG CAG GAG CGT GAA GAA GAC GAC			1110
Gly Leu Val Ala Pro Ala Pro Leu Ala Gln Glu Arg Glu Glu Asp Asp			
350	355	360	
GAG GAC GAC TGG GAC GAA GAG GAC GAC TGG TGAGCTCCCG TAATGACAAC AGA			1163
Glu Asp Asp Trp Asp Glu Glu Asp Asp Trp			
365	370		
CTTCCCGGCC ACCCGGGCCG GAAGACTTGC CAACATT			1200

## (2) INFORMATION FOR SEQ ID NO: 92:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 371 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 92:

Met	Ile	Thr	Met	Leu	Trp	His	Ala	Met	Pro	Pro	Glu	Leu	Asn	Thr	Ala
1				5					10					15	
Arg	Leu	Met	Ala	Gly	Ala	Gly	Pro	Ala	Pro	Met	Leu	Ala	Ala	Ala	Ala

200

20					25					30						
Gly	Trp	Gln	Thr	Leu	Ser	Ala	Ala	Leu	Asp	Ala	Gln	Ala	Val	Glu	Leu	
35					40					45						
Thr	Ala	Arg	Leu	Asn	Ser	Leu	Gly	Glu	Ala	Trp	Thr	Gly	Gly	Gly	Ser	
50					55					60						
Asp	Lys	Ala	Leu	Ala	Ala	Ala	Thr	Pro	Met	Val	Val	Trp	Leu	Gln	Thr	
65					70					75					80	
Ala	Ser	Thr	Gln	Ala	Lys	Thr	Arg	Ala	Met	Gln	Ala	Thr	Ala	Gln	Ala	
85					90					95						
Ala	Ala	Tyr	Thr	Gln	Ala	Met	Ala	Thr	Thr	Pro	Ser	Leu	Pro	Glu	Ile	
100					105					110						
Ala	Ala	Asn	His	Ile	Thr	Gln	Ala	Val	Leu	Thr	Ala	Thr	Asn	Phe	Phe	
115					120					125						
Gly	Ile	Asn	Thr	Ile	Pro	Ile	Ala	Leu	Thr	Glu	Met	Asp	Tyr	Phe	Ile	
130					135					140						
Arg	Met	Trp	Asn	Gln	Ala	Ala	Leu	Ala	Met	Glu	Val	Tyr	Gln	Ala	Glu	
145					150					155					160	
Thr	Ala	Val	Asn	Thr	Leu	Phe	Glu	Lys	Leu	Glu	Pro	Met	Ala	Ser	Ile	
165					170					175						
Leu	Asp	Pro	Gly	Ala	Ser	Gln	Ser	Thr	Thr	Asn	Pro	Ile	Phe	Gly	Met	
180					185					190						
Pro	Ser	Pro	Gly	Ser	Ser	Thr	Pro	Val	Gly	Gln	Leu	Pro	Pro	Ala	Ala	
195					200					205						
Thr	Gln	Thr	Leu	Gly	Gln	Leu	Gly	Glu	Met	Ser	Gly	Pro	Met	Gln	Gln	
210					215					220						
Leu	Thr	Gln	Pro	Leu	Gln	Gln	Val	Thr	Ser	Leu	Phe	Ser	Gln	Val	Gly	
225					230					235					240	
Gly	Thr	Gly	Gly	Gly	Asn	Pro	Ala	Asp	Glu	Glu	Ala	Ala	Gln	Met	Gly	
245					250					255						
Leu	Leu	Gly	Thr	Ser	Pro	Leu	Ser	Asn	His	Pro	Leu	Ala	Gly	Gly	Ser	
260					265					270						
Gly	Pro	Ser	Ala	Gly	Ala	Gly	Leu	Leu	Arg	Ala	Glu	Ser	Leu	Pro	Gly	
275					280					285						
Ala	Gly	Gly	Ser	Leu	Thr	Arg	Thr	Pro	Leu	Met	Ser	Gln	Leu	Ile	Glu	
290					295					300						
Lys	Pro	Val	Ala	Pro	Ser	Val	Met	Pro	Ala	Ala	Ala	Ala	Gly	Ser	Ser	
305					310					315					320	
Ala	Thr	Gly	Gly	Ala	Ala	Pro	Val	Gly	Ala	Gly	Ala	Met	Gly	Gln	Gly	
325					330					335						

Ala Gln Ser Gly Gly Ser Thr Arg Pro Gly Leu Val Ala Pro Ala Pro  
                   340                                  345                                  350

Leu Ala Gln Glu Arg Glu Glu Asp Asp Glu Asp Asp Trp Asp Glu Glu  
                   355                                  360                                  365

Asp Asp Trp  
                   370

(2) INFORMATION FOR SEQ ID NO: 93:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1000 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 46...969
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 93:

GACGCGACAC AGAAATCCTT AAGGCCGGCG GCCAAGGGGC CGAAG GTG AAG AAG GTG	57
Met Lys Lys Val	
1	
AAG CCC CAG AAA CCG AAG GCC ACG AAG CCG CCC AAA GTG GTG TCG CAG	105
Lys Pro Gln Lys Pro Lys Ala Thr Lys Pro Pro Lys Val Val Ser Gln	
5                                  10                                  15                                  20	
CGC GGC TGG CGA CAT TGG GTG CAT GCG TTG ACG CGA ATC AAC CTG GGC	153
Arg Gly Trp Arg His Trp Val His Ala Leu Thr Arg Ile Asn Leu Gly	
25                                  30                                  35	
CTG TCA CCC GAC GAG AAG TAC GAG CTG GAC CTG CAC GCT CGA GTC CGC	201
Leu Ser Pro Asp Glu Lys Tyr Glu Leu Asp Leu His Ala Arg Val Arg	
40                                  45                                  50	
CGC AAT CCC CGC GGG TCG TAT CAG ATC GCC GTC GTC GGT CTC AAA GGT	249
Arg Asn Pro Arg Gly Ser Tyr Gln Ile Ala Val Val Gly Leu Lys Gly	
55                                  60                                  65	
GGG GCT GGC AAA ACC ACG CTG ACA GCA GCG TTG GGG TCG ACG TTG GCT	297
Gly Ala Gly Lys Thr Thr Leu Thr Ala Ala Leu Gly Ser Thr Leu Ala	
70                                  75                                  80	
CAG GTG CGG GCC GAC CGG ATC CTG GCT CTA GAC GCG GAT CCA GGC GCC	345
Gln Val Arg Ala Asp Arg Ile Leu Ala Leu Asp Ala Asp Pro Gly Ala	
85                                  90                                  95                                  100	
GGA AAC CTC GCC GAT CGG GTA GGG CGA CAA TCG GGC GCG ACC ATC GCT	393
Gly Asn Leu Ala Asp Arg Val Gly Arg Gln Ser Gly Ala Thr Ile Ala	
105                                  110                                  115	

GAT GTG CTT GCA GAA AAA GAG CTG TCG CAC TAC AAC GAC ATC CGC GCA Asp Val Leu Ala Glu Lys Glu Leu Ser His Tyr Asn Asp Ile Arg Ala 120 125 130	441
CAC ACT AGC GTC AAT GCG GTC AAT CTG GAA GTG CTG CCG GCA CCG GAA His Thr Ser Val Asn Ala Val Asn Leu Glu Val Leu Pro Ala Pro Glu 135 140 145	489
TAC AGC TCG GCG CAG CGC GCG CTC AGC GAC GCC GAC TGG CAT TTC ATC Tyr Ser Ser Ala Gln Arg Ala Leu Ser Asp Ala Asp Trp His Phe Ile 150 155 160	537
GCC GAT CCT GCG TCG AGG TTT TAC AAC CTC GTC TTG GCT GAT TGT GGG Ala Asp Pro Ala Ser Arg Phe Tyr Asn Leu Val Leu Ala Asp Cys Gly 165 170 175 180	585
GCC GGC TTC TTC GAC CCG CTG ACC CGC GGC GTG CTG TCC ACG GTG TCC Ala Gly Phe Phe Asp Pro Leu Thr Arg Gly Val Leu Ser Thr Val Ser 185 190 195	633
GGT GTC GTG GTC GTG GCA AGT GTC TCA ATC GAC GGC GCA CAA CAG GCG Gly Val Val Val Val Ala Ser Val Ser Ile Asp Gly Ala Gln Gln Ala 200 205 210	681
TCG GTC GCG TTG GAC TGG TTG CGC AAC AAC GGT TAC CAA GAT TTG GCG Ser Val Ala Leu Asp Trp Leu Arg Asn Asn Gly Tyr Gln Asp Leu Ala 215 220 225	729
AGC CGC GCA TGC GTG GTC ATC AAT CAC ATC ATG CCG GGA GAA CCC AAT Ser Arg Ala Cys Val Val Ile Asn His Ile Met Pro Gly Glu Pro Asn 230 235 240	777
GTC GCA GTT AAA GAC CTG GTG CGG CAT TTC GAA CAG CAA GTT CAA CCC Val Ala Val Lys Asp Leu Val Arg His Phe Glu Gln Gln Val Gln Pro 245 250 255 260	825
GGC CGG GTC GTG GTC ATG CCG TGG GAC AGG CAC ATT GCG GCC GGA ACC Gly Arg Val Val Val Met Pro Trp Asp Arg His Ile Ala Ala Gly Thr 265 270 275	873
GAG ATT TCA CTC GAC TTG CTC GAC CCT ATC TAC AAG CGC AAG GTC CTC Glu Ile Ser Leu Asp Leu Leu Asp Pro Ile Tyr Lys Arg Lys Val Leu 280 285 290	921
GAA TTG GCC GCA GCG CTA TCC GAC GAT TTC GAG AGG GCT GGA CGT CGT T Glu Leu Ala Ala Ala Leu Ser Asp Asp Phe Glu Arg Ala Gly Arg Arg 295 300 305	970
GAGCGCACCT GCTGTTGCTG CTGGTCCTAC	1000

## (2) INFORMATION FOR SEQ ID NO: 94:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 308 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein  
(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 94:

```

Met Lys Lys Val Lys Pro Gln Lys Pro Lys Ala Thr Lys Pro Pro Lys
 1             5             10             15

Val Val Ser Gln Arg Gly Trp Arg His Trp Val His Ala Leu Thr Arg
      20             25             30

Ile Asn Leu Gly Leu Ser Pro Asp Glu Lys Tyr Glu Leu Asp Leu His
      35             40             45

Ala Arg Val Arg Arg Asn Pro Arg Gly Ser Tyr Gln Ile Ala Val Val
      50             55             60

Gly Leu Lys Gly Gly Ala Gly Lys Thr Thr Leu Thr Ala Ala Leu Gly
 65             70             75             80

Ser Thr Leu Ala Gln Val Arg Ala Asp Arg Ile Leu Ala Leu Asp Ala
      85             90             95

Asp Pro Gly Ala Gly Asn Leu Ala Asp Arg Val Gly Arg Gln Ser Gly
      100            105            110

Ala Thr Ile Ala Asp Val Leu Ala Glu Lys Glu Leu Ser His Tyr Asn
      115            120            125

Asp Ile Arg Ala His Thr Ser Val Asn Ala Val Asn Leu Glu Val Leu
      130            135            140

Pro Ala Pro Glu Tyr Ser Ser Ala Gln Arg Ala Leu Ser Asp Ala Asp
      145            150            155            160

Trp His Phe Ile Ala Asp Pro Ala Ser Arg Phe Tyr Asn Leu Val Leu
      165            170            175

Ala Asp Cys Gly Ala Gly Phe Phe Asp Pro Leu Thr Arg Gly Val Leu
      180            185            190

Ser Thr Val Ser Gly Val Val Val Val Ala Ser Val Ser Ile Asp Gly
      195            200            205

Ala Gln Gln Ala Ser Val Ala Leu Asp Trp Leu Arg Asn Asn Gly Tyr
      210            215            220

Gln Asp Leu Ala Ser Arg Ala Cys Val Val Ile Asn His Ile Met Pro
      225            230            235            240

Gly Glu Pro Asn Val Ala Val Lys Asp Leu Val Arg His Phe Glu Gln
      245            250            255

Gln Val Gln Pro Gly Arg Val Val Val Met Pro Trp Asp Arg His Ile
      260            265            270

Ala Ala Gly Thr Glu Ile Ser Leu Asp Leu Leu Asp Pro Ile Tyr Lys
      275            280            285

```



Arg Lys Val Leu Glu Leu Ala Ala Ala Leu Ser Asp Asp Phe Glu Arg  
 290 295 300

Ala Gly Arg Arg  
 305

(2) INFORMATION FOR SEQ ID NO: 95:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 34 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 95:

AAGAGTAGAT CTATGATGGC CGAGGATGTT CGCG

34

(2) INFORMATION FOR SEQ ID NO: 96:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 27 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 96:

CGGCGACGAC GGATCCTACC GCGTCGG

27

(2) INFORMATION FOR SEQ ID NO: 97:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 28 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 97:

CCTTGGGAGA TCTTTGGACC CCGGTTGC

28

(2) INFORMATION FOR SEQ ID NO: 98:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 25 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 98:

GACGAGATCT TATGGGCTTA CTGAC

25

(2) INFORMATION FOR SEQ ID NO: 99:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 99:

CCCCCAGAT CTGCACCACC GGCATCGGCG GGC

33

(2) INFORMATION FOR SEQ ID NO: 100

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 100:

GCGGCGGATC CGTTGCTTAG CCGG

24

(2) INFORMATION FOR SEQ ID NO: 101:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 32 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 101:

CCGGCTGAGA TCTATGACAG AATACGAAGG GC

32

(2) INFORMATION FOR SEQ ID NO: 102:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 102:

CCCCGCCAGG GAACTAGAGG CGGC

24

(2) INFORMATION FOR SEQ ID NO: 103:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 38 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 103:

CTGCCGAGAT CTACCACCAT TGTGCGCTG AAATACCC

38

(2) INFORMATION FOR SEQ ID NO: 104:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 25 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 104:

CGCCATGGCC TTACGCGCCA ACTCG

25

(2) INFORMATION FOR SEQ ID NO: 105:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 32 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 105:

GGCGGAGATC TGTGAGTTTT CCGTATTTC A TC

32

(2) INFORMATION FOR SEQ ID NO: 106:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 25 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 106:

CGCGTCGAGC CATGGTTAGG CGCAG

25

(2) INFORMATION FOR SEQ ID NO: 107:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 32 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 107:

GAGGAAGATC TATGACAACT TCACCCGACC CG

32

(2) INFORMATION FOR SEQ ID NO: 108:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 28 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 108:

CATGAAGCCA TGGCCCGCAG GCTGCATG

28

(2) INFORMATION FOR SEQ ID NO: 109:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 109:

GGCCGAGATC TGTGACCCAC TATGACGTCG TCG

33

(2) INFORMATION FOR SEQ ID NO: 110:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 110:

GGCGCCCATG GTCAGAAATT GATCATGTGG CCAACC

36

(2) INFORMATION FOR SEQ ID NO: 111:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 111:

CCGGGAGATC TATGGCAAAG CTCTCCACCG ACG

33

(2) INFORMATION FOR SEQ ID NO: 112:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 32 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 112:

CGCTGGGCAG AGCTACTTGA CGGTGACGGT GG

32

(2) INFORMATION FOR SEQ ID NO: 113:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 36 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 113:

GGCCCAGATC TATGGCCATT GAGGTTTCGG TGTTCG

36

(2) INFORMATION FOR SEQ ID NO: 114:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 26 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 114:

CGCCGTGTTG CATGGCAGCG CTGAGC

26

(2) INFORMATION FOR SEQ ID NO: 115:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 24 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 115:

GGACGTTCAA GCGACACATC GCCG

24

(2) INFORMATION FOR SEQ ID NO: 116:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 24 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 116:

CAGCACGAAC GCGCCGTCGA TGGC

24

(2) INFORMATION FOR SEQ ID NO: 117:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 26 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 117:

ACAGATCTGT GACGGACATG AACCCG

26

(2) INFORMATION FOR SEQ ID NO: 118:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 28 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 118:

TTTTCATGG TCACGGGCCC CCGGTACT

28

(2) INFORMATION FOR SEQ ID NO: 119:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 26 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 119:

ACAGATCTGT GCCCATGGCA CAGATA

26

(2) INFORMATION FOR SEQ ID NO: 120:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 120:

TTTAAGCTTC TAGGCGCCCA GCGCGGC

27

(2) INFORMATION FOR SEQ ID NO: 121:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 26 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 121:

ACAGATCTGC GCATGCGGAT CCGTGT

26

(2) INFORMATION FOR SEQ ID NO: 122:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 28 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 122:

TTTCCATGG TCATCCGGCG TGATCGAG

28

(2) INFORMATION FOR SEQ ID NO: 123:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 26 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 123:

ACAGATCTGT AATGGCAGAC TGTGAT

26

(2) INFORMATION FOR SEQ ID NO: 124:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 28 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 124:

TTTCCATGG TCAGGAGATG GTGATCGA

28

(2) INFORMATION FOR SEQ ID NO: 125:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 26 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 125:

ACAGATCTGC CGGCTACCCC GGTGCC

26

(2) INFORMATION FOR SEQ ID NO: 126:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 28 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 126:

TTTCCATGG CTATTGCAGC TTTCCGGC

28

(2) INFORMATION FOR SEQ ID NO: 127:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 50 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 127:

Ala	Glu	Asp	Val	Arg	Ala	Glu	Ile	Val	Ala	Ser	Val	Leu	Glu	Val	Val
1				5				10					15		
Val	Asn	Glu	Gly	Asp	Gln	Ile	Asp	Lys	Gly	Asp	Val	Val	Val	Leu	Leu
			20				25					30			
Glu	Ser	Met	Tyr	Met	Glu	Ile	Pro	Val	Leu	Ala	Glu	Ala	Ala	Gly	Thr
		35				40					45				
Val	Ser														
		50													

(2) INFORMATION FOR SEQ ID NO: 128:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 49 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 128:

Ala	Glu	Asp	Val	Arg	Ala	Glu	Ile	Val	Ala	Ser	Val	Leu	Glu	Val	Val
1				5				10					15		
Val	Asn	Glu	Gly	Asp	Gln	Ile	Asp	Lys	Gly	Asp	Val	Val	Val	Leu	Leu
			20				25					30			



Glu Ser Met Met Glu Ile Pro Val Leu Ala Glu Ala Ala Gly Thr Val  
                   35                  40                  45

Ser

(2) INFORMATION FOR SEQ ID NO: 129:

- (i) SEQUENCE CHARACTERISTICS:  
     (A) LENGTH: 50 amino acids  
     (B) TYPE: amino acid  
     (C) STRANDEDNESS: single  
     (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 129:

Ala Glu Asp Val Arg Ala Glu Ile Val Ala Ser Val Leu Glu Val Val  
   1                  5                  10                  15  
 Val Asn Glu Gly Asp Gln Ile Asp Lys Gly Asp Val Val Val Leu Leu  
                   20                  25                  30  
 Glu Ser Met Lys Met Glu Ile Pro Val Leu Ala Glu Ala Ala Gly Thr  
                   35                  40                  45  
 Val Ser  
   50

(2) INFORMATION FOR SEQ ID NO: 130:

- (i) SEQUENCE CHARACTERISTICS:  
     (A) LENGTH: 33 base pairs  
     (B) TYPE: nucleic acid  
     (C) STRANDEDNESS: single  
     (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 130:

CCGGGAGATC TATGGCAAAG CTCTCCACCG ACG

33

(2) INFORMATION FOR SEQ ID NO: 131:

- (i) SEQUENCE CHARACTERISTICS:  
     (A) LENGTH: 32 base pairs  
     (B) TYPE: nucleic acid  
     (C) STRANDEDNESS: single  
     (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 131:

CGCTGGGCAG AGCTACTTGA CGGTGACGGT GG

32

(2) INFORMATION FOR SEQ ID NO: 132:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 36 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 132:

GGCGCCGGCA AGCTTGCCAT GACAGAGCAG CAGTGG

36

(2) INFORMATION FOR SEQ ID NO: 133:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 26 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 133:

CGAACTCGCC GGATCCCGTG TTTCGC

26

(2) INFORMATION FOR SEQ ID NO: 134:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 32 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 134:

GGCAACCGCG AGATCTTTCT CCCGGCCGGG GC

32

(2) INFORMATION FOR SEQ ID NO: 135:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 27 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 135:

GGCAAGCTTG CCGGCGCCTA ACGAACT

27

(2) INFORMATION FOR SEQ ID NO: 136:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 30 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 136:

GGACCCAGAT CTATGACAGA GCAGCAGTGG

30

(2) INFORMATION FOR SEQ ID NO: 137:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 47 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 137:

CCGGCAGCCC CGGCCGGGAG AAAAGCTTTG CGAACATCCC AGTGACG

47

(2) INFORMATION FOR SEQ ID NO: 138:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 44 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 138:

GTTCGCAAAG CTTTCTCCC GGCCGGGGCT GCCGGTCGAG TACC

44

(2) INFORMATION FOR SEQ ID NO: 139:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 139:

CCTTCGGTGG ATCCCGTCAG

20

(2) INFORMATION FOR SEQ ID NO: 140:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 450 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 68...346
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 140:

```

TGGCGCTGTC ACCGAGGAAC CTGTCAATGT CGTCGAGCAG TACTGAACCG TTCCGAGAAA      60
GGCCAGC ATG AAC GTC ACC GTA TCC ATT CCG ACC ATC CTG CGG CCC CAC      109
    Met Asn Val Thr Val Ser Ile Pro Thr Ile Leu Arg Pro His
        1             5             10
ACC GGC GGC CAG AAG AGT GTC TCG GCC AGC GGC GAT ACC TTG GGT GCC      157
Thr Gly Gly Gln Lys Ser Val Ser Ala Ser Gly Asp Thr Leu Gly Ala
    15             20             25             30
GTC ATC AGC GAC CTG GAG GCC AAC TAT TCG GGC ATT TCC GAG CGC CTG      205
Val Ile Ser Asp Leu Glu Ala Asn Tyr Ser Gly Ile Ser Glu Arg Leu
        35             40             45
ATG GAC CCG TCT TCC CCA GGT AAG TTG CAC CGC TTC GTG AAC ATC TAC      253
Met Asp Pro Ser Ser Pro Gly Lys Leu His Arg Phe Val Asn Ile Tyr
        50             55             60
GTC AAC GAC GAG GAC GTG CGG TTC TCC GGC GGC TTG GCC ACC GCG ATC      301
Val Asn Asp Glu Asp Val Arg Phe Ser Gly Gly Leu Ala Thr Ala Ile
        65             70             75
GCT GAC GGT GAC TCG GTC ACC ATC CTC CCC GCC GTG GCC GGT GGG TGAGC      351
Ala Asp Gly Asp Ser Val Thr Ile Leu Pro Ala Val Ala Gly Gly
        80             85             90
GGAGCACATG ACACGATACG ACTCGCTGTT GCAGGCCTTG GGCAACACGC CGCTGGTTGG      411
CCTGCAGCGA TTGTCGCCAC GCTGGGATGA CGGGCGAGA      450

```

## (2) INFORMATION FOR SEQ ID NO: 141:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 93 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 141:

```

Met Asn Val Thr Val Ser Ile Pro Thr Ile Leu Arg Pro His Thr Gly
    1             5             10             15
Gly Gln Lys Ser Val Ser Ala Ser Gly Asp Thr Leu Gly Ala Val Ile
        20             25             30
Ser Asp Leu Glu Ala Asn Tyr Ser Gly Ile Ser Glu Arg Leu Met Asp
        35             40             45
Pro Ser Ser Pro Gly Lys Leu His Arg Phe Val Asn Ile Tyr Val Asn
        50             55             60

```

Asp Glu Asp Val Arg Phe Ser Gly Gly Leu Ala Thr Ala Ile Ala Asp  
 65 70 75 80

Gly Asp Ser Val Thr Ile Leu Pro Ala Val Ala Gly Gly  
 85 90

## (2) INFORMATION FOR SEQ ID NO: 142:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 480 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 88...381
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 142:

```

GGTGTTCCTCG CGGCCGGCTA TGACAACAGT CAATGTGCAT GACAAGTTAC AGGTATTAGG      60
TCCAGGTTCA ACAAGGAGAC AGGCAAC ATG GCA ACA CGT TTT ATG ACG GAT CCG      114
Met Ala Thr Arg Phe Met Thr Asp Pro
1 5
CAC GCG ATG CGG GAC ATG GCG GGC CGT TTT GAG GTG CAC GCC CAG ACG      162
His Ala Met Arg Asp Met Ala Gly Arg Phe Glu Val His Ala Gln Thr
10 15 20 25
GTG GAG GAC GAG GCT CGC CGG ATG TGG GCG TCC GCG CAA AAC ATC TCG      210
Val Glu Asp Glu Ala Arg Arg Met Trp Ala Ser Ala Gln Asn Ile Ser
30 35 40
GGC GCG GGC TGG AGT GGC ATG GCC GAG GCG ACC TCG CTA GAC ACC ATG      258
Gly Ala Gly Trp Ser Gly Met Ala Glu Ala Thr Ser Leu Asp Thr Met
45 50 55
GCC CAG ATG AAT CAG GCG TTT CGC AAC ATC GTG AAC ATG CTG CAC GGG      306
Ala Gln Met Asn Gln Ala Phe Arg Asn Ile Val Asn Met Leu His Gly
60 65 70
GTG CGT GAC GGG CTG GTT CGC GAC GCC AAC AAC TAC GAG CAG CAA GAG      354
Val Arg Asp Gly Leu Val Arg Asp Ala Asn Asn Tyr Glu Gln Gln Glu
75 80 85
CAG GCC TCC CAG CAG ATC CTC AGC AGC TAACGTCAGC CGCTGCAGCA CAATACT      408
Gln Ala Ser Gln Gln Ile Leu Ser Ser
90 95
TTTACAAGCG AAGGAGAACA GGTTCGATGA CCATCAACTA TCAGTTCGGT GATGTGACG      468
CTCATGGCGC CA      480

```

## (2) INFORMATION FOR SEQ ID NO: 143:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 98 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 143:

```

Met Ala Thr Arg Phe Met Thr Asp Pro His Ala Met Arg Asp Met Ala
 1             5             10             15

Gly Arg Phe Glu Val His Ala Gln Thr Val Glu Asp Glu Ala Arg Arg
      20             25             30

Met Trp Ala Ser Ala Gln Asn Ile Ser Gly Ala Gly Trp Ser Gly Met
      35             40             45

Ala Glu Ala Thr Ser Leu Asp Thr Met Ala Gln Met Asn Gln Ala Phe
      50             55             60

Arg Asn Ile Val Asn Met Leu His Gly Val Arg Asp Gly Leu Val Arg
      65             70             75             80

Asp Ala Asn Asn Tyr Glu Gln Gln Glu Gln Ala Ser Gln Gln Ile Leu
      85             90             95

Ser Ser

```

## (2) INFORMATION FOR SEQ ID NO: 144:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 940 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 86...868
- (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 144:

```

GCCCCAGTCC TCGATCGCCT CATCGCCTTC ACCGCGCCGCC AGCCGACCGC AGGCCACGTG      60

TCCGCCACCT AACGAAAGGA TGATC ATG CCC AAG AGA AGC GAA TAC AGG CAA      112
      Met Pro Lys Arg Ser Glu Tyr Arg Gln
              1             5

GGC ACG CCG AAC TGG GTC GAC CTT CAG ACC ACC GAT CAG TCC GCC GCC      160

```

Gly	Thr	Pro	Asn	Trp	Val	Asp	Leu	Gln	Thr	Thr	Asp	Gln	Ser	Ala	Ala	
10					15					20					25	
AAA	AAG	TTC	TAC	ACA	TCG	TTG	TTC	GGC	TGG	GGT	TAC	GAC	GAC	AAC	CCG	208
Lys	Lys	Phe	Tyr	Thr	Ser	Leu	Phe	Gly	Trp	Gly	Tyr	Asp	Asp	Asn	Pro	
			30					35						40		
GTC	CCC	GGA	GGC	GGT	GGG	GTC	TAT	TCC	ATG	GCC	ACG	CTG	AAC	GGC	GAA	256
Val	Pro	Gly	Gly	Gly	Gly	Val	Tyr	Ser	Met	Ala	Thr	Leu	Asn	Gly	Glu	
			45					50					55			
GCC	GTG	GCC	GCC	ATC	GCA	CCG	ATG	CCC	CCG	GGT	GCA	CCG	GAG	GGG	ATG	304
Ala	Val	Ala	Ala	Ile	Ala	Pro	Met	Pro	Pro	Gly	Ala	Pro	Glu	Gly	Met	
			60				65					70				
CCG	CCG	ATC	TGG	AAC	ACC	TAT	ATC	GCG	GTG	GAC	GAC	GTC	GAT	GCG	GTG	352
Pro	Pro	Ile	Trp	Asn	Thr	Tyr	Ile	Ala	Val	Asp	Asp	Val	Asp	Ala	Val	
		75				80					85					
GTG	GAC	AAG	GTG	GTG	CCC	GGG	GGC	GGG	CAG	GTG	ATG	ATG	CCG	GCC	TTC	400
Val	Asp	Lys	Val	Val	Pro	Gly	Gly	Gly	Gln	Val	Met	Met	Pro	Ala	Phe	
	90					95			100						105	
GAC	ATC	GGC	GAT	GCC	GGC	CGG	ATG	TCG	TTC	ATC	ACC	GAT	CCG	ACC	GGC	448
Asp	Ile	Gly	Asp	Ala	Gly	Arg	Met	Ser	Phe	Ile	Thr	Asp	Pro	Thr	Gly	
				110					115					120		
GCT	GCC	GTG	GGC	CTA	TGG	CAG	GCC	AAT	CGG	CAC	ATC	GGA	GCG	ACG	TTG	496
Ala	Ala	Val	Gly	Leu	Trp	Gln	Ala	Asn	Arg	His	Ile	Gly	Ala	Thr	Leu	
			125					130					135			
GTC	AAC	GAG	ACG	GGC	ACG	CTC	ATC	TGG	AAC	GAA	CTG	CTC	ACG	GAC	AAG	544
Val	Asn	Glu	Thr	Gly	Thr	Leu	Ile	Trp	Asn	Glu	Leu	Leu	Thr	Asp	Lys	
		140						145				150				
CCG	GAT	TTG	GCG	CTA	GCG	TTC	TAC	GAG	GCT	GTG	GTT	GGC	CTC	ACC	CAC	592
Pro	Asp	Leu	Ala	Leu	Ala	Phe	Tyr	Glu	Ala	Val	Val	Gly	Leu	Thr	His	
		155				160					165					
TCG	AGC	ATG	GAG	ATA	GCT	GCG	GGC	CAG	AAC	TAT	CGG	GTG	CTC	AAG	GCC	640
Ser	Ser	Met	Glu	Ile	Ala	Ala	Gly	Gln	Asn	Tyr	Arg	Val	Leu	Lys	Ala	
	170				175					180				185		
GGC	GAC	GCG	GAA	GTC	GGC	GGC	TGT	ATG	GAA	CCG	CCG	ATG	CCC	GGC	GTG	688
Gly	Asp	Ala	Glu	Val	Gly	Gly	Cys	Met	Glu	Pro	Pro	Met	Pro	Gly	Val	
				190					195					200		
CCG	AAT	CAT	TGG	CAC	GTC	TAC	TTT	GCG	GTG	GAT	GAC	GCC	GAC	GCC	ACG	736
Pro	Asn	His	Trp	His	Val	Tyr	Phe	Ala	Val	Asp	Asp	Ala	Asp	Ala	Thr	
			205					210					215			
GCG	GCC	AAA	GCC	GCC	GCA	GCG	GGC	GGC	CAG	GTC	ATT	GCG	GAA	CCG	GCT	784
Ala	Ala	Lys	Ala	Ala	Ala	Ala	Gly	Gly	Gln	Val	Ile	Ala	Glu	Pro	Ala	
		220					225					230				
GAC	ATT	CCG	TCG	GTG	GGC	CGG	TTC	GCC	GTG	TTG	TCC	GAT	CCG	CAG	GGC	832
Asp	Ile	Pro	Ser	Val	Gly	Arg	Phe	Ala	Val	Leu	Ser	Asp	Pro	Gln	Gly	
	235					240					245					

GCG ATC TTC AGT GTG TTG AAG CCC GCA CCG CAG CAA TAGGGAGCAT CCCGGG 884  
 Ala Ile Phe Ser Val Leu Lys Pro Ala Pro Gln Gln  
 250 255 260

CAGGCCCGCC GGCCGGCAGA TTCGGAGAAT GCTAGAAGCT GCCGCCGGCG CCGCCG 940

(2) INFORMATION FOR SEQ ID NO: 145:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 261 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 145:

Met	Pro	Lys	Arg	Ser	Glu	Tyr	Arg	Gln	Gly	Thr	Pro	Asn	Trp	Val	Asp	1	5	10	15
Leu	Gln	Thr	Thr	Asp	Gln	Ser	Ala	Ala	Lys	Lys	Phe	Tyr	Thr	Ser	Leu	20	25	30	
Phe	Gly	Trp	Gly	Tyr	Asp	Asp	Asn	Pro	Val	Pro	Gly	Gly	Gly	Gly	Val	35	40	45	
Tyr	Ser	Met	Ala	Thr	Leu	Asn	Gly	Glu	Ala	Val	Ala	Ala	Ile	Ala	Pro	50	55	60	
Met	Pro	Pro	Gly	Ala	Pro	Glu	Gly	Met	Pro	Pro	Ile	Trp	Asn	Thr	Tyr	65	70	75	80
Ile	Ala	Val	Asp	Asp	Val	Asp	Ala	Val	Val	Asp	Lys	Val	Val	Pro	Gly	85	90	95	
Gly	Gly	Gln	Val	Met	Met	Pro	Ala	Phe	Asp	Ile	Gly	Asp	Ala	Gly	Arg	100	105	110	
Met	Ser	Phe	Ile	Thr	Asp	Pro	Thr	Gly	Ala	Ala	Val	Gly	Leu	Trp	Gln	115	120	125	
Ala	Asn	Arg	His	Ile	Gly	Ala	Thr	Leu	Val	Asn	Glu	Thr	Gly	Thr	Leu	130	135	140	
Ile	Trp	Asn	Glu	Leu	Leu	Thr	Asp	Lys	Pro	Asp	Leu	Ala	Leu	Ala	Phe	145	150	155	160
Tyr	Glu	Ala	Val	Val	Gly	Leu	Thr	His	Ser	Ser	Met	Glu	Ile	Ala	Ala	165	170	175	
Gly	Gln	Asn	Tyr	Arg	Val	Leu	Lys	Ala	Gly	Asp	Ala	Glu	Val	Gly	Gly	180	185	190	
Cys	Met	Glu	Pro	Pro	Met	Pro	Gly	Val	Pro	Asn	His	Trp	His	Val	Tyr	195	200	205	



220

Phe Ala Val Asp Asp Ala Asp Ala Thr Ala Ala Lys Ala Ala Ala Ala  
 210 215 220

Gly Gly Gln Val Ile Ala Glu Pro Ala Asp Ile Pro Ser Val Gly Arg  
 225 230 235 240

Phe Ala Val Leu Ser Asp Pro Gln Gly Ala Ile Phe Ser Val Leu Lys  
 245 250 255

Pro Ala Pro Gln Gln  
 260

(2) INFORMATION FOR SEQ ID NO: 146:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 280 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 47...247
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 146:

CCGAAAGGCG GTGCACCGCA CCCAGAAGAA AAGGAAAGAT CGAGAA ATG CCA CAG	55
Met Pro Gln	
1	
GGA ACT GTG AAG TGG TTC AAC GCG GAG AAG GGG TTC GGC TTT ATC GCC	103
Gly Thr Val Lys Trp Phe Asn Ala Glu Lys Gly Phe Gly Phe Ile Ala	
5 10 15	
CCC GAA GAC GGT TCC GCG GAT GTA TTT GTC CAC TAC ACG GAG ATC CAG	151
Pro Glu Asp Gly Ser Ala Asp Val Phe Val His Tyr Thr Glu Ile Gln	
20 25 30 35	
GGA ACG GGC TTC CGC ACC CTT GAA GAA AAC CAG AAG GTC GAG TTC GAG	199
Gly Thr Gly Phe Arg Thr Leu Glu Glu Asn Gln Lys Val Glu Phe Glu	
40 45 50	
ATC GGC CAC AGC CCT AAG GGC CCC CAG GCC ACC GGA GTC CGC TCG CTC T	248
Ile Gly His Ser Pro Lys Gly Pro Gln Ala Thr Gly Val Arg Ser Leu	
55 60 65	
GAGTTACCCC CGCGAGCAGA CGCAAAAAGC CC	280

(2) INFORMATION FOR SEQ ID NO: 147:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 67 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein  
 (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 147:

```

Met Pro Gln Gly Thr Val Lys Trp Phe Asn Ala Glu Lys Gly Phe Gly
 1              5              10              15

Phe Ile Ala Pro Glu Asp Gly Ser Ala Asp Val Phe Val His Tyr Thr
              20              25              30

Glu Ile Gln Gly Thr Gly Phe Arg Thr Leu Glu Glu Asn Gln Lys Val
              35              40              45

Glu Phe Glu Ile Gly His Ser Pro Lys Gly Pro Gln Ala Thr Gly Val
              50              55              60

Arg Ser Leu
65
  
```

(2) INFORMATION FOR SEQ ID NO: 148:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 540 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: Coding Sequence  
 (B) LOCATION: 105...491  
 (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 148:

```

ATCGTGTTCGT ATCGAGAACC CCGGCCGGTA TCAGAACGCG CCAGAGCGCA AACCTTTATA      60

ACTTCGTGTC CCAAATGTGA CGACCATGGA CCAAGGTTCC TGAG ATG AAC CTA CGG      116
                               Met Asn Leu Arg
                               1

CGC CAT CAG ACC CTG ACG CTG CGA CTG CTG GCG GCA TCC GCG GGC ATT      164
Arg His Gln Thr Leu Thr Leu Arg Leu Leu Ala Ala Ser Ala Gly Ile
 5              10              15              20

CTC AGC GCC GCG GCC TTC GCC GCG CCA GCA CAG GCA AAC CCC GTC GAC      212
Leu Ser Ala Ala Ala Phe Ala Ala Pro Ala Gln Ala Asn Pro Val Asp
              25              30              35

GAC GCG TTC ATC GCC GCG CTG AAC AAT GCC GGC GTC AAC TAC GGC GAT      260
Asp Ala Phe Ile Ala Ala Leu Asn Asn Ala Gly Val Asn Tyr Gly Asp
              40              45              50

CCG GTC GAC GCC AAA GCG CTG GGT CAG TCC GTC TGC CCG ATC CTG GCC      308
Pro Val Asp Ala Lys Ala Leu Gly Gln Ser Val Cys Pro Ile Leu Ala
              55              60              65
  
```

GAG CCC GGC GGG TCG TTT AAC ACC GCG GTA GCC AGC GTT GTG GCG CGC 356  
 Glu Pro Gly Gly Ser Phe Asn Thr Ala Val Ala Ser Val Val Ala Arg  
       70                      75                      80

GCC CAA GGC ATG TCC CAG GAC ATG GCG CAA ACC TTC ACC AGT ATC GCG 404  
 Ala Gln Gly Met Ser Gln Asp Met Ala Gln Thr Phe Thr Ser Ile Ala  
       85                      90                      95                      100

ATT TCG ATG TAC TGC CCC TCG GTG ATG GCA GAC GTC GCC AGC GGC AAC 452  
 Ile Ser Met Tyr Cys Pro Ser Val Met Ala Asp Val Ala Ser Gly Asn  
                       105                      110                      115

CTG CCG GCC CTG CCA GAC ATG CCG GGG CTG CCC GGG TCC TAGGCGTGCG CG 503  
 Leu Pro Ala Leu Pro Asp Met Pro Gly Leu Pro Gly Ser  
                       120                      125

GCTCCTAGCC GGTCCCTAAC GGATCGATCG TGGATGC 540

## (2) INFORMATION FOR SEQ ID NO: 149:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 129 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 149:

Met Asn Leu Arg Arg His Gln Thr Leu Thr Leu Arg Leu Leu Ala Ala  
   1                      5                      10                      15

Ser Ala Gly Ile Leu Ser Ala Ala Ala Phe Ala Ala Pro Ala Gln Ala  
                       20                      25                      30

Asn Pro Val Asp Asp Ala Phe Ile Ala Ala Leu Asn Asn Ala Gly Val  
                       35                      40                      45

Asn Tyr Gly Asp Pro Val Asp Ala Lys Ala Leu Gly Gln Ser Val Cys  
   50                      55                      60

Pro Ile Leu Ala Glu Pro Gly Gly Ser Phe Asn Thr Ala Val Ala Ser  
   65                      70                      75                      80

Val Val Ala Arg Ala Gln Gly Met Ser Gln Asp Met Ala Gln Thr Phe  
                       85                      90                      95

Thr Ser Ile Ala Ile Ser Met Tyr Cys Pro Ser Val Met Ala Asp Val  
                       100                      105                      110

Ala Ser Gly Asn Leu Pro Ala Leu Pro Asp Met Pro Gly Leu Pro Gly  
                       115                      120                      125

Ser

## (2) INFORMATION FOR SEQ ID NO: 150:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 400 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 25...354
- (D) OTHER INFORMATION:

## (ix) FEATURE:

- (A) NAME/KEY: mat\_peptide
- (B) LOCATION: 109..357

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 150:

ATAGTTTGGG GAAGGTGTCC ATAA ATG AGG CTG TCG TTG ACC GCA TTG AGC	51
Met Arg Leu Ser Leu Thr Ala Leu Ser	
-28 -25 -20	
GCC GGT GTA GGC GCC GTG GCA ATG TCG TTG ACC GTC GGG GCC GGG GTC	99
Ala Gly Val Gly Ala Val Ala Met Ser Leu Thr Val Gly Ala Gly Val	
-15 -10 -5	
GCC TCC GCA GAT CCC GTG GAC GCG GTC ATT AAC ACC ACC TGC AAT TAC	147
Ala Ser Ala Asp Pro Val Asp Ala Val Ile Asn Thr Thr Cys Asn Tyr	
1 5 10	
GGG CAG GTA GTA GCT GCG CTC AAC GCG ACG GAT CCG GGG GCT GCC GCA	195
Gly Gln Val Val Ala Ala Leu Asn Ala Thr Asp Pro Gly Ala Ala Ala	
15 20 25	
CAG TTC AAC GCC TCA CCG GTG GCG CAG TCC TAT TTG CGC AAT TTC CTC	243
Gln Phe Asn Ala Ser Pro Val Ala Gln Ser Tyr Leu Arg Asn Phe Leu	
30 35 40 45	
GCC GCA CCG CCA CCT CAG CGC GCT GCC ATG GCC GCG CAA TTG CAA GCT	291
Ala Ala Pro Pro Pro Gln Arg Ala Ala Met Ala Ala Gln Leu Gln Ala	
50 55 60	
GTG CCG GGG GCG GCA CAG TAC ATC GGC CTT GTC GAG TCG GTT GCC GGC	339
Val Pro Gly Ala Ala Gln Tyr Ile Gly Leu Val Glu Ser Val Ala Gly	
65 70 75	
TCC TGC AAC AAC TAT TAAGCCCATG CGGGCCCAT CCCGCGACCC GGCATCGTCG	394
Ser Cys Asn Asn Tyr	
80	
CCGGGG	400

## (2) INFORMATION FOR SEQ ID NO: 151:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 110 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 151:

```

Met Arg Leu Ser Leu Thr Ala Leu Ser Ala Gly Val Gly Ala Val Ala
-28          -25          -20          -15

Met Ser Leu Thr Val Gly Ala Gly Val Ala Ser Ala Asp Pro Val Asp
          -10          -5          1

Ala Val Ile Asn Thr Thr Cys Asn Tyr Gly Gln Val Val Ala Ala Leu
 5          10          15          20

Asn Ala Thr Asp Pro Gly Ala Ala Ala Gln Phe Asn Ala Ser Pro Val
          25          30          35

Ala Gln Ser Tyr Leu Arg Asn Phe Leu Ala Ala Pro Pro Pro Gln Arg
          40          45          50

Ala Ala Met Ala Ala Gln Leu Gln Ala Val Pro Gly Ala Ala Gln Tyr
          55          60          65

Ile Gly Leu Val Glu Ser Val Ala Gly Ser Cys Asn Asn Tyr
          70          75          80

```

## (2) INFORMATION FOR SEQ ID NO: 152:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 990 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: cDNA

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence  
 (B) LOCATION: 93...890  
 (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 152:

```

AATAGTAATA TCGCTGTGCG GTTGCAAAAC GTGTGACCGA GGTTCGCAG TCGAGCGCTG      60
CGGGCCGCCT TCGAGGAGGA CGAACCACAG TC ATG ACG AAC ATC GTG GTC CTG      113
          Met Thr Asn Ile Val Val Leu
          1          5

ATC AAG CAG GTC CCA GAT ACC TGG TCG GAG CGC AAG CTG ACC GAC GGC      161
Ile Lys Gln Val Pro Asp Thr Trp Ser Glu Arg Lys Leu Thr Asp Gly
          10          15          20

GAT TTC ACG CTG GAC CGC GAG GCC GCC GAC GCG GTG CTG GAC GAG ATC      209

```

Asp	Phe	Thr	Leu	Asp	Arg	Glu	Ala	Ala	Asp	Ala	Val	Leu	Asp	Glu	Ile	
25						30					35					
AAC	GAG	CGC	GCC	GTG	GAG	GAA	GCG	CTA	CAG	ATT	CGG	GAG	AAA	GAG	GCC	257
Asn	Glu	Arg	Ala	Val	Glu	Glu	Ala	Leu	Gln	Ile	Arg	Glu	Lys	Glu	Ala	
40					45				50					55		
GCC	GAC	GGC	ATC	GAA	GGG	TCG	GTA	ACC	GTG	CTG	ACG	GCG	GGC	CCC	GAG	305
Ala	Asp	Gly	Ile	Glu	Gly	Ser	Val	Thr	Val	Leu	Thr	Ala	Gly	Pro	Glu	
				60					65					70		
CGC	GCC	ACC	GAG	GCG	ATC	CGC	AAG	GCG	CTG	TCG	ATG	GGT	GCC	GAC	AAG	353
Arg	Ala	Thr	Glu	Ala	Ile	Arg	Lys	Ala	Leu	Ser	Met	Gly	Ala	Asp	Lys	
			75					80					85			
GCC	GTC	CAC	CTA	AAG	GAC	GAC	GGC	ATG	CAC	GGC	TCG	GAC	GTC	ATC	CAA	401
Ala	Val	His	Leu	Lys	Asp	Asp	Gly	Met	His	Gly	Ser	Asp	Val	Ile	Gln	
		90					95					100				
ACC	GGG	TGG	GCT	TTG	GCG	CGC	GCG	TTG	GGC	ACC	ATC	GAG	GGC	ACC	GAG	449
Thr	Gly	Trp	Ala	Leu	Ala	Arg	Ala	Leu	Gly	Thr	Ile	Glu	Gly	Thr	Glu	
	105					110						115				
CTG	GTG	ATC	GCA	GGC	AAC	GAA	TCG	ACC	GAC	GGG	GTG	GGC	GGT	GCG	GTG	497
Leu	Val	Ile	Ala	Gly	Asn	Glu	Ser	Thr	Asp	Gly	Val	Gly	Gly	Ala	Val	
120					125					130					135	
CCG	GCC	ATC	ATC	GCC	GAG	TAC	CTG	GGC	CTG	CCG	CAG	CTC	ACC	CAC	CTG	545
Pro	Ala	Ile	Ile	Ala	Glu	Tyr	Leu	Gly	Leu	Pro	Gln	Leu	Thr	His	Leu	
				140					145					150		
CGC	AAA	GTG	TCG	ATC	GAG	GGC	GGC	AAG	ATC	ACC	GGC	GAG	CGT	GAG	ACC	593
Arg	Lys	Val	Ser	Ile	Glu	Gly	Gly	Lys	Ile	Thr	Gly	Glu	Arg	Glu	Thr	
			155					160					165			
GAT	GAG	GGC	GTA	TTC	ACC	CTC	GAG	GCC	ACG	CTG	CCC	GCG	GTG	ATC	AGC	641
Asp	Glu	Gly	Val	Phe	Thr	Leu	Glu	Ala	Thr	Leu	Pro	Ala	Val	Ile	Ser	
		170					175					180				
GTG	AAC	GAG	AAG	ATC	AAC	GAG	CCG	CGC	TTC	CCG	TCC	TTC	AAA	GGC	ATC	689
Val	Asn	Glu	Lys	Ile	Asn	Glu	Pro	Arg	Phe	Pro	Ser	Phe	Lys	Gly	Ile	
	185					190					195					
ATG	GCC	GCC	AAG	AAG	AAG	GAA	GTT	ACC	GTG	CTG	ACC	CTG	GCC	GAG	ATC	737
Met	Ala	Ala	Lys	Lys	Lys	Glu	Val	Thr	Val	Leu	Thr	Leu	Ala	Glu	Ile	
200					205					210				215		
GGT	GTC	GAG	AGC	GAC	GAG	GTG	GGG	CTG	GCC	AAC	GCC	GGA	TCC	ACC	GTG	785
Gly	Val	Glu	Ser	Asp	Glu	Val	Gly	Leu	Ala	Asn	Ala	Gly	Ser	Thr	Val	
				220					225					230		
CTG	GCG	TCG	ACG	CCC	AAA	CCG	GCC	AAG	ACT	GCC	GGG	GAG	AAG	GTC	ACC	833
Leu	Ala	Ser	Thr	Pro	Lys	Pro	Ala	Lys	Thr	Ala	Gly	Glu	Lys	Val	Thr	
			235					240					245			
GAC	GAG	GGT	GAA	GGC	GGC	AAC	CAG	ATC	GTG	CAG	TAC	CTG	GTT	GCC	CAG	881
Asp	Glu	Gly	Glu	Gly	Gly	Asn	Gln	Ile	Val	Gln	Tyr	Leu	Val	Ala	Gln	
		250					255					260				

AAA ATC ATC TAAGACATAC GCACCTCCCA AAGACGAGAG CGATATAACC CATGGCTGA 939  
 Lys Ile Ile  
 265

AGTACTGGTG CTCGTTGAGC ACGCTGAAGG CGCGTTAAAG AAGGTCAGCG C 990

(2) INFORMATION FOR SEQ ID NO: 153:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 266 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 153:

Met	Thr	Asn	Ile	Val	Val	Leu	Ile	Lys	Gln	Val	Pro	Asp	Thr	Trp	Ser	
1				5					10					15		
Glu	Arg	Lys	Leu	Thr	Asp	Gly	Asp	Phe	Thr	Leu	Asp	Arg	Glu	Ala	Ala	
			20					25					30			
Asp	Ala	Val	Leu	Asp	Glu	Ile	Asn	Glu	Arg	Ala	Val	Glu	Glu	Ala	Leu	
			35				40					45				
Gln	Ile	Arg	Glu	Lys	Glu	Ala	Ala	Asp	Gly	Ile	Glu	Gly	Ser	Val	Thr	
	50					55				60						
Val	Leu	Thr	Ala	Gly	Pro	Glu	Arg	Ala	Thr	Glu	Ala	Ile	Arg	Lys	Ala	
	65				70				75					80		
Leu	Ser	Met	Gly	Ala	Asp	Lys	Ala	Val	His	Leu	Lys	Asp	Asp	Gly	Met	
			85					90						95		
His	Gly	Ser	Asp	Val	Ile	Gln	Thr	Gly	Trp	Ala	Leu	Ala	Arg	Ala	Leu	
			100					105					110			
Gly	Thr	Ile	Glu	Gly	Thr	Glu	Leu	Val	Ile	Ala	Gly	Asn	Glu	Ser	Thr	
	115					120						125				
Asp	Gly	Val	Gly	Gly	Ala	Val	Pro	Ala	Ile	Ile	Ala	Glu	Tyr	Leu	Gly	
	130					135					140					
Leu	Pro	Gln	Leu	Thr	His	Leu	Arg	Lys	Val	Ser	Ile	Glu	Gly	Gly	Lys	
	145				150					155				160		
Ile	Thr	Gly	Glu	Arg	Glu	Thr	Asp	Glu	Gly	Val	Phe	Thr	Leu	Glu	Ala	
			165					170					175			
Thr	Leu	Pro	Ala	Val	Ile	Ser	Val	Asn	Glu	Lys	Ile	Asn	Glu	Pro	Arg	
			180					185					190			
Phe	Pro	Ser	Phe	Lys	Gly	Ile	Met	Ala	Ala	Lys	Lys	Lys	Glu	Val	Thr	
	195					200						205				
Val	Leu	Thr	Leu	Ala	Glu	Ile	Gly	Val	Glu	Ser	Asp	Glu	Val	Gly	Leu	

210		215		220
Ala Asn Ala Gly Ser Thr Val Leu Ala Ser Thr Pro Lys Pro Ala Lys				
225		230		240
Thr Ala Gly Glu Lys Val Thr Asp Glu Gly Glu Gly Gly Asn Gln Ile				
	245		250	255
Val Gln Tyr Leu Val Ala Gln Lys Ile Ile				
	260		265	

## (2) INFORMATION FOR SEQ ID NO: 154:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 25 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 154:

CTGAGATCTA TGAACCTACG GCGCC 25

## (2) INFORMATION FOR SEQ ID NO: 155:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 35 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 155:

CTCCCATGGT ACCCTAGGAC CCGGCAGCC CCGGC 35

## (2) INFORMATION FOR SEQ ID NO: 156:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 29 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 156:

CTGAGATCTA TGAGGCTGTC GTTGACCGC 29

## (2) INFORMATION FOR SEQ ID NO: 157:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 30 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear



(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 157:

CTCCCCGGGC TTAATAGTTG TTGCAGGAGC

30

(2) INFORMATION FOR SEQ ID NO: 158:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 158:

GCTTAGATCT ATGATTTTCT GGGCAACCAG GTA

33

(2) INFORMATION FOR SEQ ID NO: 159:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 159:

GCTTCCATGG GCGAGGCACA GGCGTGGGAA

30

(2) INFORMATION FOR SEQ ID NO: 160:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 160:

CTGAGATCTA GAATGCCACA GGGAAGTGTG

30

(2) INFORMATION FOR SEQ ID NO: 161:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 161:

TCTCCCGGGG GTAAGTCAGA GCGAGCGGAC

30

(2) INFORMATION FOR SEQ ID NO: 162:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 27 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 162:

CTGAGATCTA TGAACGTCAC CGTATCC

27

(2) INFORMATION FOR SEQ ID NO: 163:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 27 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 163:

TCTCCCGGGG CTCACCCACC GGCCACG

27

(2) INFORMATION FOR SEQ ID NO: 164:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 30 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 164:

CTGAGATCTA TGGCAACACG TTTTATGACG

30

(2) INFORMATION FOR SEQ ID NO: 165:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 30 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 165:

CTCCCCGGGT TAGCTGCTGA GGATCTGCTH

30

(2) INFORMATION FOR SEQ ID NO: 166:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 31 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 166:

CTGAAGATCT ATGCCCAAGA GAAGCGAATA C

31

(2) INFORMATION FOR SEQ ID NO: 167:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 31 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 167:

CGGCAGCTGC TAGCATTCTC CGAATCTGCC G

31

(2) INFORMATION FOR SEQ ID NO: 168:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 168:

Pro	Gln	Gly	Thr	Val	Lys	Trp	Phe	Asn	Ala	Glu	Lys	Gly	Phe	Gly
1				5				10					15	

(2) INFORMATION FOR SEQ ID NO: 169:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(ix) FEATURE:

- (A) NAME/KEY: Other
- (B) LOCATION: 15
- (D) OTHER INFORMATION: Xaa is unknown

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 169:

Asn	Val	Thr	Val	Ser	Ile	Pro	Thr	Ile	Leu	Arg	Pro	Xaa	Xaa	Xaa
1				5				10					15	

(2) INFORMATION FOR SEQ ID NO: 170:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(ix) FEATURE:

(A) NAME/KEY: Other

(B) LOCATION: 1

(D) OTHER INFORMATION: Thr Could also be Ala

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 170:

Thr	Arg	Phe	Met	Thr	Asp	Pro	His	Ala	Met	Arg	Asp	Met	Ala	Gly
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO: 171:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: None

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 171:

Pro	Lys	Arg	Ser	Glu	Tyr	Arg	Gln	Gly	Thr	Pro	Asn	Trp	Val	Asp
1		5		10				15						

(2) INFORMATION FOR SEQ ID NO:172:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 404 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:172:

Met	Ala	Thr	Val	Asn	Arg	Ser	Arg	His	His	His	His	His	His	His
1				5				10					15	
Ile	Glu	Gly	Arg	Ser	Phe	Ser	Arg	Pro	Gly	Leu	Pro	Val	Glu	Tyr
			20					25					30	
Gln	Val	Pro	Ser	Pro	Ser	Met	Gly	Arg	Asp	Ile	Lys	Val	Gln	Phe
			35				40					45		
Ser	Gly	Gly	Asn	Asn	Ser	Pro	Ala	Val	Tyr	Leu	Leu	Asp	Gly	Leu
			50				55				60			
Ala	Gln	Asp	Asp	Tyr	Asn	Gly	Trp	Asp	Ile	Asn	Thr	Pro	Ala	Phe
65					70				75				80	
Trp	Tyr	Tyr	Gln	Ser	Gly	Leu	Ser	Ile	Val	Met	Pro	Val	Gly	Gly
			85					90					95	
Ser	Ser	Phe	Tyr	Ser	Asp	Trp	Tyr	Ser	Pro	Ala	Cys	Gly	Lys	Ala
			100					105					110	
Cys	Gln	Thr	Tyr	Lys	Trp	Glu	Thr	Phe	Leu	Thr	Ser	Glu	Leu	Pro
			115				120					125		
Trp	Leu	Ser	Ala	Asn	Arg	Ala	Val	Lys	Pro	Thr	Gly	Ser	Ala	Ile

[illegible][illegible]

Figure 1. The effect of the concentration of the *Agaricus bisporus* spores on the growth of *Agaricus bisporus* on the substrate.

- Figure 1. The effect of the concentration of the *Agaricus bisporus* spores on the growth of *Agaricus bisporus* on the substrate.

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				85					90					95		
Ile	Ser	Glu	Ala	Gly	Gln	Ala	Met	Ala	Ser	Thr	Glu	Gly	Asn	Val	Thr	
			100					105					110			
Gly	Met	Phe	Ala	Lys	Leu	Phe	Ser	Arg	Pro	Gly	Leu	Pro	Val	Glu	Tyr	
		115					120					125				
Leu	Gln	Val	Pro	Ser	Pro	Ser	Met	Gly	Arg	Asp	Ile	Lys	Val	Gln	Phe	
	130					135					140					
Gln	Ser	Gly	Gly	Asn	Asn	Ser	Pro	Ala	Val	Tyr	Leu	Leu	Asp	Gly	Leu	
145				150					155						160	
Arg	Ala	Gln	Asp	Asp	Tyr	Asn	Gly	Trp	Asp	Ile	Asn	Thr	Pro	Ala	Phe	
			165					170						175		
Glu	Trp	Tyr	Tyr	Gln	Ser	Gly	Leu	Ser	Ile	Val	Met	Pro	Val	Gly	Gly	
		180					185					190				
Gln	Ser	Ser	Phe	Tyr	Ser	Asp	Trp	Tyr	Ser	Pro	Ala	Cys	Gly	Lys	Ala	
	195					200					205					
Gly	Cys	Gln	Thr	Tyr	Lys	Trp	Glu	Thr	Phe	Leu	Thr	Ser	Glu	Leu	Pro	
	210				215				220							
Gln	Trp	Leu	Ser	Ala	Asn	Arg	Ala	Val	Lys	Pro	Thr	Gly	Ser	Ala	Ala	
225				230					235						240	
Ile	Gly	Leu	Ser	Met	Ala	Gly	Ser	Ser	Ala	Met	Ile	Leu	Ala	Ala	Tyr	
			245					250					255			
His	Pro	Gln	Gln	Phe	Ile	Tyr	Ala	Gly	Ser	Leu	Ser	Ala	Leu	Leu	Asp	
		260					265					270				
Pro	Ser	Gln	Gly	Met	Gly	Pro	Ser	Leu	Ile	Gly	Leu	Ala	Met	Gly	Asp	
	275					280					285					
Ala	Gly	Gly	Tyr	Lys	Ala	Ala	Asp	Met	Trp	Gly	Pro	Ser	Ser	Asp	Pro	
	290				295						300					
Ala	Trp	Glu	Arg	Asn	Asp	Pro	Thr	Gln	Gln	Ile	Pro	Lys	Leu	Val	Ala	
305				310					315					320		
Asn	Asn	Thr	Arg	Leu	Trp	Val	Tyr	Cys	Gly	Asn	Gly	Thr	Pro	Asn	Glu	
			325					330					335			
Leu	Gly	Gly	Ala	Asn	Ile	Pro	Ala	Glu	Phe	Leu	Glu	Asn	Phe	Val	Arg	
		340				345						350				
Ser	Ser	Asn	Leu	Lys	Phe	Gln	Asp	Ala	Tyr	Asn	Ala	Ala	Gly	Gly	His	
	355					360					365					
Asn	Ala	Val	Phe	Asn	Phe	Pro	Pro	Asn	Gly	Thr	His	Ser	Trp	Glu	Tyr	
	370				375				380							
Trp	Gly	Ala	Gln	Leu	Asn	Ala	Met	Lys	Gly	Asp	Leu	Gln	Ser	Ser	Leu	
385				390					395					400		
Gly	Ala	Gly														

The invention will now be further described by the following numbered paragraphs.

1. A substantially pure or isolated polypeptide which
  - a) consists of the amino acid sequence as shown in SEQ ID NO: 88, or
  - 5      b) consists essentially of the amino acid sequence shown in SEQ ID NO: 88, and is immunologically equivalent to the amino acid sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a
  - 10      diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex, or
  - 15      c) consists essentially of an amino acid sequence with a sequence identity of at least 80% with SEQ ID NO: 88, and which is at least 6 contiguous amino acid residues of SEQ ID NO: 88, and is immunologically equivalent to the amino acid sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex; wherein "sequence identity" is a measure of the degree of similarity between two amino acid sequences of equal length calculated as  $(N_{ref} - N_{dif}) * 100 / N_{ref}$ , wherein  $N_{dif}$  is the total number of non-identical residues in the two sequences when aligned and wherein  $N_{ref}$  is the number of residues in one of the sequences.
  - 20      2. A substantially pure or isolated polypeptide which consists essentially of:
    - a) at least 6 contiguous amino acid residues of SEQ ID NO:88, or
    - b) an amino acid sequence with a sequence identity of at least 80% with SEQ ID NO. 88, or
    - 25      c) an amino acid sequence with a sequence identity of at least 80% with a); wherein the polypeptide is immunologically equivalent to the amino acid sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of
    - 30      eliciting a diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex; wherein "sequence identity" is a measure of the degree of similarity between two amino acid sequences of equal length calculated as  $(N_{ref} - N_{dif}) * 100 / N_{ref}$ , wherein  $N_{dif}$  is the total number of non-identical residues in the two sequences when aligned and wherein  $N_{ref}$  is the number of residues in one of the sequences.
    - 35      3. A substantially pure or isolated polypeptide which consists of the amino acid sequence as shown in SEQ ID NO: 88.
    - 40

4. A substantially pure or isolated polypeptide which consists essentially of the amino acid sequence as shown in SEQ ID NO: 88.

5. A substantially pure or isolated polypeptide which consists essentially of an amino acid sequence with a sequence identity of at least 80% with SEQ ID NO: 88, and  
5 which is at least 6 contiguous amino acid residues of SEQ ID NO: 88 wherein the polypeptide is immunologically equivalent to the amino acid sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a diagnostically significant immune response indicating previous or  
10 ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex; wherein "sequence identity" is a measure of the degree of similarity between two amino acid sequences of equal length calculated as  $(N_{ref} - N_{dif}) * 100 / N_{ref}$ , wherein  $N_{dif}$  is the total number of non-identical residues in the two sequences when aligned and wherein  $N_{ref}$  is the number of residues in one of the sequences.

15 6. A substantially pure or isolated polypeptide which consists essentially of at least 6 contiguous amino acid residues of SEQ ID NO: 88, wherein the polypeptide is immunologically equivalent to the amino acid sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of  
20 eliciting a diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex.

7. The polypeptide of paragraph 1 or 2 consisting essentially of a T cell epitope of SEQ ID NO: 88 that is a non-naturally occurring polypeptide that induces a release of  
25 IFN- $\gamma$  from primed memory T-lymphocytes withdrawn from a mouse within 2 weeks of primary infection or within 4 days after the mouse has been re-challenge infected with mycobacteria belonging to the tuberculosis complex, the induction performed by the addition of the polypeptide of a suspension comprising about 200,000 spleen cells per ml, the addition of the polypeptide resulting in a concentration of 1-4  $\mu$ g polypeptide per ml  
30 suspension, the release of IFN- $\gamma$  being assessable by determination of IFN- $\gamma$  in supernatant harvested 2 days after the addition of the polypeptide to the suspension, and elicits a delayed type hypersensitivity reaction.

8. A substantially pure or isolated polypeptide which consists essentially of an amino acid sequence with a sequence identity of at least 80% with SEQ ID NO: 88, or  
35 wherein the polypeptide is immunologically equivalent to the amino acid sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the  
40 tuberculosis complex; wherein "sequence identity" is a measure of the degree of similarity between two amino acid sequences of equal length calculated as  $(N_{ref} - N_{dif}) * 100 / N_{ref}$ , wherein  $N_{dif}$  is the total number of non-identical residues in the two sequences when aligned and wherein  $N_{ref}$  is the number of residues in one of the sequences.



9. A substantially pure or isolated polypeptide which consists essentially of a first amino acid sequence with a sequence identity of at least 80% with a second amino acid sequence that consists essentially of at least 6 contiguous amino acid residues of SEQ ID NO: 88, wherein the polypeptide is immunologically equivalent to the amino acid  
5 sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex; wherein "sequence identity" is a  
10 measure of the degree of similarity between two amino acid sequences of equal length calculated as  $(N_{ref} - N_{dif}) * 100 / N_{ref}$ , wherein  $N_{dif}$  is the total number of non-identical residues in the two sequences when aligned and wherein  $N_{ref}$  is the number of residues in one of the sequences.
10. The polypeptide according to any one of paragraphs 1 or 2 in essentially  
15 pure form.
11. The polypeptide according to any one of paragraphs 1 or 2 which consists essentially of an epitope for a T-helper cell.
12. A substantially pure or isolated polypeptide which consists essentially of at least 7 contiguous amino acid residues of SEQ ID NO: 88 and is immunologically  
20 equivalent to the amino acid sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex.
- 25 13. A substantially pure or isolated polypeptide which consists essentially of at least 12 contiguous amino acid residues of SEQ ID NO: 88 and is immunologically equivalent to the amino acid sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a  
30 diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex.
14. A substantially pure or isolated polypeptide which consists essentially of at least 20 contiguous amino acid residues of SEQ ID NO: 88 and is immunologically  
35 equivalent to the amino acid sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex.
15. A substantially pure or isolated polypeptide which consists essentially of at  
40 least 30 contiguous amino acid residues of SEQ ID NO: 88 and is immunologically equivalent to the amino acid sequence shown in SEQ ID NO: 88 with respect to the ability of evoking a protective immune response in mice against infections with mycobacteria belonging to the tuberculosis complex or with respect to the ability of eliciting a

diagnostically significant immune response indicating previous or ongoing sensitization with antigens derived from mycobacteria belonging to the tuberculosis complex.

16. The polypeptide according to any one of paragraphs 1 or 2 which is free from any signal sequence.

- 5        17. The polypeptide according to any one of paragraphs 1 or 2 which
- a) induces a release of IFN- $\gamma$  from primed memory T-lymphocytes withdrawn from a mouse within 2 weeks of primary infection or within 4 days after the mouse has been re-challenge infected with mycobacteria belonging to the tuberculosis complex, the induction performed by the addition of the
- 10       polypeptide to a suspension comprising about 200,000 spleen cells per ml, the addition of the polypeptide resulting in a concentration of 1-4  $\mu$ g polypeptide per ml suspension, the release of IFN- $\gamma$  being assessable by determination of IFN- $\gamma$  in supernatant harvested 2 days after the addition of the polypeptide to the suspension, and/or
- 15       b) induces a release of IFN- $\gamma$  of at least 300 pg above background level from about 1000,000 human PBMC (peripheral blood mononuclear cells) per ml isolated from TB patients in the first phase of infection, or from healthy BCG vaccinated donors, or from healthy contacts to TB patients, the induction being performed by the addition of the polypeptide to a suspension
- 20       comprising the about 1,000,000 PBMC per ml, the addition of the polypeptide resulting in a concentration of 1-4  $\mu$ g polypeptide per ml suspension, the release of IFN- $\gamma$  being assessable by determination of IFN- $\gamma$  in supernatant harvested 2 days after the addition of the polypeptide to the suspension; and/or
- 25       c) induces an IFN- $\gamma$  release from bovine PBMC derived from animals previously sensitized with mycobacteria belonging to the tuberculosis complex, said release being at least two times the release observed from bovine PBMC derived from animals not previously sensitized with mycobacteria belonging to the tuberculosis complex.

30       18. The polypeptide according to any one of paragraphs 1 or 2, wherein the sequence identity is at least 85%.

         19. The polypeptide according to paragraph 18, wherein the sequence identity is at least 90%, or at least 91%, or at least 92%, or at least 93%, or at least 94%, or at least 95%, or at least 96%, or at least 97%, or at least 98%, or at least 99%, or at least

35 99.5%.

         20. The polypeptide according to paragraph 18, wherein the sequence identity is at least 95%.

         21. The polypeptide according to paragraph 20, wherein the sequence identity is at least 96%, or at least 97%, or at least 98%, or at least 99%, or at least 99.5%.

40       22. A fusion polypeptide comprising at least one polypeptide according to any of paragraphs 1 or 2 and at least one fusion partner.

         23. A fusion polypeptide, consisting essentially of at least one polypeptide according to any one of paragraphs 1 or 2 and at least one fusion partner selected from the group consisting of ESAT-6, at least one T-cell epitope of ESAT-6, MPB64, at least one

T-cell epitope of MPB64, MPT64 at least one T-cell epitope of MPT64, and MPB59 and at least one T-cell epitope of MPB59.

24. The polypeptide according to any one of paragraphs 1 or 2 which is lipidated.

5 25. A composition comprising a polypeptide according to any one of paragraphs 1 or 2 and pharmaceutically acceptable carrier, vehicle or adjuvant.

26. An immunological composition comprising a polypeptide according to any one of paragraphs 1 or 2.

10 27. The immunological composition according to paragraph 26, further comprising an immunologically and pharmaceutically acceptable carrier, vehicle or adjuvant.

28. The immunological composition according to paragraph 27, wherein the carrier is a polymer to which the polypeptide(s) is/are bound by hydrophobic non-covalent interaction; the vehicle is selected from the group consisting of a diluent and a suspending agent; and the adjuvant is Freund's incomplete adjuvant.

29. An immunological composition comprising at least two different polypeptides according to any one of paragraphs 1 or 2.

30. An immunological composition comprising 3-20 different polypeptides according to any one of paragraphs 1 or 2.

20 31. A skin test reagent comprising the immunological composition of paragraph 26.

32. A composition for diagnosing tuberculosis in an animal, including a human being, comprising a polypeptide according to any one of paragraphs 1 or 2 optionally in combination with a means for detection.

25 33. A fusion polypeptide comprising at least one polypeptide according to any one of paragraphs 3 or 4 and at least one fusion partner.

34. A fusion polypeptide, consisting essentially of at least one polypeptide according to any one of paragraphs 3 or 4 and at least one fusion partner selected from the group consisting of ESAT-6, at least one T-cell epitope of ESAT-6, MPB64, at least one T-cell epitope of MPB64, MPT64 at least one T-cell epitope of MPT64, and MPB59 and at least one T-cell epitope of MPB59.

35. The polypeptide according to any one of paragraphs 3 or 4 which is lipidated.

36. A composition comprising a polypeptide according to any one of paragraphs 3 or 4 and pharmaceutically acceptable carrier, vehicle or adjuvant.

37. An immunological composition comprising a polypeptide according to any one of paragraphs 3 or 4.

40 38. The immunological compositions according to paragraph 37, further comprising an immunologically and pharmaceutically acceptable carrier, vehicle or adjuvant.

39. The immunological composition according to paragraph 38, wherein the carrier is a polymer to which the polypeptide(s) is/are bound by hydrophobic non-covalent interaction; the vehicle is selected from the group consisting of a diluent and a suspending agent; and the adjuvant is Freund's incomplete adjuvant.

40. An immunological composition comprising at least two different polypeptides according to paragraph 4.
41. An immunological composition comprising 3-20 different polypeptides according to paragraph 4.
- 5 42. A skin test reagent comprising the immunological composition of paragraph 37.
43. A composition for diagnosing tuberculosis in an animal, including a human being, comprising a polypeptide according to any one of paragraphs 3 or 4 optionally in combination with a means for detection.
- 10 44. A diagnostic tool comprising a combination of two or more substantially pure polypeptides, of which one or more comprises one or more amino acid sequences selected from
- (a) SEQ ID NO: 88;
  - (b) an immunogenic portion of the sequence in (a); and /or
  - 15 (c) an amino acid sequence analogue having at least 70% sequence identity to any one of the sequences in (a) or (b) and at the same time being immunogenic.
45. A serodiagnostic composition comprising a combination of two or more substantially pure polypeptides, of which one or more comprises one or more amino acid
- 20 sequences selected from
- (a) SEQ ID NO: 88;
  - (b) an immunogenic portion of the sequence in (a); and /or
  - (c) an amino acid sequence analogue having at least 70% sequence identity to any one of the sequences in (a) or (b) and at the same time being
  - 25 immunogenic.
46. A composition according to paragraph 32 or paragraph 43, which further comprises one or more amino acid sequences selected from the group consisting of:
- d) an amino acid sequence selected from the sequences shown in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66,
  - 30 68, 70, 72-86, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, and 168-171;
  - e) an immunogenic portion of any one of the sequences in (d); and
  - f) an amino acid sequence analogue having at least 70% sequence identity to any one of the sequences in (d) or (e) and at the same time being
  - 35 immunogenic.
47. A fusion protein according to paragraph 22, comprising as a fusion partner a polypeptide which comprises one or more amino acid sequences selected from the group consisting of:
- a) an amino acid sequence selected from the sequences shown in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 17-23, 42, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66,
  - 40 68, 70, 72-86, 90, 92, 94, 141, 143, 145, 147, 149, 151, 153, and 168-171;
  - b) a polypeptide fragment derived from a virulent mycobacterium, such as ESAT-6, MPB64, MPT64, TB10.4, CFP10, RD1-ORF5, RD1-ORF2, Rv1036,

Ag85A, Ag85B, Ag85C, 19kDa lipoprotein, MPT32, MPB59 and alpha-crystallin;

- c) an immunogenic portion of any one of the sequences in (a) or (b); and
- d) an amino acid sequence analogue having at least 70% sequence identity to any one of the sequences in (a), (b), or (c) and at the same time being immunogenic.

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